

BS EN 16695:2015



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Water quality — Guidance on the estimation of phytoplankton biovolume

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National foreword

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The UK participation in its preparation was entrusted to Technical Committee EH/3/5, Biological Methods.

A list of organizations represented on this committee can be obtained on request to its secretary.

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ISBN 978 0 580 84590 1

ICS 13.060.70

Compliance with a British Standard cannot confer immunity from legal obligations.

This British Standard was published under the authority of the Standards Policy and Strategy Committee on 30 September 2015.

Amendments issued since publication

Date	Text affected
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EUROPEAN STANDARD

EN 16695

NORME EUROPÉENNE

EUROPÄISCHE NORM

September 2015

ICS 13.060.70

English Version

Water quality - Guidance on the estimation of phytoplankton biovolume

Qualité de l'eau - Lignes directrices pour l'estimation
du biovolume des microalgues

Wasserbeschaffenheit - Anleitung zur Abschätzung des
Phytoplankton-Biovolumens

This European Standard was approved by CEN on 10 July 2015.

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European foreword

This document (EN 16695:2015) has been prepared by Technical Committee CEN/TC 230 “Water analysis”, the secretariat of which is held by DIN.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by March 2016 and conflicting national standards shall be withdrawn at the latest by March 2016.

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Introduction

The abundance or number of counting units of individual phytoplankton taxa does not necessarily reflect the real ratio of single taxa to the complete biomass of a phytoplankton community. Few big cells/counting units can contribute far more biomass to the system than many small ones. Hence, abundance data alone is often not an ideal measurement of population size. Biomass estimations give very important information for ecological studies, classification schemes and ecosystem modelling. Therefore, it is necessary to determine the biomass of phytoplankton taxa, particularly because phytoplankton delivers energy in the form of carbon, to other trophic levels of food webs. It is not possible to directly analyse the carbon content on the taxonomic level in natural phytoplankton samples, therefore the biovolume of the phytoplankton taxa is a suitable measure to determine the biomass of an ecosystem according to the taxonomic composition. Neither particle size analysis using laser analysis, nor flow cytometry, nor Coulter Counters, nor chemical analyses of chlorophyll-a concentration as well as total carbon allow statements on the taxon level. An estimation of the carbon content is possible using conversion factors (see Annex C).

Further, the biovolume is a quantitative basis for assessing hazards from those algae and cyanobacteria, which (can) contain noxious or toxic metabolites, and is used in combination with cell numbers or chlorophyll-a concentration within WHO guidelines and national regulations for risk assessments.

Up to now, various guidelines for estimating the biovolume of microalgae have been used in different national and international monitoring programs (e.g. [1], [2], [3], [4]). The main objective of this document is the standardization of the procedure for determining the phytoplankton biovolume in order to achieve comparability of data. For this reason, the estimation of the biovolume in phytoplankton samples in sedimentation chambers (according to Utermöhl) using an inverted microscope will be described in detail.

This European Standard is also applicable for image analysis of pictures derived from microscope and flow cytometry cameras. The use of a standard catalogue containing basic and some composed geometrical shapes is recommended. Of course, such a standard list will not reflect the variety of all naturally existing shapes and will not match the exact biovolume values of each taxon. It will always be a compromise between accuracy and efficiency. However, the usage of agreed geometrical shapes and the application of the relevant formulae will improve the comparability of phytoplankton data and will be an important step forward for the implementation of quality assurance measures in phytoplankton analysis.

1 Scope

This European Standard specifies a procedure for the estimation of biovolume of marine and freshwater phytoplankton taxa using inverted microscopy (Utermöhl technique according to EN 15204), in consideration of some heterotrophic protists (< 100 µm) that are not considered in routine zooplankton analysis and benthic microalgae, which can be found in pelagic water samples.

This European Standard describes the necessary methods for measuring cell dimensions and for the calculation of cell or counting unit volumes to estimate the biovolume in phytoplankton samples. This shall be done using harmonized assignments of geometrical shapes to avoid errors.

2 Normative references

The following documents, in whole or in part, are normatively referenced in this document and are indispensable for its application. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

EN 15204, *Water quality - Guidance standard on the enumeration of phytoplankton using inverted microscopy (Utermöhl technique)*

EN 15972, *Water quality - Guidance on quantitative and qualitative investigations of marine phytoplankton*

EN 16698, *Water quality - Guidance on quantitative and qualitative sampling of phytoplankton from inland waters*

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

3.1

biomass

total mass of living organic matter within a system or taxon

3.2

biovolume

total volume of (living) organisms within a system or taxon

Note 1 to entry: The biovolume is usually expressed in cubic millimetres per litre (mm³/l).

3.3

cell volume

counting unit volume

total volume of a single cell or one counting unit

Note 1 to entry: The cell volume or counting unit volume includes the cell wall (if existing) but excludes lorica and/or mucilaginous envelopes and cell surface structures such as spines, bristles and scales.

Note 2 to entry: The cell volume or counting unit volume is usually expressed in cubic micrometres (µm³).

4 Principle

Generally, the estimation of the total or taxon specific biovolume in phytoplankton samples of natural communities or cultures is based on measurements of a representative number of individuals. By

multiplying the average or median cell or counting unit volume with the abundance, the total biovolume of each taxon in the sample is determined.

Three approaches are feasible:

- 1) Estimation by representative measurement:** A representative number of individuals (in most cases single cells) or counting units of all recorded or dominating taxa is measured in each sample or a specified number of samples within a comparable series. These data are used to calculate the average or median cell or counting unit volume of each taxon using the applied geometrical formulae.
- 2) Estimation using size classes based on representative measurements:** For taxa with a high variability in cell size (e.g. several diatoms, different stages in life cycle) reasonable size classes can be determined first, and then the individuals are assigned to both the relevant taxon and size class. Basis for the definition of the size classes are measurements in the same manner as described in (1).
- 3) Estimation using standard volumes based on representative measurements:** A reasonable general standard cell or counting unit volume is defined for each taxon once. These standard values are determined by representative measurements and calculated by the formula of the assigned geometrical shapes as described in (1).

A geometrical shape shall be assigned to each taxon in all approaches to calculate the cell or counting unit volume. Thus, to harmonize these approaches the geometrical shapes are pre-assigned to all taxa (see Annex D). These shapes have been chosen to reflect the corresponding taxa shapes as accurately as possible, and to allow effective taxa measurement with little effort (i.e. with as few dimensions as possible; usually only two are necessary). Seventeen different geometrical shapes are utilized (for the catalogue of geometrical shapes see Annex A). If it is impossible to describe the actual shape of a taxon with a simple basic geometrical shape, composite shapes (e.g. cone with half sphere) are used. If the actual geometry of taxa does not fit exactly to the assigned shape, a “geometry correction factor” is used for the final cell or counting unit volume calculation.

Taxon lists describing the preferred geometrical shapes have been published before (see e.g. [1], [3], [4]), based on specific taxonomical levels or for particular areas. This guidance document provides harmonized geometrical shapes for phytoplankton organisms spread across European marine, brackish, and freshwater systems. Annex D contains an alphabetical list of genera with the assigned geometrical shapes. If there are divergent forms on species, subspecies, form, or variety level within a genus they are listed as well.

5 Equipment and preservatives

The following equipment is required for biovolume analysis of phytoplankton samples.

5.1 Inverted microscope equipped with a condenser featuring a numeric aperture (NA) of at least 0,5 and plan objectives with a NA of 0,9 or more allowing for total magnification between 63× and 400× at a minimum. The microscope should have binocular, bright field (additional phase contrast is useful), 10× or 12,5× eyepieces.

Though inverted microscopy is the recommended method for analysing of phytoplankton, conventional (non-inverted) compound light microscopes may also be used for measuring phytoplankton under some conditions.

5.2 Calibrated object micrometre.

5.3 Eyepiece (ocular) micrometre.

5.4 Counting-graticule.

5.5 Sedimentation chambers according to EN 15204.

5.6 Image analysis software, if available.

5.7 Sampling bottles according to EN 15204.

5.8 Preservatives, acidic Lugol's iodine and/or alkaline Lugol's iodine according to EN 15204.

6 Procedure

6.1 Sampling and sample preparation

The sampling and determination of phytoplankton abundance and composition is a precondition for the calculation of the biovolume of a phytoplankton sample. Sampling shall be carried out according to EN 16698 for freshwater samples and EN 15972 for marine samples. For counting and species determination, see EN 15204.

The dimensions needed for the biovolume estimation of the relevant phytoplankton taxa are analysed in sedimentation chambers, which are prepared in the same manner as for counting and species determination (see EN 15204), using an inverted microscope and an eyepiece micrometre or image analysis software.

For specific scientific purposes, measurements can be carried out also with a conventional (non-inverted) compound light microscope.

6.2 Calibration of the eyepiece micrometre, counting-graticule and image analysis software

The required dimensions for estimation of the cell or counting unit volume shall be measured using an eyepiece (ocular) micrometre or an image analysis software. For the application of size classes a calibrated counting-graticule can also be used.

Prior to measurement, all systems shall be calibrated with a calibrated object micrometre for every microscope and all objectives and eyepieces used.

The scale of commercially available calibrated object micrometres has a length of 1 mm (or 2 mm) where the millimetre is divided into 100 equal parts. The distance between the graduation lines is 10 µm. By aligning the scale of the eyepiece micrometre with the scale of the object micrometre or the grid boxes of the counting-graticule, the scale value (*S*) or conversion factor of the eyepiece micrometre can be determined for each magnification as follows:

$$S = \frac{n_{obj} \times 10}{n_{eye}} \quad (1)$$

where

S is the scale value (conversion factor) for the eyepiece micrometre in micrometres (µm);

n_{obj} is the number of graduation lines of the object micrometre;

n_{eye} is the number of graduation lines of the eyepiece micrometre or the number of grid boxes of the counting-graticule.

The conversion factor should be specified with up to two decimal places. The intervals between the graduation lines of the scale shall be separately determined with the calibrated object micrometre for every objective used.

If image analysis software is used, this equipment shall be calibrated with the calibrated object micrometre separately for every level of magnification, following the instructions in the operating manual of the software.

6.3 Statistical requirements for determination

The required dimensions of the relevant geometrical shape shall be measured for each taxon of interest. At least 20 individuals per taxon should be measured to ensure that the standard error of cell or counting unit volume will be generally < 10 %.

For taxa, which are very variable in size, the number of measured cells/counting units should be increased until the standard error is < 10 %, to a maximum of 50 individuals ([3], [5]).

Where the size variability of a taxon is small, the number of measured cells may be minimized to only 5 to 10 individuals. In all cases, it is advisable to check that the standard error of cell or counting unit volume is low. The standard deviation and standard error can be calculated according to Formulae (2) and (3) as follows:

$$s = \sqrt{\frac{\sum_{i=1}^n (x_i - \bar{x})^2}{n-1}} \quad (2)$$

where

- s is the taxon volume standard deviation;
- x_i is the volume of a single cell/counting unit of the taxon;
- \bar{x} is the mean volume of all measurements of the taxon;
- n is the number of the respective measured cells of the taxon.

From the standard deviation s , the standard error of the mean $\sigma_{\bar{x}}$ can be calculated using Formula (3):

$$\sigma_{\bar{x}} = \frac{s}{\sqrt{n}} \quad (3)$$

To obtain the percentage range σ_{rel} of the standard error, divide the standard error of the mean $\sigma_{\bar{x}}$ by the mean value \bar{x} of the samples using Formula (4):

$$\sigma_{rel} = \frac{\sigma_{\bar{x}}}{\bar{x}} \times 100 \quad (4)$$

From the results, the 95 % confidence limits can be determined using Formulae (5) and (6):

$$\bar{x}_u = \bar{x} + (\sigma_{\bar{x}} \times 1,96) \quad (5)$$

$$\bar{x}_l = \bar{x} - (\sigma_{\bar{x}} \times 1,96) \quad (6)$$

where

\bar{x}_u is the upper 95 % confidence limit;

\bar{x}_l is the lower 95 % confidence limit.

Since the required number of individuals to be measured for any taxon is dependent upon the size variability of that taxon, it is helpful to calculate the cell or counting unit volume after each measurement. This will allow continuous statistical analysis and check of precision, thus minimizing the number of measurements required. Ideally, the confidence limits should be set at 95 % as a measure for precision. By ensuring that only as many cells as necessary for achieving this limit will be counted, the amount of laboratory work will be minimized.

If not enough cells/counting units of a taxon are present in the sample in order to achieve the minimum statistical requirements described, all of the (few) cells of this taxon shall be measured or a mean standard biovolume may be used (see below).

Measuring a high number of cells for every taxon in every sample is a time consuming procedure. For routine monitoring programmes mean cell or counting unit volumes calculated from own measurements, for a particular project and area, may be used. These mean volumes shall be checked regularly by measuring actual cell dimensions (see 6.4) and calculating actual cell and counting unit volume (see 6.5). For taxa with a high variability in cell size and representing more than 50 % of the total biovolume, these checks are strongly advised.

6.4 Measurement

6.4.1 General

Measurement of the cells/counting units can be carried out in a separate step or parallel to the counting process. Depending on the cell size of the taxa, the determination of the required dimensions (e.g. diameter, height, length, width, etc.) should be carried out at magnifications between 63× and 1 000×, in order to obtain corresponding precision.

The so-called empty magnification, which does not reveal any new detail, should be avoided. Therefore, the total magnification of the microscope should preferably be higher than 500× but smaller than 1 000× of the NA of the used objectives to work in the area of useful magnification.

It is important that the cells to be measured are chosen randomly to avoid a discrimination of special size classes. This can be achieved by selecting the cells from randomly distributed visual boxes all over the sedimentation chamber.

If single cells can clearly be distinguished in chain-building or filamentous species or other colony forming taxa, only one cell per chain, filament or colony should be measured. Filamentous algae often form lumps and are distributed very unevenly in the chamber. Also in these cases, it shall be ensured that cells from different lumps are measured.

