
**Petroleum products — Determination
of boiling range distribution — Gas
chromatography method**

*Produits pétroliers — Détermination de la répartition dans l'intervalle
de distillation — Méthode par chromatographie en phase gazeuse*



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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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For an explanation on the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the WTO principles in the Technical Barriers to Trade (TBT) see the following URL: [Foreword - Supplementary information](#)

ISO 3924 was prepared by the European Committee for Standardization (CEN) Technical Committee CEN/TC 19, *Gaseous and liquid fuels, lubricants and related products of petroleum, synthetic and biological origin*, in collaboration with ISO Technical Committee ISO/TC 28, *Petroleum products and related products of synthetic or biological origin*, in accordance with the Agreement on technical cooperation between ISO and CEN (Vienna Agreement).

This fourth edition cancels and replaces the third edition (ISO 3924:2010), which has been technically revised. The third edition had several updates regarding the calculation of ISO 3405^[1] equivalent data. Because ISO 3924 is extensively used and referenced in many fuel specifications, a faster analysis procedure was included. Many fuel specifications concerned demand volume percentage recovered at 250°C and 350°C but this result was not part of the report of ISO 3924 in the former version as described. This is updated with this edition (see [Annex A](#)), for which an assessment has been executed by CEN/TC 19. In addition, several editorial updates have been made.

This method is originally based on the jointed IP 406^[3] and ASTM D2887^[4] methods.

Petroleum products — Determination of boiling range distribution — Gas chromatography method

WARNING — — The use of this International Standard can involve hazardous materials, operations and equipment. This International Standard does not purport to address all of the safety problems associated with its use. It is the responsibility of users of this International Standard to take appropriate measures to ensure the safety and health of personnel prior to application of the standard, and fulfil statutory and regulatory requirements for this purpose.

1 Scope

This International Standard specifies a method for the determination of the boiling range distribution of petroleum products. The method is applicable to petroleum products and fractions with a final boiling point of 538 °C or lower at atmospheric pressure as determined by this International Standard. This International Standard is not applicable to gasoline samples or gasoline components. The method is limited to products having a boiling range greater than 55 °C and having a vapour pressure sufficiently low to permit sampling at ambient temperature.

The method has successfully been applied to samples containing fatty acid methyl esters (FAME) up to 10 % (V/V).

NOTE For the purposes of this International Standard, the terms “% (m/m)” and % (V/V) are used to represent the mass fraction (μ), respectively the volume fraction (φ) of a material.

2 Normative references

The following documents, in whole or in part, are normatively referenced in this document and are indispensable for its application. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 3170, *Petroleum liquids — Manual sampling*

ISO 3171, *Petroleum liquids — Automatic pipeline sampling*

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

3.1

initial boiling point

IBP

temperature corresponding to the retention time at which a net area count equal to 0,5 % of the total sample area under the chromatogram is obtained

3.2

final boiling point

FBP

temperature corresponding to the retention time at which a net area count equal to 99,5 % of the total sample area under the chromatogram is obtained

3.3

slice rate

number of data slices acquired per unit of time used to integrate the continuous (analogue) chromatographic detector response during an analysis

Note 1 to entry: The slice rate is expressed in Hz (for example, slices per second).

4 Principle

A sample is introduced into a gas chromatographic column which separates hydrocarbons in the order of increasing boiling point. The column temperature is raised at a reproducible rate and the area under the chromatogram is recorded throughout the analysis. Boiling temperatures are assigned to the time axis from a calibration curve, obtained under the same conditions by running a known mixture of hydrocarbons covering the boiling range expected in the sample. From these data, the boiling range distribution is obtained.

[Annex A](#) presents a correlation model for the calculation of physical distillation (see References [1], [5] and [6]) equivalent data from boiling range distribution analysis by gas chromatography determined following this International Standard.

[Annex B](#) describes an alternative, accelerated analysis (see [8.2](#)).

5 Reagents and materials

5.1 Stationary phase for columns, non-polar, that elutes hydrocarbons in boiling point order.

NOTE The following materials have been used successfully as liquid phases.

For packed columns:

- silicone gum rubber UC-W98;
- silicone gum rubber GE-SE-30;
- silicone gum rubber OV-1;
- silicone gum rubber OV-101;

For capillary columns:

- polydimethylsiloxane.

5.2 Solid support for packed columns, usually consisting of crushed fire brick or chromatographic diatomaceous earth.

The particle size and support loading shall be such as to give optimum resolution and analysis time.

NOTE In general, support loadings of 3 % to 10 % have been found most satisfactory.

5.3 Carrier gas, with a minimum purity of 99,995 %, constituted of

- a) helium or hydrogen for use with thermal conductivity detectors, or
- b) nitrogen, helium, hydrogen or argon for use with flame ionization detectors.

5.4 Hydrogen, grade suitable for flame ionization detectors.

5.5 Compressed air, free of oil and water, regulated for flame ionization detectors.

5.6 Calibration mixture, consisting of an accurately weighed mixture of *n*-alkanes covering the range from C₅ to C₄₄ and dissolved in carbon disulfide (5.8).

For packed columns, the final concentration should be approximately 10 parts of the alkane mixture to 100 parts of carbon disulfide. For capillary columns, the final concentration should be approximately 1 part of the alkane mixture to 100 parts of carbon disulfide.

The following mixture of alkanes has been found to be satisfactory for most samples: C₅, C₆, C₇, C₈, C₉, C₁₀, C₁₂, C₁₄, C₁₆, C₁₈, C₂₀, C₂₄, C₂₈, C₃₂, C₃₆, C₄₀, C₄₄. At least one component of the mixture shall have a boiling point lower than the initial boiling point of the sample and at least one component shall have a boiling point higher than the final boiling point of the sample. The boiling points of alkanes are listed in [Table 1](#).

Table 1 — Boiling points of normal alkanes

Carbon no.	Boiling point °C	Carbon no.	Boiling point °C
2	-89	24	391
3	-42	25	402
4	0	26	412
5	36	27	422
6	69	28	431
7	98	29	440
8	126	30	449
9	151	31	458
10	174	32	466
11	196	33	474
12	216	34	481
13	235	35	489
14	254	36	496
15	271	37	503
16	287	38	509
17	302	39	516
18	316	40	522
19	330	41	528
20	344	42	534
21	356	43	540
22	369	44	545
23	380		

NOTE API Project 44, October 31, 1972[Z], is believed to have provided the original normal paraffin boiling point data that were listed in former editions of this International Standard. However, over the years some of the data contained in both API Project 44 (Thermodynamics Research Center Hydrocarbon Project) and the test methods have changed, and they are no longer equivalent. This Table represents the current normal paraffin boiling point values accepted by ISO, ASTM and the Energy Institute.

If the test sample contains significant quantities of *n*-alkanes which can be identified on the chromatogram, these peaks may be used as internal boiling point calibration points. However, it is advisable to use the calibration mixture to be sure of peak identifications.

Propane and butane may be added non-quantitatively to the calibration mixture, if necessary, to comply with 5.6. This may be done by bubbling a small amount of the gaseous hydrocarbon into a septum-sealed vial of the calibration mixture using a gas syringe.

If stationary phases other than those listed in the note in [5.1](#) are used, the retention times of a few alkylbenzenes across the boiling range such as *o*-xylene, *n*-butylbenzene, 1,3,5-tri-isopropylbenzene, *n*-decylbenzene and *n*-tetradecylbenzene shall also be checked to make certain that the column is separating according to the boiling point order (see [Annex C](#)).

5.7 Reference material, the primary reference material used shall be the ASTM Reference Gas Oil No.1.

5.8 Carbon disulfide, reagent grade (CAS RN 75-15-0).

6 Apparatus

6.1 Chromatograph, any gas chromatograph that has the following performance characteristics may be used.

6.1.1 Detector, of either the flame ionization or thermal conductivity type.

The detector shall have sufficient sensitivity to detect a mass fraction of 1,0 % of dodecane with a peak height of at least 10 % of full scale on the recorder under the conditions specified in this International Standard, and without loss of resolution as defined in [8.3](#). When operating at this sensitivity level, detector stability shall be such that a baseline drift of not more than 1 % of full scale per hour is obtained. The detector shall be capable of operating continuously at a temperature equivalent to the maximum column temperature employed. The detector shall be connected to the column in such a way that any cold spots between the detector and the column are avoided.

NOTE It is not desirable to operate thermal conductivity detectors at a temperature higher than the maximum column temperature employed. Operation at higher temperatures only serves to shorten the useful life of the detector, and generally contributes to higher noise levels and greater drift.

6.1.2 Column temperature programmer, capable of programmed temperature operation over a range sufficient to establish a retention time of at least 1 min for the initial boiling point and to elute the entire sample within the temperature ramp.

The programming rate shall be sufficiently reproducible to obtain retention time repeatability of 6 s for each component in the calibration mixture ([5.6](#)).

If the initial boiling point is less than approximately 93 °C, an initial column temperature below ambient can be required. However, excessively low initial column temperatures shall be avoided, to ensure that the stationary phase remains liquid. The initial temperature of the column shall be only low enough to obtain a calibration curve meeting the requirements of this International Standard.

6.1.3 Sample inlet system, either be capable of operating continuously at a temperature equivalent to the maximum column temperature employed or provide on-column injection with some means of programming the entire column, including the point of sample introduction, up to the maximum temperature required.

The sample inlet system shall be connected to the chromatographic column in such a way that any cold spots between the inlet system and the column are avoided.

6.2 Column, any column and conditions may be used, provided that, under the conditions of the test, separations are in the order of boiling points as given in [Table 1](#), and the column resolution, *CR*, is at least 3 ([8.3](#)). Typical column operating conditions are given in [Table 2](#) and [3](#).

Table 2 — Typical operating conditions for packed columns

Packed columns	1	2
Column length, (m)	0,7	0,5
Column outside diameter, (mm)	3,2	3,2
Stationary phase	OV-101	UC-W98
Percent stationary phase	5	10
Support material	G ^a	P ^b
Support mesh size (µm)	80/100	80/100
Initial column temperature, (°C)	-40	-30
Final column temperature, (°C)	350	360
Programming rate, (°C/min)	10	10
Carrier gas	Helium	Nitrogen
Carrier gas flow, (ml/min)	30	25
Detector	FID	FID
Detector temperature, (°C)	370	360
Injection-port temperature, (°C)	370	350
Sample size, (µl)	0,5	1
^a Chromosorb® G (AW-DMS).		
^b Chromosorb® P (AW).		

Table 3 — Typical operating conditions for capillary columns

Capillary columns	3	4	5
Column length (m)	7,5	5	10
Column inner diameter (mm)	0,53	0,53	0,53
Column	DB-1	HP-1	HP-1
Stationary phase thickness (µm)	1,5	0,88	2,65
Carrier gas	Nitrogen	Helium	Helium
Carrier gas flow rate (ml/min)	30	12	20
Initial column temperature (°C)	40	35	40
Final column temperature (°C)	340	350	350
Programming rate (°C/min)	10	10	15
Detector	FID	FID	FID
Detector temperature (°C)	350	380	350
Injector temperature (°C)	340	Cool on-column type	Programmed temperature vaporization type
Sample size (µl)	0,5	1	0,2
Sample concentration [% (m/m)]	25	10	Neat

6.3 Recorder/plotter, this apparatus is used for plotting the chromatogram. This may be accomplished using a 0 mV to 1 mV recording potentiometer having a full-scale response time of 2 s or less and a minimum chart width of approximately 120 mm. Alternatively, a computer or other device may be used, provided it is capable of graphics presentation of the same or better quality as a potentiometric recorder.

6.4 Integrator/computer, this apparatus is used for determining the accumulated area under the chromatogram. This may be achieved by using a computer-based chromatography data system or an electronic integrator. The integrator/computer system shall have normal chromatographic software for measuring the retention times and areas of eluting peaks. In addition, the system shall be capable of

converting the continuously integrated detector signal into area slices of fixed duration. These contiguous area slices, collected for the entire analysis, shall be stored for later processing. The electronic range of the integrator/computer (e.g. 1 V) shall be within the linear range of the detector/electrometer system used. The system shall be capable of subtracting the area slice of a blank run from the corresponding area slice of a sample run.