By rotating the eyepiece micrometre and moving the sedimentation chamber with the microscope stage, the scale of the eyepiece micrometre is placed over the required dimension of the cell/counting unit to be measured. The number of covered graduation lines is read, and with the application of the conversion factor (see 6.2), the length of the dimension is calculated by multiplication.

The measurement results should be reported in micrometres, with up to two decimal places. If the end of the cell dimension to be measured is between two graduation lines of the ocular scale, the share is to be estimated to a maximum of one decimal place.

When using the size classes approach based on representative measurements, the calibrated counting-graticule may also be used for the assignment of the individuals to the corresponding size classes.

When image analysis software is used, the corresponding details of the operating manual shall be followed.

6.4.2 Using size classes based on representative measurements

The appropriate number of size classes depends on the size variation of each taxon. The size classes, for example, can be derived from cluster analyses applied to a representative dataset of measured cells of the taxon (see 6.4.1).

For each size class the average cell or counting unit volume is calculated using the formula of the taxon specific geometrical shape with mean dimension lengths. By multiplication with the respective abundance and addition of all size classes of a taxon, the total taxon specific biovolume in the sample is determined. In routine monitoring programmes, standard size classes should be used (e.g. [4]). If regional lists of size classes are available, they should be used instead [4]. It is recommended to create a new size class, when the biovolume of individuals significantly exceeds the biovolume of existing size classes.

6.4.3 How to deal with hidden dimensions

For some particular taxa, it is often not possible to measure all dimensions needed to calculate the cell volume ("hidden dimensions", e.g. height of prismatic cells of different shapes or small diameter of elliptical cells) during routine analysis. Then it is necessary to estimate the length of missing dimensions as a proportion of one of the visible dimensions using a calculated species-specific factor. The visible to hidden size relationship can be obtained from measurements of other samples from the same sample series or from the same area if the position of the cells allows this. Measurements should be used preferentially over estimates. The taxon list in Annex D gives suggestions for the dimension relations for most of the taxa, which usually have a "hidden dimension". For those taxa, where no suggestion for the "hidden dimension" is given, corresponding nominal values shall be taken from the literature if available. If the "hidden dimension" shall be determined exactly for a special problem, the cells can be turned around in paraffin oil. With special microscopes, it is also possible to measure the distance between focus of the upper and lower end of the "hidden dimension".

6.4.4 Measurement of filamentous taxa

With some filamentous taxa (e.g. cyanobacteria), it is often difficult to distinguish individual cells within a filament, especially when the cells are directly connected without any gaps. In such cases, filament pieces of a fixed length, e.g. 100 µm or 10 µm, can be counted and measured (100 µm or 10 µm length and diameter), and multiplied by the total enumeration of this counting unit in the sample.

Alternatively, mean dimensions of filaments can be measured to calculate the volume of one filament, a value that is then multiplied by the number of filaments in the sample.

A third method, which is more precise for filamentous forms, particularly those, which have no distinct boundaries between cells, is as follows: Instead of counting individual filaments, the total length of the fraction of each filament that is within the boundaries of a counting grid shall be measured, ignoring the fraction outside of the grid boundaries (see Figure 1). The sum of the total length of all fractions of filaments within the grid shall be calculated after counting of the transect is completed. Afterwards, the diameter of at least 20 filaments shall be measured and the mean filament diameter is calculated. To obtain the biovolume of the respective taxon, the sum of total filament lengths (h) shall be multiplied with the square of the median filament diameter and the factor of $\pi/4$, because the filament is a cylinder with the volume:

$$V = \frac{1}{4} \times \pi \times d^2 \times h \quad (7)$$

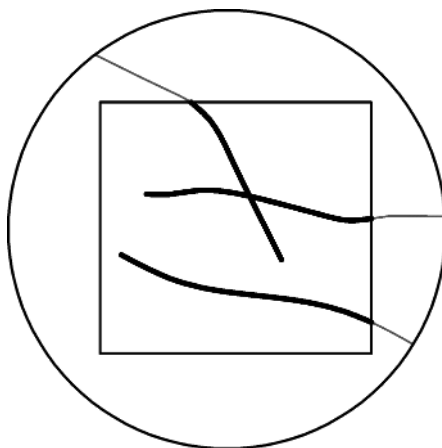


Figure 1 — Determining biovolume for filaments without distinct cell boundaries using a counting grid

NOTE The lengths of all fractions within the counting grid (black rectangle) are measured, excluding fractions outside of the counting grid boundaries [6].

6.4.5 Measurement and counting of colony- and coenobium-forming taxa

In most cases, the assignment of a geometrical shape should be based on the shape of an individual cell. For some colony- and coenobium-forming species where individual cells are difficult to distinguish or have very complex contours, it can be expedient to assign a geometrical shape based on the shape of the whole colony or coenobium or to use ultrasonic treated samples (where the association of cells have become disintegrated into individual cells) for counting and measuring the cells.

For example in some species, small individual cells are aggregated into compact spatial colonies. The individual cells are usually indistinguishable, and the number may be very difficult to assess in these colonies or coenobia. In such cases, the estimation of the volume can be based on the geometry of the entire colony or coenobium, in particular during the analysis of routine monitoring samples. For some taxa an additional geometrical shape correction factor shall be taken into account for the volume calculation. The same applies for colony- or coenobium-forming taxa with very complex contours of individual cells. Table 1 lists examples. In Annex D, it is specified for the respective taxa if the entire colony or coenobium with its geometry should be measured.

Table 1 — Geometrical shapes and correction factors for some colony- and coenobium-forming taxa.

Taxon name	Geometry based on colony	Geometrical shape correction factor	Hidden dimension factor
<i>Woronichinia</i>	sphere	0,2	-
<i>Coelosphaerium</i>	sphere	0,2	-
<i>Snowella</i>	sphere	0,75	-
<i>Botryococcus</i>	spheroid	-	-
<i>Eudorina</i>	sphere	0,25	-
<i>Pandorina</i>	sphere	-	-
<i>Coelastrum</i>	sphere	-	-
<i>Pediastrum</i>	cylinder	species dependend	height of colony = height of single cell (factor see Annex D)
<i>Crucigenia</i>	cuboid	species dependend	third edge length (height) = 0,5×second edge length (width)

A more precise method to achieve the biovolume of such colonial forms, especially the cyanobacterial genera *Microcystis*, *Aphanothece*, *Aphanocapsa*, and the *Dolichospermum* species forming “ball of yarn” colonies, is to measure cell dimensions and the greatest axial linear dimension of the colony in the sedimented sample, and then separate the colonies by ultrasonic treatment, and count the individual cells in the treated sample [7].

6.4.6 Measurement of complex geometrical shapes

Some taxa show very complex cell outlines requiring a composition of multiple geometrical shapes and thus, the application of complicated formulae for biovolume calculations. In order to ease that work, some simplified combined forms have been assigned to these taxa that can be easily measured and will require only minimum effort for the estimation. However, depending on the required precision, more complex and thus more precise geometrical subdivisions shall be applied to those taxa. Another possibility is to resort to pre-determined cell volumes from literature (mean standard cell or counting unit volume), bearing in mind that the size of the cells depends on a number of environmental factors and thus, may vary widely. On the other hand, cell volume estimation for some species is easier on half-cell basis (for example in some desmid genera like *Cosmarium* and *Staurastrum*).

In any case, everything shall be documented in the protocol. As an example, Annex E shows how to measure the needed dimensions for four taxa from the conducted interlaboratory comparison as well as for two additional taxa, which are not easy to measure.

6.5 Calculation of biovolume

The cell or counting unit volume is calculated on the basis of the taxon-specific geometrical shape and the linear dimensions determined for the individual cell or counting unit (e.g. diameter, height, length, width, etc.; see Annex A). For some taxa, the associated geometrical form will not fit exactly to the actual cell shape, for example, if a cuboid is assigned to a pennate diatom with rounded apical cell ends. For such cases, the cell volume shall be multiplied by a correction factor, which is given in Annex D for relevant taxa.

The average cell or counting unit volumes of the various taxa shall be generally calculated as the median of all individual cell or counting unit volumes.

NOTE If the individual volumes per taxon are normally distributed (according to Chi-squared test or Kolmogorov-Smirnov test), the arithmetic mean can be used for the calculation of the average volume instead of the median. For answering specific questions in ecology, it can be necessary to use also the arithmetic mean for non-normally distributed values.

These calculations shall be carried out for every phytoplankton taxon recorded in all or selected samples using the “representative measurement” approach (see Clause 4) or basically once for characteristic samples using the “standard factor” approach (see Clause 4). If applying the “size class” approach (see Clause 4 and 6.4.2) the average cell or counting unit volume for each size class is calculated using the mean of the upper and lower dimensions of size class borders.

The biovolume per taxon in a sample is calculated by multiplying the number of cells/l (or cells/ml) or counting units (e.g. number of 100 µm filament pieces per liter) with the median (or mean) of the determined taxon-specific cell or counting unit volumes (µm³) as measured by one of the three methods listed above:

$$V_{\text{bio},i} = \frac{n_i \times \tilde{V}_i}{10^9} \quad (8)$$

where

$V_{\text{bio},i}$ is the biovolume of taxon or size class i in cubic millimetres per litre (mm³/l);

n_i is the number of cells (or number of counting units) of taxon or size class i per litre (l⁻¹);

\tilde{V}_i is the median (or mean) of the cell or counting unit volumes of taxon or size class i in cubic micrometres (μm^3).

The total biovolume to be determined for each sample results from the sum of the biovolume determined for each phytoplankton taxon or size class.

Statistical performance data from the conducted European wide interlaboratory comparison for validation are listed in Annex B.

6.6 Biovolume biomass relations

In phytoplankton ecology, the biomass is usually expressed as chlorophyll-a concentration ($\mu\text{g/l}$), biovolume (mm^3/l) or carbon content ($\mu\text{g/l}$). Assuming the density of organisms being equal to the density of water ($1,0 \text{ g/cm}^3$, [8]), the biomass as wet weight may be estimated as follows:

$$1 \text{ mm}^3/\text{l} (\text{biovolume}) = 1 \text{ cm}^3/\text{m}^3 (\text{biovolume}) = 1 \text{ mg/l} (\text{wet weight});$$

$$1 \text{ mm}^3/\text{m}^3 (\text{biovolume}) = 10^6 \mu\text{m}^3/\text{l} (\text{biovolume}) = 1 \mu\text{g/l} (\text{wet weight}).$$

NOTE For the estimation of the carbon content, see Annex C.

6.7 Reporting

The specific cell or counting unit volumes are expressed in cubic micrometres (μm^3) without decimal places or for picoplankton with two significant places (e.g. $4,2 \mu\text{m}^3$). The biovolume of a single taxon and the complete sample is given in cubic millimetres per litre (mm^3/l or mm^3/ml) with three significant digits or places (e.g. $12,5 \text{ mm}^3/\text{l}$, $3,75 \text{ mm}^3/\text{l}$, $0,138 \text{ mm}^3/\text{l}$, $0,004 \text{ mm}^3/\text{l}$).

As an example, the taxon specific part of a sample report can contain information as shown in Table 2.

Table 2 — Example report for taxon specific sample part.

Taxon name	Abundance	Geometry	Average cell volume	Total biovolume	Carbon content (according to ...)
	n/l		μm^3	mm^3/l	$\mu\text{g/l}$
<i>Thalassiosira nordenskiöldii</i>	12 109	cylinder	9 817	0,119	6,53
<i>Odontella aurita</i>	265 725	elliptic cylinder	14 091	3,74	188,5
....					
Sum of sample 5	765 180			4,43	231,2

Some taxa (e.g. flagellates without solid cell wall) are inclined to shrink during the fixation process. Shrinkage depends on many influencing factors (preservative, life stage, physiological status, species, etc., even diatoms can shrink). Often it is difficult to find good correction factors for this process in the literature. As a consequence, applying a general correction factor to preserved material can lead either to overestimation or to underestimation of the biovolume. For monitoring activities with the objective of detecting trends, the exact compensation by preservation correction factors is not of the highest priority. In other cases where it might be mandatory to determine the exact biomass, correction factors for influence of fixation should be derived from comparisons between fixated and living organisms. This shall be noted in the protocol.

7 Quality Assurance

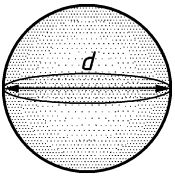
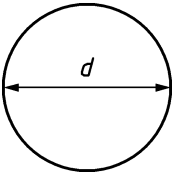
The quality assurance associated with this European Standard should be in accordance with EN 14996.