NOTE Some gas chromatographs have an algorithm built into their operating software that allows a mathematical model of the baseline profile to be stored in the memory. This profile can be automatically subtracted from the detector signal on subsequent sample analysis to compensate for any baseline offset. Some integration systems also store and automatically subtract a blank analysis from subsequent sample analysis.

6.5 Flow/pressure controllers.

6.5.1 If a packed column is used, the chromatograph shall be equipped with constant-flow controllers capable of maintaining the carrier gas flow constant to ± 1 % over the full operating temperature range.

6.5.2 If a wide-bore capillary column is used, the chromatograph shall be equipped with a controller of carrier gas flow or pressure appropriate for the inlet used.

6.6 Micro-syringe, this apparatus is used to introduce the sample into the chromatograph. Sample injection may be either manual or automatic. Automatic sample injection is preferred because it gives better retention time precision.

7 Sampling

Unless otherwise specified, samples shall be taken by the procedures described in ISO 3170 or ISO 3171.

8 Preparation of apparatus

8.1 Column preparation, any satisfactory method that will produce a column meeting the requirements of 6.2 may be used. The column shall be conditioned at the maximum operating temperature to reduce baseline shifts due to bleeding of the column substrate.

8.1.1 Packed columns, an acceptable method of column conditioning, which has been found effective for columns with an initial loading of 10 % liquid phase, consists of purging the column with carrier gas at the normal flow rate while holding the column at the maximum operating temperature for 12 h to 16 h.

8.1.2 Capillary columns, capillary columns may be conditioned using the following procedure.

- a) Install the column following the manufacturer's instructions. Set the column and detector gas flows. Ensure that the system is leak free.
- b) Allow the system to purge with carrier gas at ambient temperature for at least 30 min. Then increase the oven temperature by approximately 5 °C/min to 10 °C/min to the final operating temperature and hold for approximately 30 min.
- c) Cycle the chromatograph through its temperature programme several times until a stable baseline is obtained.

NOTE 1 Capillary columns with cross-linked and bonded phases are available from many manufacturers and are usually preconditioned. These columns have much lower column bleed than packed columns.

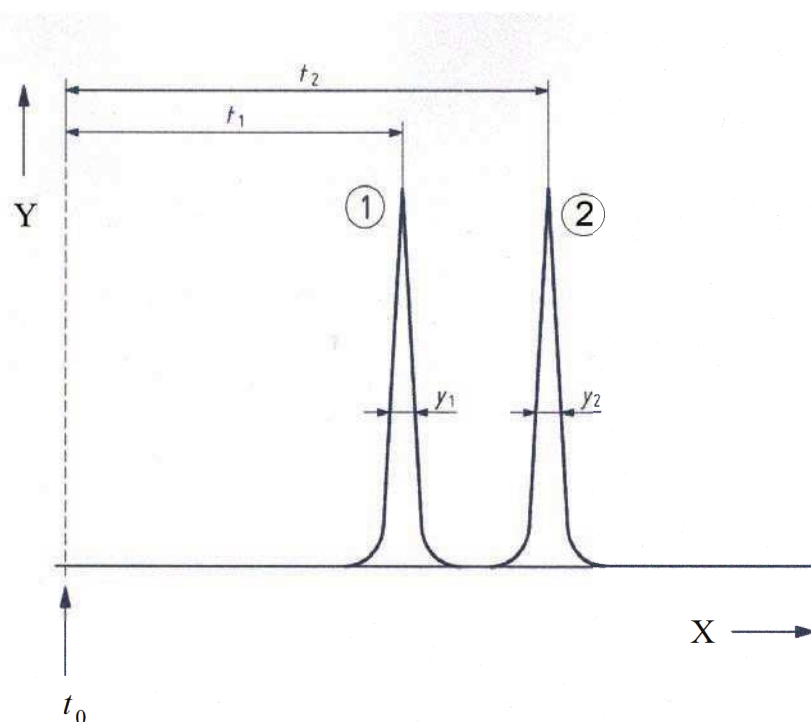
NOTE 2 The column is not always connected to the FID when making a first conditioning of the column to overcome that initial column bleed affects the detector's sensitivity.

8.2 Chromatograph, place the chromatograph in service in accordance with the manufacturer's instructions. Typical operating conditions are shown in [Tables 2](#) and [3](#).

If a flame ionization detector is used, the deposits formed in the detector from combustion of the silicone decomposition products shall be removed regularly, as they change the response characteristics of the detector.

NOTE Without any instrumental adaptation, it is possible to decrease analysis time. [Annex B](#) describes such an accelerated analysis.

8.3 Column resolution, analyse the calibration mixture under the same conditions as those used for the samples. Using the procedure illustrated in [Figure 1](#), calculate the resolution, CR , from the time between the hexadecane and octadecane peaks at the peak maxima t_1 and t_2 and the widths y_1 and y_2 of the peaks at half height, as given by [Formula \(1\)](#).



Key

X	time (s)	y_1	width of hexadecane peak at half height, in s
Y	detector signal	y_2	width of octadecane peak at half height, in s
t_0	start analysis time	1	hexadecane
t_1	retention time hexadecane, in s	2	octadecane
t_2	retention time octadecane, in s		

Figure 1 — Column resolution parameters

$$CR = \frac{2(t_2 - t_1)}{1,699(y_1 + y_2)} \quad (1)$$

where

t_1 is the retention time, in seconds, for hexadecane peak maximum;

t_2 is the retention time, in seconds, for octadecane peak maximum;

y_1 is the width, in seconds, at half height of hexadecane peak;

y_2 is the width, in seconds, at half height of octadecane peak.

The resolution, CR , obtained from the [Formula \(1\)](#), shall be at least three.

8.4 Detector response check, this method assumes that the detector response to petroleum hydrocarbons is proportional to the mass of individual components. This shall be verified when the system is put into service and whenever any changes are made to the system or operational parameters. Analyse the calibration mixture ([5.6](#)) using the same conditions as those used for the samples. Calculate the response factor, F_n , for each alkane relative to decane using [Formula \(2\)](#):

$$F_n = \frac{m_n / A_n}{m_{10} / A_{10}} \quad (2)$$

where

F_n is the relative response factor;

m_n is the mass of the alkane in the mixture;

A_n is the peak area of the alkane;

m_{10} is the mass of decane in the mixture;

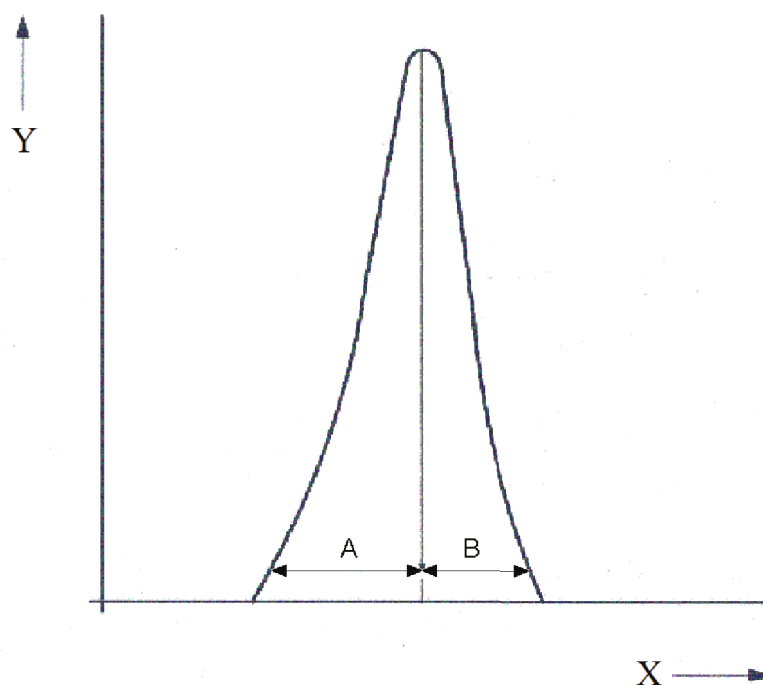
A_{10} is the peak area of decane.

The relative response factor, F_n , of each alkane shall not deviate from 1,0 by more than $\pm 0,1$.

8.5 Peak skewness, determine the peak skewness (the ratio A/B) of the largest peak in the calibration mixture ([5.6](#)) as shown in [Figure 2](#).

The peak skewness shall be not less than 0,5 and not more than 2,0. If peak skewness is outside these parameters, reanalyse the calibration mixture using a smaller sample size or a more dilute solution, if necessary, to avoid peak distortion.

NOTE Skewness is often an indication of overloading the column that results in displacement of the peak apex relative to non-overloaded peaks. Distortion in retention time measurement and hence errors in boiling point determination will be likely if column overloading occurs. The column liquid phase loading has a direct bearing on the acceptable sample size.



Key

X time (s)

Y detector signal

A width of the leading part of the peak at 5 % of peak height, in s

B width of the trailing part of the peak at 5 % of peak height, in s

Figure 2 — Peak skewness

9 Calibration

9.1 Analysis sequence protocol

9.1.1 Define and use for all runs a predetermined schedule of analysis events to achieve maximum reproducibility. The schedule shall include cooling the oven to the initial starting temperature, equilibration time, sample injection and system start; analysis and final temperature hold time.

9.1.2 After the chromatographic conditions have been set to meet performance requirements, programme the column temperature upward to the maximum temperature to be used and hold that temperature for the selected time. Following the analysis sequence protocol, cool the column to the initial starting temperature.

9.1.3 During the cool down and equilibration time, prepare the integrator/computer system for data acquisition. If a retention time or detector response calibration is being performed, use the peak detection mode. For samples and baseline compensation determinations, use the area slice mode of integration. The recommended slice rate for this method is 1 Hz (one slice per second).

9.1.4 At the exact time set by the schedule, inject either the calibration mixture (5.6) or sample into the chromatograph; or make no injection (baseline blank). At the time of injection and/or at the start of the

baseline blank, start the chromatograph time cycle and the integrator/computer data acquisition. Follow this analysis sequence protocol for all subsequent analysis, blanks or calibrations.

9.2 Baseline compensation analysis

9.2.1 A baseline compensation analysis, or baseline blank, shall be performed at least once each day that the test is run, using the same technique for a sample analysis except that no injection is made.

NOTE The blank analysis is necessary due to the normal occurrence of chromatographic baseline rise near the maximum column temperature. Factors that influence baseline stability are column bleed, septum bleed, detector temperature control, constancy of carrier and detector gas flows, leaks, instrument drift, etc.

9.2.2 Subtract the blank analysis from the sample analysis to remove any non-sample slice area from the chromatographic data.

The blank analysis is typically performed prior to sample analysis, but can be useful if determined between samples or at the end of a sample sequence to provide additional data regarding instrument operation or residual sample carry-over from previous sample analysis.

9.2.3 Carry out periodic baseline blank analysis in accordance with the analysis sequence protocol to give an indication of baseline stability.

9.3 Retention time versus boiling point calibration

9.3.1 A retention time versus boiling point calibration shall be performed at least once each day that the test is run. Inject an appropriate aliquot (0,2 µl to 2,0 µl) of the calibration mixture (5.6) into the chromatograph following the analysis sequence protocol.

9.3.2 Prepare a calibration table based on the results of the analysis of the calibration mixture (5.6) by recording the retention time and the boiling temperature for each component in the mixture. Boiling temperatures of alkanes are listed in Table 1.

9.3.3 Plot the retention time of each peak versus the corresponding boiling temperature for that component. A typical calibration curve is shown in Figure 3.

9.3.4 Ensure that calibration points bracket the boiling range of the sample at both the low and high ends. Ideally, the calibration plot of retention time versus boiling temperature should be linear, but it is impractical to operate the chromatograph such that curvature is eliminated completely.

NOTE The greatest potential for deviation from linearity is associated with the lower boiling point alkanes, which elute from the column relatively quickly and have the largest difference in boiling temperatures. In general, the lower the sample initial boiling point, the lower the starting point of the analysis will be.