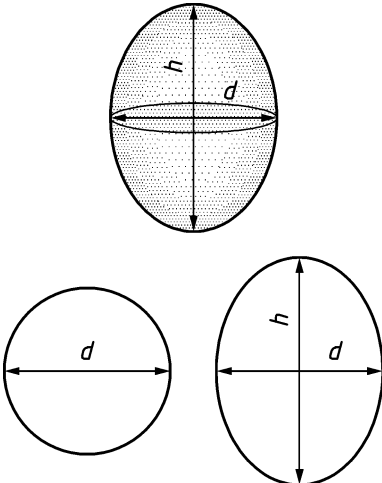
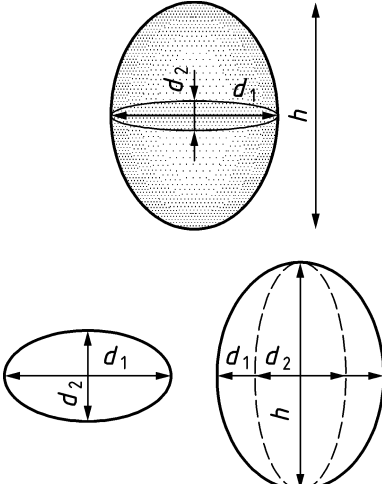
Annex A (informative)

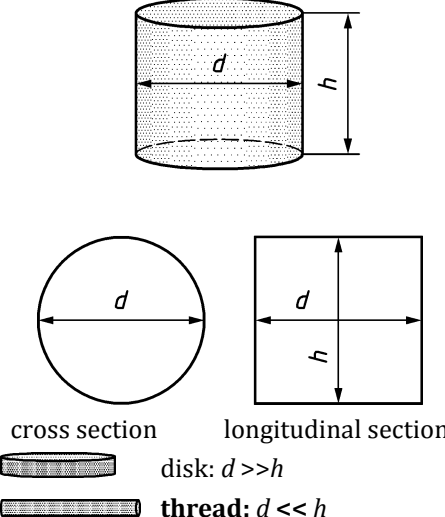
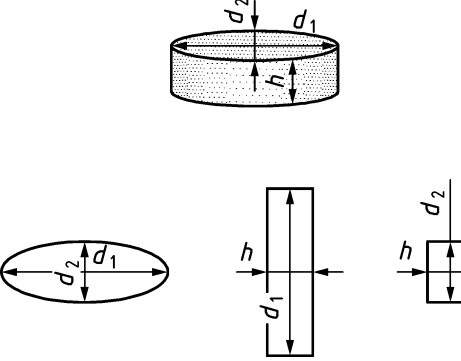
List of geometrical shapes

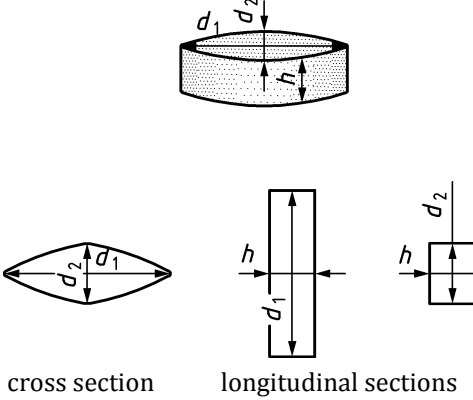
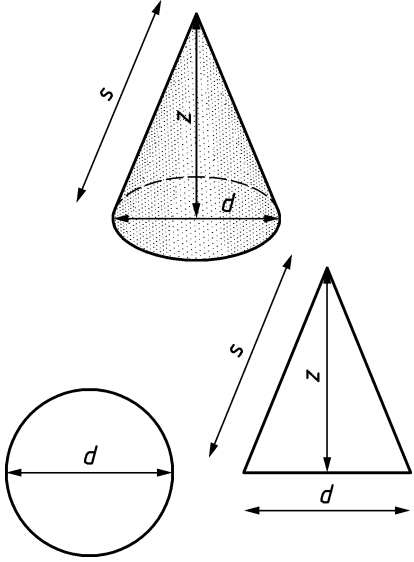
Table A.1 provides information about 17 different geometrical shapes used for the biovolume estimation of phytoplankton. The first column simply contains a consecutive number (ID-number) for each shape. Listed in the second column is the official geometrical name of the form alongside any other synonymous names. The images of each shape show a 3D-view, a cross-section, and a longitudinal-section in which the dimensions are designated to be used for biovolume calculation. The third column shows some general information about each shape, a description about the needed dimensions as well as some taxon examples. Firstly, it is listed if it is a basic shape or a composed form. If the latter is the case it is also indicated from which basic shapes the form is composed. The volume calculation precision distinguishes between exact calculation, if there is a clearly defined formula for volume, and approximate calculation, if the formula is derived by approximate integration. The dimensions necessary for calculation of biovolume are listed with full name and abbreviation as well as with information about measurability of these dimensions in phytoplankton organisms using an inverted microscope. “m” means that this dimension is measurable, “hd” means that this is a hidden dimension that cannot be measured in a sedimentation chamber. If both specifications are listed (m; hd), this indicates that cells can have different positions, the orientation of which can affect if the dimension is measurable or not. Finally, some typical taxon examples for each shape are listed. In the last column, the formulae needed for biovolume calculation are specified, with the deduced form based on diameter that can clearly be measured in the cells, and with the original geometrical form based on radius.

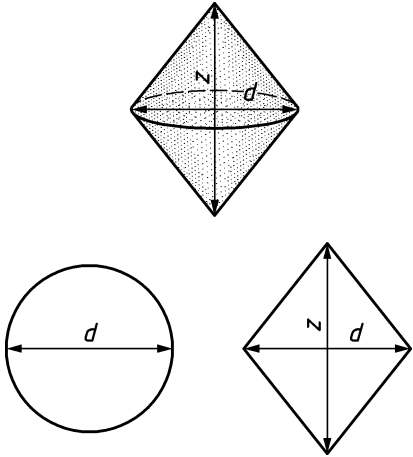
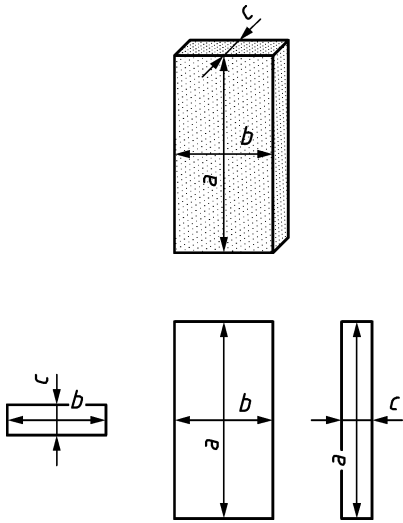
Table A.1 — Geometrical shapes

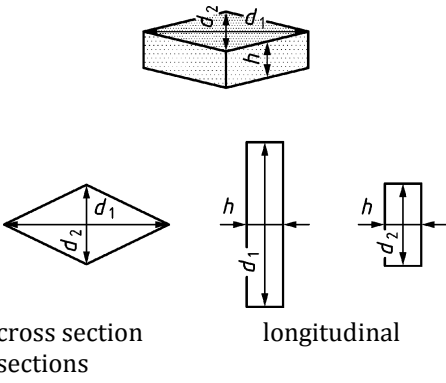
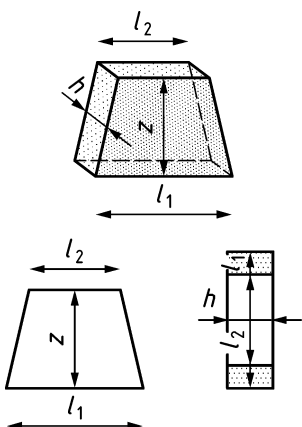
ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
1	Sphere   cross section	<i>basic shape</i> <i>Volume calculation precision:</i> <i>exact</i> Dimensions: <i>diameter (d) - m</i> Examples: <i>Chroococcus, Coelastrum,</i> <i>Gloeocapsa, Microcystis, Oblea,</i> <i>Pterosperma</i>	$V = \frac{1}{6} \times \pi \times d^3$ For radius $r = \frac{d}{2}$: $V = \frac{4}{3} \times \pi \times r^3$

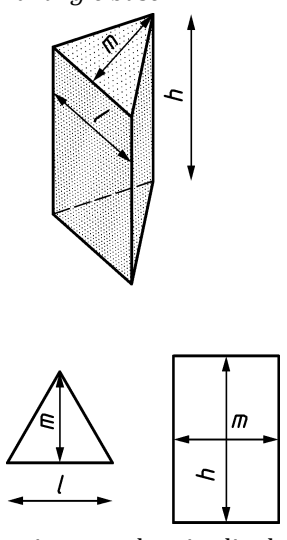
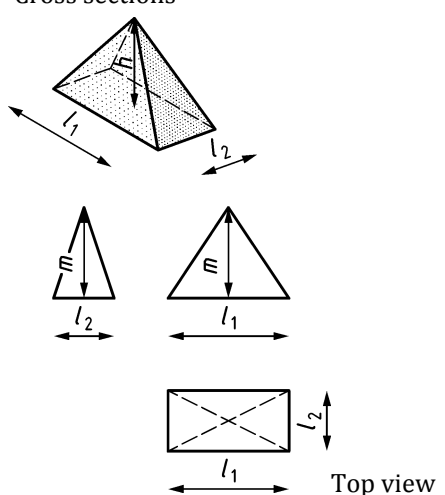
ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
2	<p>Spheroid <i>rotational ellipsoid</i> <i>ellipsoid of revolution</i></p>  <p>cross section longitudinal section</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>diameter (d) - m</i> <i>height (h) - m</i></p> <p>Examples: <i>Aphanothece, Chlamydomonas, Desmodesmus, Trachelomonas, Woronichinia</i></p>	$V = \frac{1}{6} \times \pi \times d^2 \times h$ <p>For radius $r = \frac{d}{2}$:</p> $V = \frac{4}{3} \times \pi \times r^2 \times \frac{1}{2} \times h$
3	<p>Ellipsoid <i>tri-axial ellipsoid,</i> <i>flattened ellipsoid</i></p>  <p>cross section longitudinal section</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>large diameter (d₁) - m</i> <i>small diameter (d₂) - hd</i> <i>height (h) - m</i></p> <p>Examples: <i>Amphidinium, Cosmarium, Dinophysis, Gymnodinium, Mallomonas</i></p>	$V = \frac{1}{6} \times \pi \times d_1 \times d_2 \times h$ <p>For radius r:</p> $V = \frac{4}{3} \times \pi \times r_1 \times r_2 \times r_3$ <p>with</p> $r_1 = \frac{d_1}{2}; \quad r_2 = \frac{d_2}{2}; \quad r_3 = \frac{h}{2}$

ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
4	<p>Cylinder <i>circle based cylinder</i></p>  <p>cross section longitudinal section</p> <p>disk: $d \gg h$ thread: $d \ll h$</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: diameter (d) - m height (h) - m; hd</p> <p>Examples: <i>Aphanizomenon, Aulacoseira, Coscinodiscus, Phormidium, Rhizosolenia, Thalassiosira</i></p>	$V = \frac{1}{4} \times \pi \times d^2 \times h$ <p>For radius $r = \frac{d}{2}$:</p> $V = \pi \times r^2 \times h$
5	<p>Elliptic cylinder <i>prism on elliptic base</i> <i>oval cylinder</i></p>  <p>cross section longitudinal sections</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: large diameter (d_1) - m small diameter (d_2) - m; hd height of cylinder (h) - hd; m</p> <p>Examples: <i>Achnanthes, Chaetoceros, Fragilaria, Odontella, Surirella</i></p>	$V = \frac{1}{4} \times \pi \times d_1 \times d_2 \times h$ <p>For radius r:</p> $V = \pi \times r_1 \times r_2 \times h$ <p>with</p> $r_1 = \frac{d_1}{2}; \quad r_2 = \frac{d_2}{2}$

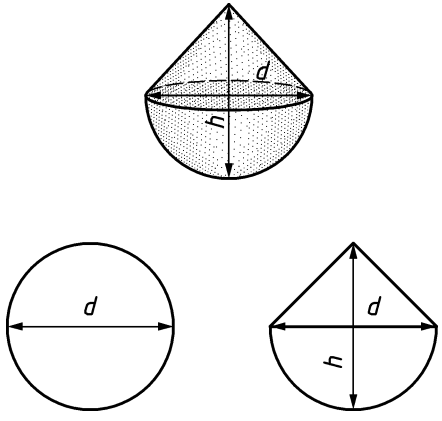
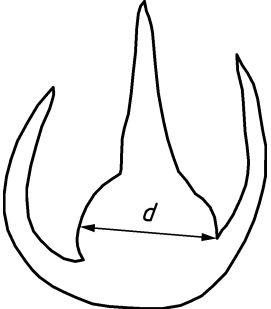
ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
6	<p>Lanceolate cylinder <i>naviculoid</i></p>  <p>cross section longitudinal sections</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision:</i> <i>approximate</i></p> <p>Dimensions: large diameter (d_1) - m small diameter (d_2) - m; hd height of cylinder (h) - hd; m</p> <p>Examples: <i>Haslea, Navicula</i></p>	$V = \frac{2}{\pi} \times d_1 \times d_2 \times h$ <p>For radius r:</p> $V = \frac{8}{\pi} \times r_1 \times r_2 \times h$ <p>with</p> $r_1 = \frac{d_1}{2}; \quad r_2 = \frac{d_2}{2}$
7	<p>Cone</p>  <p>cross section longitudinal section</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision:</i> <i>exact</i></p> <p>Dimensions: diameter (d) - m slant height (s) - m height of cone (z) - hd</p> <p>NOTE With an inverted microscope, the slant height can exactly be measured. The cone height is a hidden dimension because of the oblique projection.</p> <p>Examples: <i>Calciopappus, Minuscula, Pyramimonas longicauda</i></p>	$V = \frac{1}{12} \times \pi \times d^2 \times \sqrt{s^2 - \frac{1}{4} \times d^2}$ <p>For radius $r = \frac{d}{2}$ and</p> <p>height $z = \sqrt{s^2 - r^2}$:</p> $V = \frac{1}{3} \times \pi \times r^2 \times z$

ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
8	<p>Double cone <i>two cones</i></p>  <p>cross section longitudinal section</p>	<p><i>composed shape</i> <i>basic shape: cone</i> <i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>diameter (d) - m</i> <i>height of double cone (z) - m</i></p> <p>NOTE Due to the position in the sedimentation chamber, the total height can only be measured approximately because of the oblique projection.</p> <p>Examples: <i>Gyrodinium spirale,</i> <i>Heterocapsa triquetra,</i> <i>Protoperidinium crassipes</i></p>	$V = \frac{1}{12} \times \pi \times d^2 \times z$ <p>For radius $r = \frac{d}{2}$:</p> $V = 2 \times \frac{1}{3} \times \pi \times r^2 \times \frac{1}{2} \times z$
9	<p>Cuboid <i>rectangular box</i> <i>rectangular parallelepiped</i></p>  <p>cross section longitudinal sections</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>first edge length (a) - m</i> <i>second edge length (b) - m; hd</i> <i>third edge length (c) - hd; m</i></p> <p>Examples: <i>Bacillaria, Hantzschia,</i> <i>Pinnularia, Tabellaria</i></p>	$V = a \times b \times c$

ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
10	<p>Rhombic prism <i>parallelepiped prism on parallelogram base</i></p>  <p>cross section sections longitudinal sections</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>large diagonal (d₁) - m</i> <i>small diagonal (d₂) - m; hd</i> <i>height of prism (h) - hd; m</i></p> <p>Examples: <i>Brachysira, Nitzschia, Pseudonitzschia</i></p>	$V = \frac{1}{2} \times d_1 \times d_2 \times h$
11	<p>Trapezoid prism</p>  <p>cross section longitudinal section</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>length of large trapezoid side (l₁) - m</i> <i>length of small trapezoid side (l₂) - m</i> <i>height of trapezoid (z) - m; hd</i> <i>height of prism (h) - hd; m</i></p> <p>Examples: <i>Crucigenia fenestrata</i></p>	$V = \frac{1}{2} \times (l_1 + l_2) \times z \times h$

ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
12	<p>Triangular prism <i>prism on triangle base</i></p>  <p>cross section longitudinal section</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>length of triangle side (l) - m</i> <i>height of triangle (m) - m; hd</i> <i>height of prism (h) - hd; m</i></p> <p>NOTE For a prism on basis of an equilateral triangle, only the triangle side is needed. The triangle height can be expressed by this side.</p> <p>Examples: <i>Bellerochea, Ditylum, Goniochloris, Lithodesmium, Triceratium</i></p>	$V = \frac{1}{2} \times l \times m \times h$ <p>or</p> $V = \frac{1}{4} \times \sqrt{3} \times l^2 \times h$ <p>for a prism on basis of an equilateral triangle with</p> $m = \frac{1}{2} \times \sqrt{3} \times l$
13	<p>Pyramid</p> <p>Cross sections</p>  <p>Top view</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>length of first basic side (l₁) - m</i> <i>length of second basic side (l₂) - m; hd</i> <i>height of pyramid (h) - hd; m</i></p> <p>Examples: <i>Gomphonema, Licmophora</i></p>	$V = \frac{1}{3} \times l_1 \times l_2 \times h$

ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula
14	<p>Tetrahedron</p> <p>lateral view top view</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: exact</i></p> <p>Dimensions: edge length (<i>a</i>) - m height of tetrahedron (<i>h</i>) - hd</p> <p>Examples: <i>Staurastrum, Staurodesmus, Tetraedriella, Tetraedron</i></p>	$V = \frac{1}{12} \times \sqrt{3} \times a^2 \times h$ <p>or</p> $V = \frac{1}{12} \times \sqrt{2} \times a^3$ <p>for a regular tetrahedron with</p> $h = \frac{1}{3} \times \sqrt{6} \times a;$
15	<p>Spindle</p> <p>cross section longitudinal section</p>	<p><i>basic shape</i></p> <p><i>Volume calculation precision: approximate</i></p> <p>Dimensions: diameter (<i>d</i>) - m height (<i>h</i>) - m</p> <p>Examples: <i>Ankistrodesmus, Closterium, Monoraphidium</i></p>	$V = \frac{2}{15} \times \pi \times d^2 \times h$ <p>For radius $r = \frac{d}{2}$:</p> $V = \frac{8}{15} \times \pi \times r^2 \times h$

ID	Geometrical shape (<i>Synonyms</i>)	Information	Volume formula																																				
16	<p>Cone with half sphere <i>cone with hemisphere</i></p>  <p>cross section longitudinal section</p>	<p><i>composed shape</i> <i>basic shapes: cone, sphere</i> <i>Volume calculation precision: exact</i></p> <p>Dimensions: <i>diameter (d) - m</i> <i>total height (h) - m</i></p> <p>NOTE Due to the position in the sedimentation chamber, the total height can only be measured approximately because of the oblique projection.</p> <p>Examples: <i>Gonyaulax, Hemiselmis, Katodinium, Teleaulax</i></p>	$V = \frac{1}{12} \times \pi \times d^2 \times \left(h + \frac{1}{2} \times d \right)$ <p>For radius $r = \frac{d}{2}$:</p> $V = \frac{1}{3} \times \pi \times r^2 \times (h - r) + \frac{1}{2} \times \frac{4}{3} \times \pi \times r^3$																																				
17	<p>Ceratium-shape</p>  <p>lateral view</p>	<p><i>Complicated composed shape</i></p> <p><i>Volume calculation precision: approximate</i></p> <p>Dimensions: <i>girdle diameter (d) - m</i></p> <p>Examples: <i>Ceratium</i></p>	$V = x \times d^y$ <p>x and y are factors derived from measurements [4], [9]</p> <table border="0"> <tr> <td><i>Ceratium</i></td> <td>x: 0,32359</td> </tr> <tr> <td></td> <td>y: 2,9953</td> </tr> <tr> <td><i>C, furca</i></td> <td>x: 2,3038</td> </tr> <tr> <td></td> <td>y: 2,532</td> </tr> <tr> <td><i>C, fusus</i></td> <td>x: 35,198</td> </tr> <tr> <td></td> <td>y: 1,9156</td> </tr> <tr> <td><i>C, hirundinella</i></td> <td>x: 2,3038</td> </tr> <tr> <td></td> <td>y: 2,532</td> </tr> <tr> <td><i>C, horridum</i></td> <td>x: 0,32437</td> </tr> <tr> <td></td> <td>y: 3,0474</td> </tr> <tr> <td><i>C, lineatum</i></td> <td>x: 1,2375</td> </tr> <tr> <td></td> <td>y: 2,5989</td> </tr> <tr> <td><i>C, longipes</i></td> <td>x: 0,32437</td> </tr> <tr> <td></td> <td>y: 3,0474</td> </tr> <tr> <td><i>C, macroceros</i></td> <td>x: 0,32437</td> </tr> <tr> <td></td> <td>y: 3,0474</td> </tr> <tr> <td><i>C, tripos</i></td> <td>x: 0,32359</td> </tr> <tr> <td></td> <td>y: 2,9953</td> </tr> </table>	<i>Ceratium</i>	x: 0,32359		y: 2,9953	<i>C, furca</i>	x: 2,3038		y: 2,532	<i>C, fusus</i>	x: 35,198		y: 1,9156	<i>C, hirundinella</i>	x: 2,3038		y: 2,532	<i>C, horridum</i>	x: 0,32437		y: 3,0474	<i>C, lineatum</i>	x: 1,2375		y: 2,5989	<i>C, longipes</i>	x: 0,32437		y: 3,0474	<i>C, macroceros</i>	x: 0,32437		y: 3,0474	<i>C, tripos</i>	x: 0,32359		y: 2,9953
<i>Ceratium</i>	x: 0,32359																																						
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Annex B (informative)

Performance data

The performance data given in Table B.1 to Table B.4 were determined in a European interlaboratory comparison for validation carried out in March 2014 on two mixed marine and two freshwater samples taken from the North Sea (Helgoland, Germany), the Baltic Sea (Stralsund, Germany) and different lakes around Oldenburg (Germany) and Bad Saarow (Germany). 52 participants from the following European countries have provided results: Denmark (4), Finland (3), France (1), Germany (20), Ireland (1), Italy (3), Lithuania (3), Norway (1), Poland (2), Portugal (3), Spain (2), Sweden (3) and United Kingdom (6).

All results have been calculated according to the German standard document DIN 38402-45:2014-06.

Table B.1 — Performance data for measurements of first dimension in biovolume analysis of phytoplankton species in natural samples - Abbreviations for dimensions according to Annex A.

Taxon name	l	n	X μm	X_{med} μm	X_{mean} μm	X_{HS} μm	S_{R} μm	$C_{\text{V,R}}$ %	S_{r} μm	$C_{\text{V,r}}$ %
<i>Dactyliosolen fragilissimus (d)</i>	45	1 225	11,74	11,93	12,06	12,04	2,01	16,7	1,80	14,9
<i>Dinophysis acuminata (d₁)</i>	42	961	33,48	33,90	34,07	34,07	4,24	12,5	3,23	9,5
<i>Ceratium tripos (d₁)</i>	44	998	66,79	65,93	66,74	66,66	6,12	9,2	4,56	6,8
<i>Pseudo-nitzschia cf.pungens (d₁)</i>	44	1 054	111,87	111,69	111,12	111,19	12,43	11,2	11,18	10,1
<i>Thalassionema nitzschioides (d₁)</i>	42	1 013	32,95	32,82	33,15	33,15	4,29	13,0	4,25	12,8
<i>Rhizosolenia imbricata (d)</i>	42	1 154	9,68	10,19	10,16	10,19	2,49	24,4	2,14	21,0
<i>Ditylum brightwellii (l)</i>	40	1 114	26,76	23,83	23,64	23,69	4,89	20,7	4,18	17,6
<i>Stephanopyxis turris (d)</i>	37	964	43,15	40,35	39,96	39,96	7,60	19,0	7,07	17,7
<i>Odontella sinensis (d₁)</i>	40	1 106	167,28	163,87	175,38	174,21	40,62	23,3	33,02	19,0
<i>Chaetoceros debilis (d₁)</i>	39	982	13,39	14,25	13,65	13,65	3,84	28,2	2,77	20,3
<i>Fragilaria crotonensis (d₁)</i>	53	1 391	76,94	76,58	78,15	77,47	12,51	16,2	14,27	18,4
<i>Tabellaria fenestrata (a)</i>	47	1 052	51,62	50,91	50,77	50,77	4,77	9,4	4,20	8,3
<i>Aulacoseira granulata (d)</i>	50	1 183	5,48	5,88	5,89	5,90	1,02	17,3	0,84	14,2
<i>Cryptomonas erosa (d₁)</i>	50	1 133	12,63	12,83	12,96	12,96	1,58	12,2	1,32	10,2
<i>Rhodomonas lacustris (d)</i>	49	1 249	5,20	5,27	5,28	5,28	0,66	12,5	0,71	13,5
<i>Planktothrix agardhii (d)</i>	47	1 260	3,32	3,34	3,44	3,42	0,72	21,0	0,15	4,3
<i>Monoraphidium arcuatum (d)</i>	42	1 085	1,73	1,69	1,83	1,78	0,54	30,3	0,32	17,8
<i>Woronichinia naegeliana (d)</i>	44	1 184	2,92	3,27	3,26	3,26	0,60	18,3	0,39	12,0
<i>Cosmarium ocellatum (d₁)</i>	43	977	28,60	26,21	26,44	26,46	2,54	9,6	1,85	7,0
<i>Trachelomonas hispida (d)</i>	45	1 068	21,15	20,96	20,86	20,87	2,08	10,0	1,75	8,4
Explanation of symbols										
l	number of laboratories with $ z_u\text{-score} < 3$									
n	number of total individual test results of all laboratories with $ z_u\text{-score} < 3$									
X	assigned reference value based on 7 replicates with each having a minimum of 30 single measurements									
X_{med}	interlaboratory ^a median value									
X_{mean}	interlaboratory ^a arithmetic mean value									
X_{HS}	interlaboratory ^a robust mean value (Hampel Schaetzer)									
S_{R}	reproducibility standard deviation									
$C_{\text{V,R}}$	relative reproducibility standard deviation									
S_{r}	repeatability standard deviation									
$C_{\text{V,r}}$	relative repeatability standard deviation									
^a	Based on arithmetic means of individual laboratory measurements; only laboratories with $ z_u\text{-score} < 3$ included.									

Table B.2 — Performance data for measurements of second dimension in biovolume analysis of phytoplankton species in natural samples - Abbreviations for dimensions according to Annex A.

Taxon name	<i>l</i>	<i>n</i>	<i>X</i> µm	<i>x</i> _{med} µm	<i>x</i> _{mean} µm	<i>x</i> _{HS} µm	<i>s</i> _R µm	<i>C</i> _{V,R} %	<i>s</i> _r µm	<i>C</i> _{V,r} %
<i>Dactyliosolen fragilissimus (h)</i>	45	1 225	60,61	59,69	59,32	59,32	12,01	20,3	11,86	20,0
<i>Dinophysis acuminata (d₂)</i>	43	994	20,09	19,45	20,58	20,32	5,58	27,5	1,90	9,4
<i>Ceratium tripos (d₂)</i>	45	1 018	43,41	38,00	38,53	38,09	11,82	31,0	2,12	5,6
<i>Pseudo-nitzschia cf.pungens (d₂)</i>	43	1 024	4,81	4,51	4,59	4,58	1,01	22,0	0,84	18,4
<i>Thalassionema nitzschioides (d₂)</i>	42	1 013	3,99	4,35	4,26	4,23	0,99	23,4	0,77	18,2
<i>Rhizosolenia imbricata (h)</i>	42	1 154	188,20	201,34	200,16	200,46	45,13	22,5	41,59	20,7
<i>Ditylum brightwellii (m)</i>	40	1 114	23,17	22,99	22,87	22,96	4,47	19,5	4,51	19,7
<i>Stephanopyxis turris (h)</i>	37	964	66,83	59,20	58,30	58,30	10,36	17,8	9,62	16,5
<i>Odontella sinensis (d₂)</i>	40	1 106	66,91	64,27	64,17	63,63	20,62	32,4	11,50	18,1
<i>Chaetoceros debilis (d₂)</i>	38	959	10,04	9,31	9,53	9,40	2,59	27,6	1,59	16,9
<i>Fragilaria crotonensis (d₂)</i>	47	1 171	4,40	3,96	3,99	3,98	0,96	24,2	0,23	5,9
<i>Tabellaria fenestrata (b)</i>	47	1 052	5,16	7,57	7,39	7,41	2,33	31,4	1,07	14,4
<i>Aulacoseira granulata (h)</i>	43	966	27,33	26,04	26,11	26,11	3,16	12,1	2,68	10,3
<i>Cryptomonas erosa (d₂)</i>	49	1 113	10,73	10,13	9,95	9,99	2,10	21,1	0,95	9,5
<i>Rhodomonas lacustris (h)</i>	49	1 249	11,43	10,53	10,61	10,61	1,85	17,4	1,67	15,7
<i>Planktothrix agardhii</i>	-	-	-	-	-	-	-	-	-	-
<i>Monoraphidium arcuatum (h)</i>	41	1 065	49,31	42,00	41,25	41,25	7,92	19,2	4,98	12,1
<i>Woronichinia naegeliana (h)</i>	43	1 077	5,35	5,38	5,33	5,36	0,69	12,9	0,49	9,2
<i>Cosmarium ocellatum (d₂)</i>	43	956	14,30	13,39	13,52	12,19	2,52	20,7	0,87	7,2
<i>Trachelomonas hispida (h)</i>	45	1 068	27,12	27,13	26,83	26,80	3,01	11,2	2,35	8,8

For explanation of symbols, see Table B.1.