9.4 Analysis of reference material

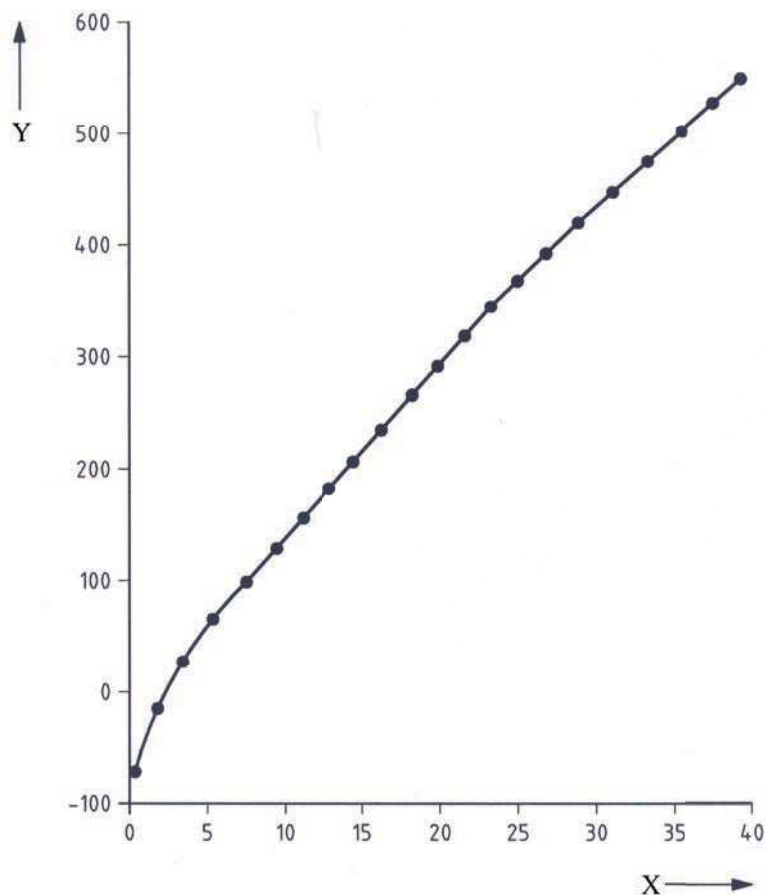
9.4.1 The reference material (5.7) is used to verify both the chromatographic and calculation processes involved in this method.

A secondary reference material may be used, providing it satisfies the following criteria:

- a) it is similar in nature and boiling range to the samples to be analysed;
- b) the boiling range distribution values assigned to that obtained by averaging multiple analysis of the secondary reference material on a system that is first shown to be operating properly with the primary reference material (5.7).

9.4.2 Analyse the primary reference material (5.7) or a secondary reference material at least once each day that the test is run. Perform an analysis of the reference material following the analysis sequence protocol (see 9.1). Collect the area slice data and provide a boiling point distribution report in accordance with 12.1.

9.4.3 The results of the analysis of the reference material (either batch 1 or batch 2 can be used) shall not deviate more from the values for that batch given in Table 4 than the range specified by the reproducibility of this International Standard (see 13.3).



Key

X retention time (min)

Y boiling point (°C)

Figure 3 — Typical calibration curve

Table 4 — Specified temperature-recovery values for ASTM Gas Oil No. 1

Percent recovered %	Batch No. 1	Batch No. 2
	Temperature °C	Temperature °C
IBP	114	115
5	143	151
10	169	176
15	196	201
20	221	224
30	258	259
40	287	289
50	312	312
60	332	332
70	354	354
80	376	378
90	404	407
95	425	428
FBP	475	475

10 Procedure

10.1 Sample preparation

10.1.1 The amount of sample injected shall not overload the column stationary phase capacity nor exceed the detector linear range.

NOTE A narrow boiling range sample will require the injection of a smaller amount than a wider boiling range sample.

10.1.2 The column stationary phase capacity can be estimated from the chromatogram of the calibration mixture (5.6). Different volumes of the calibration mixture (5.6) can be injected to find the maximum amount of a component that the stationary phase can tolerate without overloading (see 8.5, Note). Note the peak height for this amount of sample. The maximum sample signal intensity shall not exceed this peak height.

10.1.3 Samples that are of low enough viscosity to be sampled with a syringe at ambient temperature shall be injected undiluted. Samples that are too viscous or waxy to be sampled with a syringe may be diluted with carbon disulfide (5.8).

10.1.4 Typical sample injection volumes are shown in Tables 5 and 6.

10.2 Sample analysis

Using the analysis sequence protocol (see 9.1), inject a sample aliquot into the gas chromatograph. At the time of injection, start the chromatograph time cycle and the integrator/computer data acquisition.

Table 5 — Typical sample injection volumes for packed columns

Stationary phase loading %	Neat sample volume μl
10	1,0
5	0,5

Table 6 — Typical sample injection volumes for capillary columns

Film thickness μm	Neat sample volume μl
0,8 to 1,5	0,1 to 0,2
1,8 to 3,0	0,1 to 0,5
3,0 to 5,0	0,2 to 1,0

11 Calculation

11.1 Correct the sample area slices for non-sample detector response by subtracting each blank analysis area slice from each sample area slice at the equivalent slice time. Sum the corrected area slices to obtain the cumulative corrected areas for each time interval during the run.

11.2 At the point on the chromatogram where the baseline at the end of the run first becomes steady, record the total cumulative area counts. Move back along the chromatogram until the cumulative area equals 99,5 % of the total area. Mark this point as the final boiling point (FBP).

NOTE Location of the final boiling point can be the most difficult step in this method. Some samples have extremely long tail-end portions due to gradually decreasing amounts of heavy material. This fact, coupled with the natural tendency of the chromatographic baseline to rise at the end of the run due to septum or column bleed or elution of traces of heavy components from previous samples, can preclude the possibility of the chromatogram returning precisely to the original baseline established prior to the initial boiling point of the sample. Thus, the most satisfactory procedure is to inspect the chromatogram and the area counts at each interval near the end of the run to determine the point at which the rate of change of the chromatographic signal has reached a constant low value of no greater than 0,000 01 % of the total area counts per second.