Table B.3 — Performance data for measurements of third dimension in biovolume analysis of phytoplankton species in natural samples - Abbreviations for dimensions according to Annex A

Taxon name	<i>l</i>	<i>n</i>	<i>X</i> µm	<i>x</i> _{med} µm	<i>x</i> _{mean} µm	<i>x</i> _{HS} µm	<i>s</i> _R µm	<i>C</i> _{V,R} %	<i>s</i> _r µm	<i>C</i> _{V,r} %
<i>Dactyliosolen fragilissimus</i>	-	-	-	-	-	-	-	-	-	-
<i>Dinophysis acuminata (h)</i>	42	961	46,69	48,71	48,88	48,77	5,38	11,0	4,58	9,4
<i>Ceratium tripos (h₁)</i>	42	938	40,33	42,67	41,39	41,44	6,27	15,1	3,34	8,1
<i>Pseudo-nitzschia cf.pungens (h)</i>	44	1 054	4,81	4,06	4,14	4,12	1,51	36,7	0,66	16,1
<i>Thalassionema nitzschioides (h)</i>	42	1 013	4,59	4,69	4,77	4,76	1,00	21,0	0,78	16,4
<i>Rhizosolenia imbricata</i>	-	-	-	-	-	-	-	-	-	-
<i>Ditylum brightwellii (h)</i>	40	1 114	74,17	70,10	71,00	70,88	17,30	24,4	17,01	24,0
<i>Stephanopyxis turris</i>	-	-	-	-	-	-	-	-	-	-
<i>Odontella sinensis (h)</i>	40	1 106	195,30	180,70	175,04	175,34	47,20	26,9	42,85	24,4
<i>Chaetoceros debilis (h)</i>	38	954	9,14	8,83	8,99	8,91	2,10	23,6	1,57	17,6
<i>Fragilaria crotonensis (h)</i>	49	1 291	4,40	3,76	3,48	3,48	0,98	28,2	0,30	8,6
<i>Tabellaria fenestrata (c)</i>	41	929	7,82	4,88	5,47	5,40	2,06	38,2	0,70	13,0
<i>Aulacoseira granulata</i>	-	-	-	-	-	-	-	-	-	-
<i>Cryptomonas erosa (h)</i>	49	1 113	25,85	25,75	25,91	25,91	3,64	14,1	3,53	13,6
<i>Rhodomonas lacustris</i>	-	-	-	-	-	-	-	-	-	-
<i>Planktothrix agardhii</i>	-	-	-	-	-	-	-	-	-	-
<i>Monoraphidium arcuatum</i>	-	-	-	-	-	-	-	-	-	-
<i>Woronichinia naegeliana</i>	-	-	-	-	-	-	-	-	-	-
<i>Cosmarium ocellatum (h)</i>	46	1 014	26,82	26,86	27,04	27,04	2,32	8,6	2,12	7,8
<i>Trachelomonas hispida</i>	-	-	-	-	-	-	-	-	-	-
For explanation of symbols, see Table B.1.										

Table B.4 — Performance data for biovolume analysis of phytoplankton species in natural samples

Taxon name	<i>l</i>	<i>n</i>	<i>X</i> µm ³	<i>x</i> _{med} µm ³	<i>x</i> _{mean} µm ³	<i>x</i> _{HS} µm ³	<i>s</i> _R µm ³	<i>C</i> _{V,R} %	<i>s</i> _r µm ³	<i>C</i> _{V,r} %
<i>Dactyliosolen fragilissimus</i>	45	1 225	6 824	6 911	7 045	7 033	2 566	36,5	2 497	35,5
<i>Dinophysis acuminata</i>	43	992	16 718	16 334	17 241	17 190	6 906	40,2	3 940	22,9
<i>Ceratium tripos</i>	44	998	109 160	85 983	95 007	88 516	37 516	42,4	13 216	14,9
<i>Pseudo-nitzschia cf.pungens</i>	44	1 054	1 355	1 058	1 167	1 135	541	47,7	348	30,7
<i>Thalassionema nitzschioides</i>	41	1 006	490	555	536	535	218	40,7	161	30,0
<i>Rhizosolenia imbricata</i>	42	1 154	13 980	16 089	17 031	16 988	8 001	47,1	6 982	41,1
<i>Ditylum brightwellii</i>	40	1 114	24 055	18 753	20 080	20 080	8 571	42,7	7 689	38,3
<i>Stephanopyxis turris</i>	37	964	99 642	80 331	77 611	77 611	34 081	43,9	27 694	35,7
<i>Odontella sinensis</i>	40	1 106	1 785 462	1 560 584	1 624 194	1 586 715	785 689	49,5	723 632	45,6
<i>Chaetoceros debilis</i>	39	982	1 022	854	938	937	468	49,9	328	35,0
<i>Fragilaria crotonensis</i>	50	1 311	688	526	503	498	247	49,7	85	17,1
<i>Tabellaria fenestrata</i>	45	1 009	2 111	1 648	1 863	1 758	880	50,1	463	26,3
<i>Aulacoseira granulata</i>	45	1 032	680	753	761	760	272	35,9	216	28,4
<i>Cryptomonas erosa</i>	50	1 133	1 914	1 725	1 809	1 809	655	36,2	545	30,2
<i>Rhodomonas lacustris</i>	49	1 249	102	100	103	102	33	32,4	31	30,8
<i>Planktothrix agardhii</i>	47	1 260	866	884	969	941	362	38,4	68	7,2
<i>Monoraphidium arcuatum</i>	47	1 246	64	51	57	55	27	49,5	19	35,1
<i>Woronichinia naegeliana</i>	45	1 151	24	30	31	31	12	39,7	8	25,3
<i>Cosmarium ocellatum</i>	47	1 057	5 820	5 083	5 071	5 070	1 624	32,0	987	19,5
<i>Trachelomonas hispida</i>	45	1 068	6 410	6 414	6 302	6 287	1 716	27,3	1 309	20,8

For explanation of symbols, see Table B.1.

Annex C (informative)

Carbon content calculation

Organic carbon is a main constituent of organisms. It represents the energy source transported within food webs and therefore is an important basis for ecological modelling. The carbon content of phytoplankton in natural samples cannot be determined directly (e.g. with a CN-Analyser), because chemical methods are not suitable to discriminate between the C-contents of different organism groups such as phytoplankton, zooplankton, bacteria, detritus, etc. Additionally, it will be impossible to distinguish phytoplankton groups on the taxonomic level. Therefore, a common method for estimating carbon biomass is the calculation from the microscopically determined biovolume using conversion factors. Mainly due to the correlation between total cell volume and vacuole size as mentioned before, the estimation of the carbon content with an exponential function applied on the cell volume should be preferred to the use of fixed conversion factors per volume:

$$c(C)_i = a \cdot (V_{\text{cell},i})^b \quad (\text{C.1})$$

where

$c(C)_i$ is the carbon content of taxon i in picograms per cell (pg/cell);

a and b are constant factors, see Table C.1;

$V_{\text{cell},i}$ is the average cell volume of taxon i in cubic micrometres per cell ($\mu\text{m}^3/\text{cell}$).

Because the carbon content of phytoplankton cells varies depending on different physiological and environmental factors, there are no universal values for the factors a and b . Some values used in the literature are shown in Table B.1. Due to the fact that the ratio of total cell volume to vacuole size may significantly differ from one species to another, especially for diatoms and other taxonomic groups under the same conditions, different conversion factors are used.

Table C.1 — Conversion factors for converting cell volume to carbon content from different sources

Group	Function $c(C)_i = a \cdot (V_{\text{cell},i})^b$		Source
	a	b	
pelagic diatoms	0,485	0,712	Strathmann (1967) [10]
	0,263	0,740	Tagushi (1976) [11]
	0,357	0,814	Vidal (1978) [12]
	0,513	0,757	Eppley et al. (1970) [13]
	0,131	0,890	Pavlovskaya & Kondratyeva (1981) [14]
	1,094	0,859	Blasco et al. (1982) [15]
marine dinoflagellates	0,288	0,811	Menden-Deuer & Lessard (2000) [16]
phytoplankton without diatoms	0,387	0,900	Pavlovskaya & Kondratyeva (1981) [14]
	0,347	0,866	Strathmann (1967) [10]
	0,251	0,940	Eppley et al. (1970) [13]
total phytoplankton	0,216	0,939	Menden-Deuer & Lessard (2000) [16]
	0,436	0,829	Moal et al. (1987) [17]
	0,109	0,991	Montagnes et al. (1994) [18]

The calculation of the carbon content is not mandatory, but if this procedure is carried out, then it is recommended to use the factors by Menden-Deuer & Lessard (2000) [16] or Eppley et al. (1970) [13]. Reference to what carbon factors have been used should always be included in the data report. Nevertheless, more extensive studies for different environmental conditions are needed to standardize the estimation of the carbon content of phytoplankton organisms via the biovolume.

Sometimes colony forming taxa shall be counted in cell aggregates. In these cases, it is necessary to distinguish between counting of cell packages (e.g. 100 cells of *Microcystis* sp.) and filaments (e.g. 100 µm pieces of *Nodularia* sp.). For these two special cases, the carbon content shall be calculated by using the following formulae (see also [2]):

For multi-cell colonies as

$$c(C)_i = a \cdot CPU \cdot \left(\frac{VCU}{CPU} \right)^b \quad (C.2)$$

and for filaments as

$$c(C)_i = a \cdot \frac{LCU}{CL} \cdot \left(\frac{VCU \cdot CL}{LCU} \right)^b \quad (C.3)$$

where

VCU is the volume of the counting unit in cubic micrometres (µm³);

CPU is the number of cells per counting unit;

CL is the cell length;

LCU is the length of the counting unit in micrometres (µm), mostly 100 µm.

Especially for small cells (e.g. cyanobacterial picoplankton) the carbon content can be calculated by multiplying the average cell volume with a fixed factor using a simple linear dependency (e.g. [19]):

$$c(C)_i = x \cdot V_{\text{cell},i} \quad (C.4)$$

where

c(C)_i is the carbon content of taxon *i* in picograms per cell (pg/cell);

x is a constant factor (pg/µm³);

V_{cell,i} is the average cell volume of taxon *i* in cubic micrometres per cell (µm³/cell).

Annex D (informative)

Alphabetical list of recommended geometrical shapes and hidden dimension factors

Table D.1 contains recommended geometrical shapes and additional information, which should be used for biovolume estimation. This information is generally assigned on genus level. If some species, subspecies, forms, or varieties within a genus are different, they are listed additionally. The taxon list also contains genera of typical benthic microalgae as well as some heterotrophic flagellates or other protists. Both groups often are determined in the routine monitoring in parallel to the real phytoplankton organisms (in the strict ecological sense), either because they are stirred up from the sediment into the pelagial or because they belong to taxonomical groups also containing autotrophic species (e.g. Euglenophyceae, Dinophyceae or Cryptophyceae).

Basis for the used taxonomical names is the current valid status provided by the international databases "World Register of Marine Species" (WoRMS) [20] and AlgaeBase [21]. These taxa list can not be a complete list of all phytoplankton taxa occurring in European waters but is intended as a recommendation for the most important taxa. If some taxa seem to be missing, possibly due to using synonyms, the validity of the used name has to be checked in the above mentioned databases to find the assigned geometrical shape.

For the listed taxa the geometrical shape relating to Annex A is given. Additionally it is listed to which unit (cell, filament, coenobium, etc.) the geometry has been assigned. Sometimes the allocated geometrical shape does not correctly cover the effective volume. In these cases, the result has to be multiplied by a "geometrical shape correction factor" to estimate the effective volume more precisely. Some of these factors have been taken from the Helsinki Commission (HELCOM) monitoring program [4]. All others are estimated suggestions based on the results of the mandated CEN project.

For some particular taxa, it is often not possible to measure all dimensions needed for the calculation of the cell volume during routine analysis. In the last column of Table D.1 recommendations are given for relations to measurable dimensions for most of the taxa, which usually have such a "hidden dimension" depending on the position in the microscopic view. Some of these indicated factors have also been taken from the HELCOM monitoring program [4] or derived from their size class dimensions. Others have been calculated from measurements or from several pictures from different taxonomical databases or literature.

Various taxa are quite variable in appearance. Thus, few species sometimes can be rather spherical, in other cases rather elliptical. Moreover, different stages within the life cycle (e.g. cysts) may have various forms. In addition, amoeboid taxa exist. In the following taxon-list, the assignments of a geometry have been made according to the common appearance of the vegetative form. If a corresponding taxon in the sample to be analysed shows a different shape deviating from the list, an appropriate geometrical shape will have to be selected according to Annex A.

If taxa are analysed on a higher taxonomical level than genus (e.g. family, order, class, etc.) a well-fitting shape according to Annex A will have to be assigned.

Table D.1 — Assigned geometrical shapes and correction factors to taxa (* for the explanation of the dimensions see Annex A).