11.3 Observe the area counts at the start of the run until the point is reached where the cumulative area count is equal to 0,5 % of the total area. Mark this point as the initial boiling point (IBP) of the sample. If carbon disulfide is used as the solvent, its response shall be ignored in the calculations.

11.4 Divide the cumulative area at each interval between the initial and final boiling points by the total area and multiply by 100 to give the percentage of the sample recovered at each time interval.

11.5 Tabulate the cumulative percentage recovered at each interval and the retention time at the end of the interval. Using linear interpolation where necessary, determine the retention time associated with each percentage between 1 % and 99 %.

11.6 For each percentage and its associated retention time, determine the corresponding boiling temperature from the calibration table (see [9.3.2](#)). Use linear interpolation between data points.

12 Expression of results

12.1 Report the temperature to the nearest 0,5 °C at 1 % intervals between 1 % and 99 % and at the IBP and the FBP.

12.2 If a plot of the boiling point distribution curve is required, use graph paper with uniform subdivisions and plot each boiling temperature against its corresponding percentage recovered. Plot the initial boiling point at 0 % and the final boiling point at 100 % recovered. Draw a smooth curve connecting the points.

13 Precision

13.1 General

The precision, as determined by statistical examination in accordance with ISO 4259[2] of interlaboratory test results, is given in 13.2 and 13.3.

13.2 Repeatability

The difference between two test results, obtained by the same operator with the same apparatus under constant operating conditions on identical test material would, in the long run, in the normal and correct operation of the test method, exceed the values given in Table 7 in only one case in 20.

Table 7 — Repeatability values

Percent recovered	Repeatability °C
IBP	0,011 <i>X</i>
5 %	0,003 2 (<i>X</i> + 100)
10 % to 40 %	0,8
50 % to 90 %	1,0
95 %	1,2
FBP	3,2
NOTE <i>X</i> is the average of the two results, in °C.	

13.3 Reproducibility

The difference between two single and independent test results, obtained by different operators working in different laboratories on identical test material would, in the long run, in the normal and correct operation of the test method, exceed the values given in Table 8 in only one case in 20.

Table 8 — Reproducibility values

Percent recovered	Reproducibility °C
IBP	0,066 <i>X</i>
5 % to 20 %	0,015 (<i>X</i> + 100)
30 %	0,013 (<i>X</i> + 100)
40 % to 90 %	4,3
95 %	5,0
FBP	11,8
NOTE <i>X</i> is the average of the two results, in °C.	

14 Test report

The test report shall contain at least the following information:

- a) reference to this International Standard, i.e. ISO 3924:2016;
- b) type and complete identification of the product tested;
- c) result of the test (see [Clause 12](#));
- d) any deviation, by agreement or otherwise, from the procedure specified;
- e) date of the test.

Annex A (informative)

Calculation of ISO 3405 equivalent data

A.1 General

A correlation model is presented for the calculation of ISO 3405^[1] equivalent data from boiling range distribution analysis by gas chromatography following the main part of this International Standard.

The correlation model is only valid for diesel and jet fuels and should obey the sample specification given in [Clause 1](#).

The correlation model is validated by an Analysis of Variance procedure according to ASTM D6708.^[8]

Valid data for conversion to ISO 3405 equivalent data can be obtained by the use of this Annex.

[A.4](#) describes the calculation of percent volume recoveries at temperature cutpoint intervals from the data obtained through this correlation model.

A.2 Procedure

ISO 3405 equivalent data are calculated from this International Standard's data using [Formula \(A.1\)](#) and coefficients specified in [Table A.1](#).

$$t_n = a_0 + a_1 \times T_{n-1} + a_2 \times T_n + a_3 \times T_{n+1} \quad (\text{A.1})$$

where

t_n n^{th} boiling temperature of ISO 3405 equivalent;

a_i i^{th} coefficient from [Table A.1](#);

T_n n^{th} boiling temperature as calculated and reported in [Clause 12](#).

A.3 Justification

The correlation model is based on data from 46 jet fuel samples and 39 diesel samples analysed using methods according to ISO 3405 and this International Standard. From these results, a correlation model is determined using regression, specifying coefficients per recovery. A model of the remaining bias is determined by use of the procedure as described in ASTM D6708, on a data set from the ASTM interlaboratory crosscheck program containing five jet fuels and six diesels analysed by 38 laboratories using the method as described in this International Standard and 201 laboratories using ISO 3405.

The bias correction model has been used to correct the results from the correlation model, resulting in a new correlation matrix given in [Table A.1](#).

Both methods are found sufficiently precise to distinguish among the samples.

Table A.1 — Correlation coefficients

t_n	a_0	a_1	a_2	a_3	T_n		
IBP	25,351	0,322 16	0,711 87	-0,042 21	T_{IBP}	T_5	T_{10}
5 %	18,822	0,066 02	0,158 03	0,778 98	T_{IBP}	T_5	T_{10}
10 %	15,173	0,201 49	0,306 06	0,482 27	T_5	T_{10}	T_{20}
20 %	13,141	0,226 77	0,290 42	0,460 23	T_{10}	T_{20}	T_{30}
30 %	5,776 6	0,372 18	0,303 13	0,311 18	T_{20}	T_{30}	T_{50}
50 %	6,375 3	0,077 63	0,689 84	0,183 02	T_{30}	T_{50}	T_{70}
70 %	-2,843 7	0,163 66	0,421 02	0,382 52	T_{50}	T_{70}	T_{80}
80 %	-0,215 36	0,256 14	0,409 25	0,279 95	T_{70}	T_{80}	T_{90}
90 %	0,099 66	0,243 35	0,320 51	0,373 57	T_{80}	T_{90}	T_{95}
95 %	0,898 80	-0,097 90	1,038 16	-0,008 94	T_{90}	T_{95}	T_{FBP}
FBP	19,444	-0,381 61	1,085 71	0,177 29	T_{90}	T_{95}	T_{FBP}

A.4 Calculating volume percent recoveries at temperature cutpoint intervals

The % (V/V) recovery (x) at a certain temperature cutpoint is obtained through linear interpolation between two known recoveries by using [Formula \(A.2\)](#):

$$x = x_1 + (y - y_1) \frac{(x_2 - x_1)}{(y_2 - y_1)} \quad (\text{A.2})$$

where

- y is the required temperature cutpoint;
- x_1 is known recovery at the temperature below y ;
- x_2 is known recovery at the temperature above y ;
- y_1 is temperature cutpoint belonging to x_1 ;
- y_2 is temperature cutpoint belonging to x_2 .