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Acanthoceras</i> H.Honigmann, 1909	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Acanthochloris</i> Pascher, 1930	cell	sphere	
<i>Acanthocorbis</i> S.Hara & E.Takahashi, 1984	cell	spheroid	
<i>Acanthoeca</i> W.Ellis	cell	spheroid	
<i>Acanthogonyaulax</i> (Kofoid) Graham, 1942	cell	spheroid	
<i>Acanthoica</i> Lohmann, 1903	cell	spheroid	
<i>Acanthosphaera</i> Lemmermann, 1899	cell	sphere	
<i>Achnantheiopsis</i> Lange-Bertalot, 1997	cell	elliptic cylinder	
<i>Achnanthes</i> Bory de Saint-Vincent, 1822	cell	elliptic cylinder	$d_2=0,65 \times d_1$; $h=0,65 \times d_2$
<i>Achnanthes amoena</i> (Hustedt) Bukhtiyarova, 1999	cell	elliptic cylinder × 0,9	
<i>Achnanthes bremeyeri</i> Lange-Bertalot, 1989	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Achnanthes brevipes</i> C.Agardh, 1824	cell	elliptic cylinder	$d_2=0,35 \times d_1$; $h=1,15 \times d_2$
<i>Achnanthes caledonica</i> Lange-Bertalot	cell	cuboid × 0,9	
<i>Achnanthes lanceolata</i> subsp. <i>frequentissima</i> (Lange-Bertalot) Round & L.Bukhtiyarova, 1996	cell	elliptic cylinder × 0,9	
<i>Achnanthes longipes</i> C.Agardh, 1824	cell	elliptic cylinder	$d_2=0,25 \times d_1$; $h=1,50 \times d_2$
<i>Achnanthes taeniata</i> Grunow, 1880	cell	elliptic cylinder	$d_2=0,29 \times d_1$; $h=0,84 \times d_2$
<i>Achnanthes trinodis</i> (W.Smith) Grunow, 1880	cell	elliptic cylinder	
<i>Achnanthes ventralis</i> (Krasske) Lange-Bertalot, 1989	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Achnanthes zieglerei</i> Lange-Bertalot, 1993	cell	elliptic cylinder	
<i>Achradina</i> Lohmann, 1902	cell	spheroid	
<i>Achroonema</i> Skuja, 1948	filament	cylinder	
<i>Actidesmium</i> Reinsch, 1875	cell	spindle	
<i>Actinastrum</i> Lagerheim, 1882	cell	spindle	
<i>Actinastrum aciculare</i> Playfair	cell	cone with half sphere	
<i>Actinastrum gracillimum</i> Smith	cell	cylinder	
<i>Actinella</i> F.W.Lewis, 1864	cell	cuboid	
<i>Actiniscus</i> Ehrenberg, 1841	cell	spheroid	$h=0,80 \times d$
<i>Actinochloris</i> Korshikov, 1953	cell	spheroid	
<i>Actinocyclus</i> Ehrenberg, 1837	cell	cylinder	$h=0,50 \times d$
<i>Actinomonas</i> Kent, 1880	cell	spheroid	
<i>Actinophaenia</i> Shadbolt, 1854	cell	cylinder	$h=0,25 \times d$
<i>Actinoptychus</i> Ehrenberg, 1843	cell	cylinder	$h=0,50 \times d$
<i>Actinoptychus splendens</i> (Shadbolt) Ralfs ex Pritchard, 1861	cell	cylinder	$h=0,25 \times d$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Actinotaenium</i> (Nägeli) Teiling, 1954	cell	spheroid	
<i>Acutodesmus</i> (Hegewald) Tsarenko, 2001	cell	spindle	
<i>Adenoides</i> Balech, 1956	cell	ellipsoid	
<i>Adlafia</i> Gerd Moser, Lange-Bertalot & Metzeltin, 1998	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Adlafia aquaeductae</i> (Krasske) Gerd Moser, Lange-Bertalot & D.Metzeltin, 1998	cell	cuboid × 0,85	$c=1,00 \times b$
<i>Adlafia minuscula</i> (Grunow) Lange-Bertalot, 1999	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Adoneis</i> G.W.Andrews & P.Rivera, 1987	cell	rhombic prism	
<i>Akashiwo</i> G.Hansen & Moestrup, 2000	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Akashiwo sanguinea</i> (K.Hirasaka) G.Hansen & Ø.Moestrup, 2000	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Alexandrium</i> Halim, 1960	cell	spheroid	
<i>Alexandrium catenella</i> (Whedon & Kofoid) Balech, 1985	cell	ellipsoid	
<i>Alexandrium leei</i> Balech, 1985	cell	sphere	
<i>Alexandrium margalefii</i> Balech, 1994	cell	sphere	
<i>Algirosphaera</i> Schlauder, 1945	cell	spheroid	
<i>Algirosphaera robusta</i> (Lohmann) R.E.Norris, 1984	cell	sphere	
<i>Alisphaera</i> Heimdal, 1973	cell	sphere	
<i>Ammatoidea</i> West & G.S.West, 1897	filament	cylinder	
<i>Amoebophrya</i> Koeppen, 1894	cell	sphere	
<i>Amoenoscapa</i>	cell	spheroid	
<i>Amphicampa</i> (Ehrenberg) Ralfs, 1861	cell	elliptic cylinder × 1,03	$r_2=0,50 \times h; h=2,00 \times r_2$
<i>Amphichrysis</i> Korshikov, 1929	cell	spheroid	
<i>Amphidiniopsis</i> Woloszynska, 1928	cell	ellipsoid	
<i>Amphidinium</i> Claperède & Lachmann, 1859	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Amphidinium acutissimum</i> Schiller, 1933	cell	double cone	
<i>Amphidinium crassum</i> Lohmann, 1908	cell	ellipsoid	$d_2=0,72 \times d_1$
<i>Amphidinium longum</i> Lohmann, 1908	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Amphidinium sphenoides</i> Wülf, 1916	cell	double cone	
<i>Amphidoma</i> Stein, 1883	cell	cone with half sphere	
<i>Amphikrikos</i> Korshikov, 1953	cell	cylinder	
<i>Amphikrikos nanus</i> (Fott & Heynig) Hindák, 1977	cell	spheroid	
<i>Amphipleura</i> Kützing, 1844	cell	elliptic cylinder	
<i>Amphipleura pellucida</i> (Kützing) Kützing, 1844	cell	lanceolate cylinder	
<i>Amphiprora</i> Ehrenberg, 1843	cell	elliptic cylinder	$d_2=0,15 \times d_1; h=0,67 \times d_1$
<i>Amphirhiza</i> Skuja, 1948	cell	sphere	
<i>Amphisolenia</i> Stein, 1883	cell	spindle	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Amphora</i> Ehrenberg ex Kützing, 1844	cell	elliptic cylinder \times 0,65	$d_2=0,50 \times h$; $h=2,00 \times d_2$
<i>Amylax</i> Meunier, 1910	cell	cone with half sphere \times 0,75	$h=1,50 \times d$
<i>Anabaena</i> Bory de Saint-Vincent ex Bornet & Flahault, 1886	cell	sphere	
<i>Anabaena cylindrica</i> Lemmermann, 1896	cell	cylinder	
<i>Anabaena elliptica</i> Lemmermann, 1898	cell	spheroid	
<i>Anabaena pseudoscillatoria</i> Bory de Saint-Vincent, 1822	cell	spheroid	
<i>Anabaena smithii</i> (Komárek) M.Watanabe	cell	spheroid	
<i>Anabaena sphaerica</i> Bornet & Flahault, 1888	cell	spheroid	
<i>Anabaena tenericaulis</i> Nygaard	cell	spheroid	
<i>Anabaena utermohli</i> (Utermöhl) Geitler, 1925	cell	spheroid	
<i>Anabaenopsis</i> V.V.Miller, 1923	cell	spheroid	
<i>Anacystis</i> Meneghini, 1837	cell	spheroid	
<i>Anathece</i> (Komárek & Anagnostidis) Komárek, Kaštovský & Jezberová, 2011	cell	spheroid	
<i>Anaulus</i> Ehrenberg, 1844	cell	elliptic cylinder	$d_2=0,60 \times d_1$
<i>Aneumastus</i> D.G.Mann & A.J.Stickle, 1990	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Aneumastus tuscula</i> (Ehrenberg) D.G.Mann & A.J.Stickle, 1990	cell	elliptic cylinder \times 0,9	$h=1,00 \times d_2$
<i>Anisonema</i> Dujardin, 1841	cell	ellipsoid	
<i>Ankistrodesmus</i> Corda, 1838	cell	spindle	
<i>Ankyra</i> Fott, 1957	cell	spindle	
<i>Anomooneis</i> E.Pfitzer, 1871	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Anoplosolenia</i> Deflandre, 1952	cell	spindle	
<i>Anorthoneis</i> Grunow, 1868	cell	elliptic cylinder	
<i>Anthophysa</i> Bory de Saint-Vincent, 1822	cell	cone with half sphere	
<i>Anthosphaera</i> Kamptner, 1936	cell	sphere	
<i>Apatococcus</i> F.Brand, 1925	cell	spheroid	
<i>Apedinella</i> Thronsen, 1971	cell	sphere	
<i>Aphanizomenon</i> A.Morren ex Bornet & Flahault, 1888	filament	cylinder	
<i>Aphanocapsa</i> Nägeli, 1849	cell	sphere	
<i>Aphanothece</i> Nägeli, 1849	cell	spheroid	
<i>Aphanothece paralleliformis</i> G. Cronberg, 2003	cell	cylinder	
<i>Apiocystis</i> Nägeli, 1849	cell	sphere	
<i>Apodochloris</i> Komárek, 1959	cell	ellipsoid	
<i>Arachnochloris</i> Pascher, 1930	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Arachnoidiscus</i> H.Deane ex G.Shadbolt, 1852	cell	cylinder	
<i>Archaeoperidinium</i> Jörgensen, 1912	cell	sphere × 0,8	
<i>Arcocellulus</i> G.R.Hasle, H.A.von Stosch & E.E.Syvrtsen, 1983	cell	elliptic cylinder	
<i>Ardissonea</i> De Notaris, 1870	cell	elliptic cylinder	
<i>Arthronema</i> J.Komárek & J.Lukavský, 1988	filament	cylinder	
<i>Arthrospira</i> Sitzenberger ex Gomont, 1892	filament	cylinder	
<i>Astartiella</i> A.Witkowski, Lange-Bertalot & D.Metzeltin, 1998	cell	elliptic cylinder	
<i>Astasia</i> Dujardin, 1841	cell	ellipsoid	
<i>Asterionella</i> Hassall, 1850	cell	cuboid	$b=1,00 \times c$
<i>Asterionella ralfsii</i> var. <i>hustedtiana</i> Körner, 1970	cell	elliptic cylinder × 0,9	
<i>Asterionellopsis</i> Round, 1990	cell	pyramid × 0,9	$l_1=0,75 \times l_2$
<i>Asterococcus</i> Scherffel, 1908	cell	spheroid	
<i>Asterolampra</i> Ehrenberg, 1844	cell	cylinder	
<i>Asteromonas</i> A.Artari, 1913	cell	spheroid	
<i>Asteromphalus</i> Ehrenberg, 1844	cell	cylinder	
<i>Asteroplanus</i> C.Gardner & R.M.Crawford, 1997	cell	elliptic cylinder	$d_2=0,14 \times d_1$
<i>Astrosiga</i> W.S.Kent	cell	spheroid	
<i>Attheya</i> T.West, 1860	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Attheya longicornis</i> R.M.Crawford & C.Gardner, 1994	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Attheya septentrionalis</i> (Østrup) R.M.Crawford, 1994	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Aulacodiscus</i> C.G.Ehrenberg, 1844	cell	cylinder	$h=0,50 \times d$
<i>Aulacomonas</i> Skuja, 1939	cell	ellipsoid	
<i>Aulacoseira</i> Thwaites, 1848	cell	cylinder	
<i>Aulacoseira karelica</i> (Molder) Simonsen	cell	elliptic cylinder	
<i>Auliscus</i> Ehrenberg, 1843	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Aulomonas</i> Lackey, 1942	cell	spheroid	
<i>Aulosira</i> Kirchner ex Bornet & Flahault, 1886	cell	spheroid	
<i>Aureococcus</i> Hargraves & Sieburth, 1988	cell	sphere	
<i>Aureodinium</i> Dodge, 1967	cell	ellipsoid	
<i>Auricula</i> Castracane, 1873	cell	elliptic cylinder × 0,65	$d_2=0,50 \times h; h=2,00 \times d_2$
<i>Auxenochlorella</i> (I.Shihira & R.W.Krauss) T.Kalina & M.Puncochárová, 1987	cell	spheroid	
<i>Azadinium</i> Elbrächter & Tillmann, 2009	cell	spheroid	
<i>Azpeitia</i> M.Peragallo, 1912	cell	cylinder	
<i>Bacillaria</i> J.F.Gmelin, 1791	cell	cuboid × 0,95	$b=1,00 \times c; c=1,00 \times b$
<i>Bacteriastrium</i> Shadbolt, 1854	cell	cylinder	$h=1,00 \times d$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Bacterosira</i> Gran, 1900	cell	cylinder	
<i>Bacularia</i> Borzi, 1905	cell	spheroid	
<i>Balaniger</i> Thomsen & Oates, 1978	cell	spheroid	
<i>Balechina</i> Loeblich Jr. & Loeblich III, 1968	cell	spheroid	
<i>Bambusina</i> Kützing ex Kützing, 1849	cell	cylinder \times 0,8	
<i>Basycladia</i> W.E.Hoffmann & Tilden, 1930	filament	cylinder	
<i>Bathycoccus</i> W.Eikrem & J.Thronksen, 1990	cell	spheroid	
<i>Beggiatoa</i> V.B.A.Trevisan, 1845	filament	cylinder	
<i>Bellerochea</i> Van Heurck, 1885	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$; $h=0,35 \times l$
<i>Belonastrum</i> (Lemmermann) Round & N.I.Maidana, 2001	cell	elliptic cylinder	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Berkeleya</i> Greville, 1827	cell	elliptic cylinder	
<i>Bernardinium</i> Chodat, 1923	cell	ellipsoid	
<i>Biceratium</i> Vanhöffen, 1896	cell	cone with half sphere \times 0,65	
<i>Bicosoeca</i> H.J.Clark, 1866	cell	spheroid	