A typical example is given in [Tables A.2](#) and [A.3](#).

Table A.2 — Example data of temperature versus percent volume recovery

Volume recovered % (V/V)	Temperature °C
0,5	199,9
5,0	215,6
10,0	228,2
20,0	246,8
30,0	261,3
50,0	280,5
70,0	305,3
80,0	318,1
90,0	335,4
95,0	348,7
99,5	365,4

Table A.3 — Calculated recoveries in percent volume from data in [Table A.2](#)

Cutpoint °C	Volume recovered % (V/V)
250	22,2
350	95,4

A.5 Precision and bias

The reproducibility of the converted chromatographic data into ISO 3405 equivalent data are in accordance with the reproducibility of the gas chromatographic data described in [13.3](#)

Cross-method reproducibility after conversion of chromatographic data into ISO 3405 equivalent data are specified in [Table A.4](#).

Table A.4 — Cross-method reproducibility

t_n	IBP	5 %	10 %	20 %	30 %	50 %	70 %	80 %	90 %	95 %	FBP
R	13,71	11,80	10,73	8,83	7,39	6,96	7,03	7,62	8,85	17,32	12,94
NOTE R is the reproducibility in °C.											

The reproducibility of the calculated recoveries in % (V/V) at 250 °C and 350 °C can be estimated from [Table 8](#) by linear interpolation between the nearest values below and above the calculated recovery.

EXAMPLE Reproducibility (R) calculation using the results from [Table A.3](#):

R at 20 % is 5,2 °C and R at 30 % is 4,7 °C → R at 22,2 % (V/V) = 5,1 °C

R at 95 % is 5,0 °C and R at 99,5 % is 11,8 °C → R at 95,4 % (V/V) = 5,6 °C

Annex B (informative)

Accelerated analysis

B.1 General

Because the test method is extensively used for all kinds of products, there is a need for an accelerated method in order to save analysis time. Without any instrumental adaptation it is possible to decrease analysis time by a factor five. This Annex describes a set up to reduce the original analysis time of 40 min to less than 10 min. Such methods are usually referred to as accelerated analysis.

Simulated distillation methods have been reported where analysis times were less than two minutes. These methods are referred to as fast analysis and are not described in this Annex.

A research report with supporting data are available (see Reference [9]).

B.2 Procedure

B.2.1 Column dimensions as needed for an accelerated procedure fall within the ones described in [Table 3](#), except for the programming rate, which is set at a typical value of 35 °C/min. [Table B.1](#) gives the typical operating conditions.

Table B.1 — Typical operating conditions for accelerated analysis

Column length (m)	10
Column inner diameter (mm)	0,53
Column	HP-1
Stationary phase thickness (µm)	0,88
Carrier gas	helium
Carrier gas flow rate (ml/min)	26
Initial column temperature (°C)	40
Final column temperature (°C)	360
Programming rate (°C/min)	35
Detector	FID
Detector temperature (°C)	360
Injector temperature initial (°C)	100
Injector programming rate (°C/min)	35
Injector temperature final (°C)	360
Sample size (µl)	0,1
Sample concentration	neat

B.2.2 Slice rate as given in [9.1.3](#) should be adjusted so that the total amount of data points stays around 1 500.

B.2.3 The provision as defined in [6.2](#) will not be met, e.g. the retention time for the IBP will be smaller than 1 min. A negative impact could not be found.

B.3 Justification

A comparison between the test method as defined in the main body of this International Standard (standard procedure) and an accelerated procedure has been made based on 40 and 26 instruments, respectively.

B.4 Precision and bias

Repeatability is according to [Table 7](#) (see Reference [9]).

Reproducibility is according to [Table 8](#) (see Reference [9]).

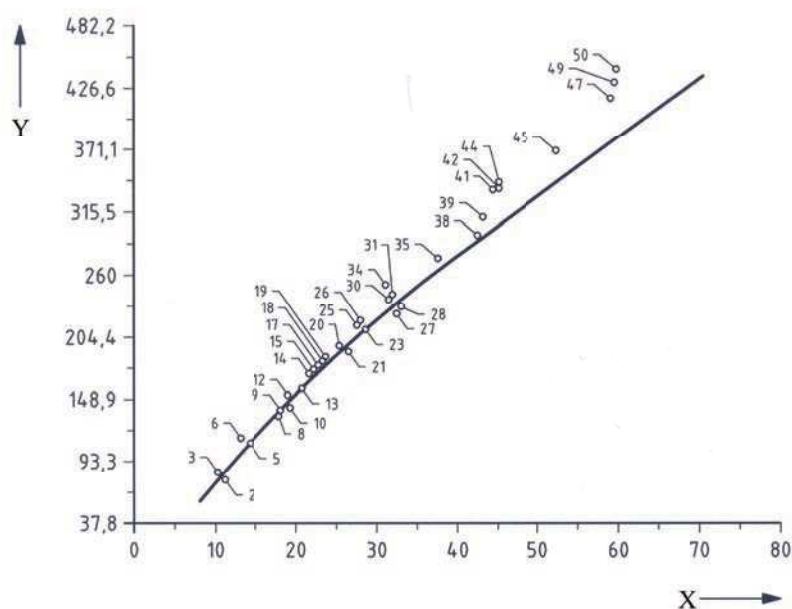
No significant bias between the standard procedure and the accelerated procedure could be found in comparison studies.^[9] If an accelerated procedure is implemented, a primary reference material ([5.7](#)), should be analysed, so that the bias will be verified.

Annex C (informative)

Boiling points of non-normal alkane hydrocarbons

C.1 There is an apparent discrepancy in the boiling point versus retention time of certain high-boiling multiple-ring-type compounds. When the retention times of these compounds are compared with those of *n*-alkanes of equivalent atmospheric boiling point, these ring compounds appear to be eluted early from silicone rubber columns. A graph showing 36 compounds other than *n*-alkanes plotted along the calibration curve for *n*-alkanes alone is shown in [Figure C.1](#). The numbered dots are identified in [Table C.1](#). In [Figure C.1](#), the atmospheric boiling points are plotted against the observed retention times.