<i>Bicosta</i> B.S.C.Leadbeater, 1978	cell	spheroid	
<i>Biddulphia</i> S.F.Gray, 1821	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Biddulphia antediluviana</i> (Ehrenberg) Van Heurck, 1885	cell	cuboid \times 0,9	$b=1,00 \times a$; $c=1,40 \times a$
<i>Biddulphia biddulphiana</i> (J.E.Smith) Boyer, 1900	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Biddulphia rhombus</i> (Ehrenberg) W.Smith, 1854	cell	rhombic prism	$d_2=0,60 \times d_1$; $h=1,00 \times d_2$
<i>Biddulphiopsis</i> von Stosch & R.Simonsen, 1984	cell	elliptic cylinder	
<i>Biecheleria</i> Moestrup, Lindberg & Daugbjerg, 2009	cell	ellipsoid	
<i>Binuclearia</i> Wittrock, 1886	cell	cylinder	
<i>Biremis</i> D.G.Mann & E.J.Cox, 1990	cell	elliptic cylinder	
<i>Bitrichia</i> Woloszynska, 1914	cell	spheroid	
<i>Bleakeleya</i> Round, 1990	cell	cuboid	
<i>Blennothrix</i> Kützing ex Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Blepharocysta</i> Ehrenberg, 1873	cell	spheroid	
<i>Bodo</i> Ehrenberg, 1832	cell	spheroid	
<i>Boreadinium</i> Dodge & Hermes, 1981	cell	ellipsoid	
<i>Borghiella</i> Moestrup, G.Hansen & Daugberg, 2008	cell	ellipsoid	
<i>Borzia</i> Cohn ex Gomont, 1892	filament	cylinder	
<i>Botrydiopsis</i> Borzi, 1889	cell	sphere	
<i>Botrydium</i> Wallroth, 1815	cell	ellipsoid	
<i>Botryochloris</i> Pascher, 1930	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Botryococcus</i> Kützing, 1849	cell	spheroid	
<i>Botryosphaerella</i> P.C.Silva, 1970	cell	sphere	
<i>Braarudosphaera</i> Deflandre, 1947	cell	sphere	
<i>Brachiomonas</i> Bohlin, 1897	cell	spheroid	
<i>Brachysira</i> Kützing, 1836	cell	rhombic prism	
<i>Brachysira aponina</i> Kützing, 1836	cell	lanceolate cylinder	
<i>Brachysira brebissonii</i> R.Ross, 1986	cell	lanceolate cylinder	
<i>Brachysira calcicola</i> Lange-Bertalot, 1994	cell	elliptic cylinder	
<i>Brachysira calcicola</i> subsp. <i>pfisteri</i> H.Lange-Bertalot, 2004	cell	elliptic cylinder	
<i>Brachysira follis</i> (Ehrenberg) R.Ross, 1986	cell	elliptic cylinder × 0,8	$h=0,33 \times d_2$
<i>Brachysira hofmanniae</i> Lange-Bertalot, 1994	cell	elliptic cylinder × 0,9	
<i>Brachysira procera</i> Lange-Bertalot & Gerd Moser, 1994	cell	lanceolate cylinder	
<i>Brachysira serians</i> (Brébisson) Round & D.G.Mann, 1981	cell	lanceolate cylinder	
<i>Brachysira styriaca</i> (Grunow) R.Ross, 1986	cell	lanceolate cylinder	
<i>Brachysira vitrea</i> (Grunow) R.Ross, 1986	cell	elliptic cylinder	
<i>Brachysira zellensis</i> (Grunow) Cleve	cell	elliptic cylinder	
<i>Brebissonia</i> Grunow, 1860	cell	rhombic prism	$h=0,50 \times d_2$
<i>Brevisira</i> Krammer	cell	cylinder × 0,9	
<i>Brockmanniella</i> Hasle, Stosch & Syvertsen, 1983	cell	elliptic cylinder	$d_2=0,40 \times d_1$
<i>Bulbochaete</i> C.Agardh, 1817	filament	cylinder	
<i>Bumilleria</i> Borzì, 1889	cell	cylinder	
<i>Bumilleriopsis</i> Printz, 1914	cell	cylinder	
<i>Cachonina</i> Loeblich III, 1968	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Caecitellus</i> D.J.Patterson, K.Nygaard, G.Steinberg & Turley, 1993	cell	spheroid	
<i>Cafeteria</i> T.Fenchel & D.J.Patterson, 1988	cell	spheroid	
<i>Calcidiscus</i> Kamptner, 1950	cell	sphere	
<i>Calciopappus</i> Gaarder & Ramsfjell, 1954	cell	cone	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Calciosolenia</i> Gran, 1912	cell	spindle	
<i>Calliakantha</i> B.S.C.Leadbeater, 1978	cell	spheroid	
<i>Caloneis</i> P.Cleve, 1894	cell	elliptic cylinder	$d_2=1,50 \times h; h=0,67 \times d_2$
<i>Caloneis amphisbaena</i> (Bory de Saint Vincent) Cleve, 1894	cell	elliptic cylinder × 0,95	$h=0,75 \times d_2$
<i>Caloneis amphisbaena</i> var. <i>subsalina</i> (Donkin) Cleve, 1894	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Caloneis leptosoma</i> (Grunow) Krammer, 1985	cell	elliptic cylinder × 0,9	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Caloneis limosa</i> (Kützing) R.M.Patrick, 1966	cell	cuboid	
<i>Caloneis macedonica</i> Hustedt, 1945	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Caloneis permagna</i> (J.W.Bailey) Cleve, 1894	cell	lanceolate cylinder	
<i>Caloneis tenuis</i> (W.Gregory) Krammer in Krammer & Lange-Bertalot, 1985	cell	cuboid $\times 0,85$	
<i>Caloneis undulata</i> (Gregory) Krammer, 1985	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Calothrix</i> C.Agardh ex Bornet & Flahault, 1886	filament	cylinder	
<i>Calycomonas</i> Lohmann, 1908	cell	spheroid	
<i>Calyptrolithina</i> Heimdal, 1982	cell	spheroid	
<i>Calyptrolithophora</i> Heimdal, 1980	cell	spheroid	
<i>Calyptosphaera</i> Lohmann, 1902	cell	sphere	
<i>Camptothrix</i> West & G.S.West, 1897	filament	cylinder	
<i>Camptylonemopsis</i> T.V.Desikachary, 1948	filament	cylinder	
<i>Campylodiscus</i> Ehrenberg ex Kützing, 1844	cell	elliptic cylinder	$d_2=0,90 \times d_1; h=0,40 \times d_2$
<i>Campylomonas</i> Hill, 1991	cell	spheroid	
<i>Campyloneis</i> Grunow, 1862	cell	elliptic cylinder	
<i>Campylopyxis</i> L.K.Medlin, 1985	cell	elliptic cylinder	
<i>Campylosira</i> Grunow ex Van Heurck, 1885	cell	elliptic cylinder	$h=0,75 \times r_2$
<i>Caneosphaera</i> Gaarder, 1977	cell	sphere	
<i>Capartogramma</i> Kufferath, 1956	cell	elliptic cylinder $\times 0,9$	
<i>Capsosira</i> Kützing ex Bornet & Flahault, 1886	filament	cylinder	
<i>Carteria</i> Diesing, 1866	cell	sphere	
<i>Carteria klebsii</i> (P.A.Dangeard) Francé, 1921	cell	spheroid	
<i>Carteria marina</i> Diesing, 1866	cell	spheroid	
<i>Catacombas</i> D.M.Williams & Round, 1986	cell	cuboid $\times 0,8$	
<i>Catella</i> G.Ålvik, 1934	cell	cylinder	
<i>Catena</i> Chodat, 1900	filament	cylinder	
<i>Catenochrysis</i> Perman, 1955	cell	spheroid	
<i>Catenula</i> Mereschkowsky, 1903	cell	elliptic cylinder	
<i>Cavinula</i> D.G.Mann & A.J.Stickle, 1990	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Cavinula lacustris</i> (W.Gregory) D.G.Mann & Stickle, 1990	cell	lanceolate cylinder	$d_2=1,50 \times h; h=0,67 \times d_2$
<i>Cecidochloris</i> Skuja, 1948	cell	spheroid	
<i>Centrtractus</i> E.Lemmermann, 1900	cell	cylinder $\times 0,85$	
<i>Centrodinium</i> Kofoed, 1907	cell	cone with half sphere	
<i>Centronella</i> Max Voigt, 1902	cell part	cuboid	$c=1,00 \times b$
<i>Cerataulina</i> H.Peragallo ex Schütt, 1896	cell	cylinder	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Cerataulus</i> Ehrenberg, 1844	cell	cylinder	$h=0,67 \times d$
<i>Cerataulus smithii</i> Ralfs ex Pritchard, 1861	cell	cylinder × 0,75	$h=0,60 \times d$
<i>Ceratium</i> Schrank, 1793	cell	Ceratium - girdle diameter (see Annex A)	
<i>Ceratium brachyceros</i> Daday, 1907	cell	cone with half sphere × 0,65	
<i>Ceratium carolinianum</i> (Bailey) E.G.Jørgensen, 1911	cell	cone with half sphere × 0,65	
<i>Ceratium cornutum</i> (Ehrenberg) Claparède & J.Lachmann, 1859	cell	cone with half sphere × 0,65	
<i>Ceratium furcoides</i> (Levander) Langhans, 1925	cell	cone with half sphere × 0,65	
<i>Ceratium massiliense</i> (Gourret) Karsten, 1906	cell	cone with half sphere × 0,65	
<i>Ceratium platycorne</i> Daday, 1888	cell	cone with half sphere × 0,65	
<i>Ceratium rhomvodes</i> Hickel, 1987	cell	cone with half sphere × 0,65	
<i>Ceratocorys</i> Stein, 1883	cell	ellipsoid	
<i>Ceratolithus</i> Kamptner, 1950	cell	sphere	
<i>Cercaria</i> O.F.Müller, 1773	cell	Ceratium - girdle diameter (see Annex A)	
<i>Chaetoceros</i> Ehrenberg, 1844	cell	elliptic cylinder	$d_2=0,74 \times d_1$
<i>Chaetoceros affinis</i> Lauder, 1864	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Chaetoceros anastomosans</i> Grunow, 1882	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros anastomosans</i> var. <i>externus</i> (Gran) Cleve-Euler	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros atlanticus</i> Cleve, 1873	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros borealis</i> J.W.Bailey, 1854	cell	elliptic cylinder	$d_2=0,85 \times d_1$
<i>Chaetoceros brevis</i> F.Schütt, 1895	cell	elliptic cylinder	$d_2=0,83 \times d_1$
<i>Chaetoceros calcitrans</i> (Paulsen) Takano, 1968	cell	elliptic cylinder	$d_2=0,82 \times d_1$
<i>Chaetoceros calcitrans</i> f. <i>pumilus</i> Takano	cell	elliptic cylinder	$d_2=0,82 \times d_1$
<i>Chaetoceros castracanei</i> Karsten, 1905	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros ceratosporus</i> Ostenfeld, 1910	cell	elliptic cylinder	$d_2=0,82 \times d_1$; $h=1,60 \times d_2$
<i>Chaetoceros circinalis</i> (Meunier) K.G. Jensen & Moestrup	cell	elliptic cylinder	$d_2=0,85 \times d_1$
<i>Chaetoceros compressus</i> Lauder, 1864	cell	elliptic cylinder	$d_2=0,85 \times d_1$
<i>Chaetoceros concavicornis</i> Mangin, 1917	cell	elliptic cylinder	$d_2=0,60 \times d_1$; $h=1,75 \times d_2$
<i>Chaetoceros constrictus</i> Gran, 1897	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Chaetoceros convolutus</i> Castracane, 1886	cell	elliptic cylinder	$d_2=0,60 \times d_1$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Chaetoceros coronatus</i> Gran, 1897	cell	elliptic cylinder	$d_2=0,81 \times d_1$
<i>Chaetoceros costatus</i> Pavillard, 1911	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros crinitus</i> F.Schütt, 1895	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Chaetoceros curvisetus</i> Cleve, 1889	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Chaetoceros danicus</i> Cleve, 1889	cell	elliptic cylinder	$d_2=0,75 \times d_1$; $h=1,25 \times d_2$
<i>Chaetoceros debilis</i> Cleve, 1894	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros decipiens</i> Cleve, 1873	cell	elliptic cylinder	$d_2=0,77 \times d_1$
<i>Chaetoceros densus</i> (Cleve) Cleve, 1899	cell	elliptic cylinder	$d_2=0,63 \times d_1$
<i>Chaetoceros diadema</i> (Ehrenberg) Gran, 1897	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros didymus</i> Ehrenberg, 1845	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Chaetoceros eibenii</i> Grunow, 1882	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros externus</i> Gran, 1897	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros galvestonensis</i> Collier & Murphy	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros gracilis</i> Pantocsek, 1892	cell	elliptic cylinder	$d_2=0,80 \times d_1$; $h=1,70 \times d_2$
<i>Chaetoceros holsaticus</i> F.Schütt, 1895	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros ingolfianus</i> Ostensfeld, 1902	cell	elliptic cylinder	$d_2=0,70 \times d_1$
<i>Chaetoceros lacinosus</i> F.Schütt, 1895	cell	elliptic cylinder	$d_2=0,85 \times d_1$
<i>Chaetoceros lauderi</i> Ralfs, 1864	cell	elliptic cylinder	$d_2=0,90 \times d_1$
<i>Chaetoceros lorenzianus</i> Grunow, 1863	cell	elliptic cylinder	$d_2=0,59 \times d_1$