If columns containing different percentages of stationary phase or different temperature-programming rates were used, the slope and curvature of the *n*-alkane curve (solid line) remains essentially the same. Deviations of distillation boiling points, as estimated from the curve, from true boiling points for a few compounds are shown in [Table C.2](#). The deviations obtained by plotting boiling points at 1,333 kPa rather than 101,325 kPa are also tabulated. It is apparent that the deviation is much less at 1,333 kPa pressure. This indicates that the distillation data produced by gas chromatography closely approximate those obtained in reduced pressure distillation. Since the vapour pressure versus temperature curves for multiple-ring-type compounds do not have the same slope or curvature as those for *n*-alkanes, an apparent discrepancy would exist when *n*-alkane boiling points at atmospheric pressure were used.



Key

- X retention time (min)
- Y boiling point (°C)

Figure C.1 — Boiling point-retention time relationship for several multiple-ring-type compounds with high boiling points

Table C.1 — Compound identification corresponding to numbered dots from [Figure C.1](#)

No.	Boiling point °C	Compound
2	80	Benzene
3	84	Thiophene
5	111	Toluene
6	116	Pyridine
8	136	2,5-Dimethylthiophene
9	139	1,4-Xylene
10	143	Dipropylsulfide
12	152	Cumene
13	159	<i>trans</i> -Hexahydroindane
14	171	Dec-1-ene
15	173	<i>sec</i> -Butylbenzene
17	178	2,3-Dihydroindene
18	183	Butylbenzene
19	186	<i>trans</i> -Decalin
20	194	<i>cis</i> -Decalin
21	195	Dipropyldisulfide
23	213	Dodec-1-ene
25	218	Naphthalene
26	221	2,3-Benzothiophene
27	227	Diamylsulfide
28	234	1,3,5-Tri-isopropylbenzene
30	241	2-Methylnaphthalene
31	245	1-Methylnaphthalene
34	254	Indole
35	279	Acenaphthene
38	298	Decylbenzene
39	314	Octadec-1-ene
41	339	Phenanthrene
42	342	Anthracene
44	346	Acridine
45	395	Pyrene
47	424	Triphenylene
49	438	Naphthacene
50	447	Chrysene

Table C.2 — Deviations from true boiling points of boiling points obtained from this International Standard

Compound	True boiling point, TBP °C at 101,325 kPa	Deviations from true boiling point	
		°C at 101,325 kPa	°C at 1,333 kPa
Benzene	80	+4	-2
Thiophene	86	+4	+1
Toluene	111	+2	-1
^a No data exists at 1,333 kPa for chrysene.			

Table C.2 (continued)

Compound	True boiling point, TBP °C at 101,325 kPa	Deviations from true boiling point	
		°C at 101,325 kPa	°C at 1,333 kPa
1,4-Xylene	139	0	+2
Dodec-1-ene	213	0	0
Naphthalene	218	-12	-4
2,3-Benzothiophene	221	-13	0
2-Methylnaphthalene	241	-12	-2
1- Methylnaphthalene	245	-12	-5
Dibenzothiophene	332	-32	-6
Phenanthrene	339	-35	-8
Anthracene	342	-36	-8
Pyrene	395	-48	-16
Chrysene	447	-60	a

^a No data exists at 1,333 kPa for chrysene.

C.2 However, this discrepancy does not introduce any significant error when compared with laboratory distillation, because the pressure is reduced in such procedures when overhead temperatures reach approximately 260 °C, to prevent cracking of the sample. Thus, distillation data are subject to the same deviations experienced in distillation by gas chromatography. A comparison of data obtained from true boiling point (TBP) distillations with those obtained from simulated distillation by gas chromatography of three high-boiling petroleum fractions is shown in [Table C.3](#). The TBP distillations were made on 100 theoretical plate-spinning band columns at 0,133 kPa.

C.3 The decanted oil is of particular interest because it contains a high percentage of polycyclic aromatic compounds and the high sulfur coker gas oil should contain ring-type sulfur compounds and complex olefinic types.

Table C.3 — Distillation of heavy gas oils

% (m/m)	Virgin gas oil		High sulfur coker gas oil		"Decanted" oil	
	TBP ^a °C	GC ^b °C	TBP ^a °C	GC ^b °C	TBP ^a °C	GC ^b °C
IB	230	215	223	209	190	176
10	269	263	274	259	318	302
20	304	294	296	284	341	338
30	328	321	316	312	357	358
40	343	348	336	344	377	375
50	367	373	356	364	390	391
60	394	398	377	386	410	409
70	417	424	399	410	425	425
80	447	451	427	434	445	449
90	—	448	462	467	—	469
95	—	511	482	494	—	492
100	—	541	—	—	—	541

^a TBP, true boiling point.

^b GC, boiling point determined in accordance with this International Standard (gas chromatography).

Bibliography

- [1] ISO 3405, *Petroleum products — Determination of distillation characteristics at atmospheric pressure*
- [2] ISO 4259, *Petroleum products — Determination and application of precision data in relation to methods of test*
- [3] IP 406/14, *Petroleum products — Determination of boiling range distribution by gas chromatography*
- [4] ASTM D2887-13, *Standard Test Method for Boiling Range Distribution of Petroleum Fractions by Gas Chromatography*
- [5] IP 123, *Petroleum products — Determination of distillation characteristics at atmospheric pressure*
- [6] ASTM D86, *Standard Test Method for Distillation of Petroleum Products at Atmospheric Pressure*
- [7] API Project 44, Report, 31 October, 1972
- [8] ASTM D6708-13, *Standard Practice for Statistical Assessment and Improvement of Expected Agreement Between Two Test Methods that Purport to Measure the Same Property of a Material*
- [9] EI Research Report IP 406/05, *Determination of boiling range distribution of distillates and lubricating oils — Gas chromatography method — Precision Evaluation in IP 406*, available from the Energy Institute, 61 New Cavendish Street, London W1G 7AR, England