<i>Chaetoceros minimus</i> (Levander) D.Marino, G.Giuffre, M.Montresor & A.Zingone, 1991	cell	cylinder	
<i>Chaetoceros muelleri</i> Lemmermann, 1898	cell	elliptic cylinder	$d_2=0,86 \times d_1$; $h=1,50 \times d_2$
<i>Chaetoceros neogracile</i> S.L.VanLandingham, 1968	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros orientale</i> Schiller	cell	elliptic cylinder	$d_2=0,55 \times d_1$
<i>Chaetoceros perpusillus</i> Cleve, 1897	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros peruvianus</i> Brightwell, 1856	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros protuberans</i> H.S.Lauder, 1864	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Chaetoceros pseudocrinitus</i> Ostensfeld, 1901	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros radicans</i> F.Schütt, 1895	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros salsugineus</i> Takano, 1983	cell	elliptic cylinder	$d_2=0,81 \times d_1$
<i>Chaetoceros seiracanthus</i> Gran, 1897	cell	elliptic cylinder	$d_2=0,76 \times d_1$
<i>Chaetoceros seriacanthus</i> Gran	cell	elliptic cylinder	$d_2=0,75 \times d_1$
<i>Chaetoceros similis</i> Cleve, 1896	cell	elliptic cylinder	$d_2=0,70 \times d_1$
<i>Chaetoceros simplex</i> Ostensfeld, 1901	cell	elliptic cylinder	$d_2=0,85 \times d_1$; $h=1,60 \times d_2$
<i>Chaetoceros socialis</i> H.S.Lauder, 1864	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Chaetoceros socialis</i> f. <i>radians</i> (F.Schütt) A.I.Proshkina-Lavrenko, 1963	cell	elliptic cylinder	$d_2=0,50 \times d_1$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Chaetoceros subsecundus</i> (Grunow ex Van Heurck) Hustedt, 1927	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetoceros subtilis</i> Cleve, 1896	cell	elliptic cylinder	$d_2=0,85 \times d_1$
<i>Chaetoceros tenuissimus</i> Meunier, 1913	cell	elliptic cylinder	$d_2=0,87 \times d_1$; $h=1,40 \times d_2$
<i>Chaetoceros teres</i> Cleve, 1896	cell	elliptic cylinder	$d_2=0,85 \times d_1$
<i>Chaetoceros thronsenii</i> (Marino, Montresor & Zingone) Marino, Montresor & Zingone, 1991	cell	elliptic cylinder	$d_2=0,87 \times d_1$
<i>Chaetoceros tortissimus</i> Gran, 1900	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Chaetoceros willei</i> Grunow, 1897	cell	elliptic cylinder	$d_2=0,67 \times d_1$
<i>Chaetonema</i> Filipjev, 1927	filament	cylinder	
<i>Chaetopeltis</i> Berthold, 1878	cell	spheroid	
<i>Chaetophora</i> Schrank, 1783	filament	cylinder	
<i>Chaetosphaeridium</i> Klebahn, 1892	cell	sphere	
<i>Chamaecalyx</i> Komárek & Anagnostidis, 1986	cell group	spheroid	
<i>Chamaepinnularia</i> Lange-Bertalot & Krammer, 1996	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Chamaepinnularia krookiformis</i> (K.Krammer) Lange-Bertalot & K.Krammer in H.Lange-Bertalot & S.I.Genkal, 1999	cell	cuboid × 0,9	$b=1,11 \times c$; $c=0,90 \times b$
<i>Chamaepinnularia witkowskii</i> (H.Lange-Bertalot & D.Metzeltin) M.Kulikovskiy & H.Lange-Bertalot, 2010	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Chamaesiphon</i> A.Braun, 1864	filament	cylinder	
<i>Characiopsis</i> Borzi, 1895	cell	spindle	
<i>Characium</i> Braun ex Kützing, 1849	cell	spindle	
<i>Chattonella</i> B.Biecheler, 1936	cell	spheroid	
<i>Chilomonas</i> Ehrenberg ex Ralfs, 1831	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Chlainomonas</i> H.R.Christen, 1959	cell	spheroid	
<i>Chlamydocapsa</i> Fott, 1972	cell	spheroid	
<i>Chlamydocapsa planctonica</i> (West & G.S.West) Fott	cell	sphere	
<i>Chlamydomonas</i> Ehrenberg, 1833	cell	spheroid	
<i>Chlamydomonas gloeocystiformis</i> (O.Dill) A.Nakazawa, 2001	cell	sphere	
<i>Chlamydomonas tetragama</i> (Bohlin) Wille, 1909	cell	spindle	
<i>Chlorallanthus</i> Pascher, 1930	cell	ellipsoid	
<i>Chlorangiella</i> De Toni, 1889	cell	ellipsoid	
<i>Chlorangiopsis</i> Korshikov, 1932	cell	spheroid	
<i>Chlorella</i> M.Beijerinck, 1890	cell	spheroid	
<i>Chlorella vulgaris</i> Beyerinck [Beijerinck], 1890	cell	sphere	
<i>Chlorhormidium</i> B.Fott, 1960	filament	cylinder	
<i>Chloridella</i> Pascher, 1932	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Chlorobotrys</i> Bohlin, 1901	cell	sphere	
<i>Chlorochytrium</i> Cohn, 1872	cell	spheroid	
<i>Chlorocloster</i> Pascher, 1925	cell	spheroid	
<i>Chlorococcum</i> Meneghini, 1842	cell	spheroid	
<i>Chlorococcum wimmeri</i> (F.W.Hilse) Rabenhorst, 1868	cell	sphere	
<i>Chlorodesmos</i> F.W.Phillips, 1882	cell	spheroid	
<i>Chlorogibba</i> Geitler, 1928	cell	sphere	
<i>Chlorogloea</i> Wille, 1900	cell	spheroid	
<i>Chlorogonium</i> Ehrenberg, 1837	cell	spindle	
<i>Chloroidium</i> Nadson, 1906	cell	spheroid	
<i>Chlorolobion</i> Korshikov, 1953	cell	spindle	
<i>Chloromonas</i> Gobi, 1899-1900	cell	spheroid	
<i>Chlorotetraedron</i> F.J.MacEntee, H.C.Bold & P.A.Archibald, 1978	cell	cuboid	$c=0,67 \times b$
<i>Chlorothecium</i> Borzi, 1885	cell	spindle	
<i>Chlorotylum</i> Kützing, 1843	cell	cylinder	
<i>Chodatella</i> Lemmermann, 1898	cell	spheroid	
<i>Chondrocystis</i> Lemmermann, 1899	cell	sphere	
<i>Choricystis</i> (Skuja) Fott, 1976	cell	spheroid	
<i>Chromophysomonas</i> H.R.Preisig & D.J.Hibberd, 1982	cell	sphere	
<i>Chromulina</i> L.Cienkowsky, 1870	cell	spheroid	
<i>Chroococciopsis</i> Geitler, 1933	cell	sphere	
<i>Chroococcidium</i> Geitler, 1933	cell	spheroid	
<i>Chroococcopsis</i> Geitler, 1925	cell	spheroid	
<i>Chroococcus</i> Nägeli, 1849	cell	spheroid	
<i>Chroococcus aphanocapsoides</i> Skuja, 1964	cell	sphere	
<i>Chroococcus cumulatus</i> Bachmann, 1921	cell	sphere	
<i>Chroococcus dispersus</i> (Keissler) Lemmermann, 1904	cell	sphere	
<i>Chroococcus distans</i> (G.M.Smith) Komárková-Legnerová & Cronberg, 1993	cell	sphere	
<i>Chroococcus microscopicus</i> J.Komárková-Legnerová & G.Cronberg, 1994	cell	sphere	
<i>Chroococcus minimus</i> (Keissler) Lemmermann, 1904	cell	sphere	
<i>Chroococcus minor</i> (Kützing) Nägeli, 1849	cell	sphere	
<i>Chroococcus minutus</i> (Kützing) Nägeli, 1849	cell	sphere	
<i>Chroococcus turgidus</i> (Kützing) Nägeli, 1849	cell	sphere	
<i>Chroomonas</i> Hansgirg, 1885	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Chroomonas caudata</i> (L.Geitler) D.R.A.Hill, 1991	cell	cone with half sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Chrysamoeba</i> G.A.Klebs, 1892	cell	sphere	
<i>Chrysanthemodiscus</i> A.Mann, 1925	cell	cylinder × 0,9	
<i>Chrysopsis</i> Pascher, 1910	cell	spheroid	
<i>Chrysastrella</i> Chodat, 1922	cell	sphere	
<i>Chrysidiastrum</i> Lauterborn, 1913	cell	spheroid	
<i>Chrysocapsa</i> Pascher, 1913	cell	sphere	
<i>Chrysocapsella</i> Bourrelly, 1957	cell	spheroid	
<i>Chrysochromulina</i> Lackey, 1939	cell	spheroid	
<i>Chrysochromulina parva</i> Lackey, 1939	cell	ellipsoid	$d_2=0,60 \times d_1$
<i>Chrysococcus</i> G.A.Klebs, 1892	cell	sphere	
<i>Chrysodendron</i> Pascher, 1927	cell	spheroid	
<i>Chrysolykos</i> B.Mack, 1951	cell	spheroid	
<i>Chrysomonas</i> F.Stein, 1878	cell	sphere	
<i>Chrysopora</i> Pascher, 1925	cell	spheroid	
<i>Chrysopyxis</i> F.Stein, 1878	cell	sphere	
<i>Chrysosaccus</i> Pascher, 1925	cell	sphere	
<i>Chrysosphaera</i> Pascher, 1914	cell	spheroid	
<i>Chrysosphaerella</i> Lauterborn, 1896	cell	spheroid	
<i>Chrysostephanosphaera</i> Scherffel, 1911	cell	sphere	
<i>Chrysotilos</i> Pascher, 1931	cell	spheroid	
<i>Chrysoxys</i> Skuja, 1948	cell	spheroid	
<i>Ciliophrys</i> Cienkowski, 1876	cell	sphere	
<i>Citharistes</i> Stein, 1883	cell	ellipsoid	
<i>Cladopyxis</i> Stein, 1883	cell	sphere	
<i>Clastidium</i> Kirchner, 1880	cell	cylinder	
<i>Clastidium setigerum</i> Kirchner, 1880	cell	spindle	
<i>Climacodium</i> Grunow, 1868	cell	elliptic cylinder	
<i>Closteriopsis</i> Lemmermann, 1899	cell	spindle	
<i>Closterium</i> Nitzsch ex Ralfs, 1848	cell	spindle	
<i>Closterium closterioides</i> var. <i>intermedium</i> (J.Roy & Bisset) Ruzicka, 1973	cell	spindle × 1,2	
<i>Closterium directum</i> W.Archer, 1862	cell	cylinder	
<i>Closterium navicula</i> (Brébisson) Lütkemüller, 1905	cell	spindle × 1,2	
<i>Closterium praelongum</i> Brébisson, 1856	cell	cylinder	
<i>Closterium praelongum</i> var. <i>brevius</i> (Nordstedt) Willi Krieger, 1935	cell	cylinder	
<i>Coccolithus</i> E.H.L.Schwarz, 1894	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Coccomonas</i> Stein, 1878	cell	spheroid	
<i>Coccomyxa</i> Léger & Hesse, 1907	cell	spheroid	
<i>Cocconeis</i> Ehrenberg, 1837	cell	elliptic cylinder	$h=0,35 \times d_2$
<i>Cocconeis pediculus</i> Ehrenberg, 1838	cell	elliptic cylinder	$h=0,33 \times d_2$
<i>Cocconeis placentula</i> var. <i>placentula</i> Ehrenberg	cell	elliptic cylinder	$h=0,30 \times d_2$
<i>Cocconeis scutellum</i> var. <i>scutellum</i> Ehrenberg, 1838	cell	elliptic cylinder	$h=0,10 \times d_2$
<i>Coccolopia</i> Troickaja, 1922	cell	spheroid	
<i>Cochlodinium</i> Schütt, 1896	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Cochlodinium achromaticum</i> Lebour, 1925	cell	spheroid	
<i>Cochlodinium brandtii</i> Wulff, 1916	cell	spheroid	
<i>Cochlodinium pulchellum</i> Lebour, 1917	cell	spindle	
<i>Cochlodinium vinctum</i> Kofoid & Swezy, 1921	cell	spheroid	
<i>Codomonas</i> Lackey, 1939	cell	spheroid	
<i>Codonomonas</i> van Goor, 1925	cell	spheroid	
<i>Codonosigopsis</i> (Rabin) Senn	cell	spheroid	
<i>Codosiga</i> H.J.Clark, 1866	cell	spheroid	
<i>Coelastrum</i> Nägeli, 1849	cell	sphere	
<i>Coelastrum pseudomicroporum</i> Korshikov, 1953	cell	spheroid	
<i>Coelomoron</i> Buell, 1938	cell	spheroid	
<i>Coelosphaerium</i> Nägeli, 1849	cell	sphere	
<i>Coenochloris</i> Korshikov, 1953	cell	sphere	
<i>Coenococcus</i> Korshikov, 1953	cell	sphere	
<i>Coenocystis</i> Korshikov, 1953	cell	spheroid	
<i>Colacium</i> Ehrenberg, 1834	cell	spheroid	
<i>Coleochaete</i> Brébisson, 1844	colony	cylinder	
<i>Coleochlamys</i> Korshikov, 1953	cell	spheroid	
<i>Coleodesmium</i> Borzi ex Geitler, 1942	filament	cylinder	
<i>Coleofasciculus</i> Siegesmund, J.R.Johansen & Friedl, 2008	filament	cylinder	
<i>Collodictyon</i> H.J.Carter, 1865	cell	spheroid	
<i>Comasiella</i> E.Hegewald, M.Wolf, A.Keller, Friedl & Krienitz, 2010	cell	spheroid	
<i>Commation</i> Thomsen & Larsen, 1993	cell	cone	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Conradiella</i> Pascher, 1925	cell	spheroid	
<i>Conradocystis</i> A.Hollande, 1952	cell	spheroid	
<i>Conticribra</i> K.Stachura-Suchoples & D.M.Williams, 2009	cell	cylinder	$h=0,80 \times d$
<i>Coolia</i> Meunier, 1919	cell	ellipsoid	$d_2=0,90 \times d_1$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Corethron</i> Castracane, 1886	cell	cylinder \times 0,86	
<i>Corisphaera</i> Kamptner, 1936	cell	sphere	
<i>Coronastrum</i> R.H.Thompson, 1938	cell	spheroid	
<i>Coronosphaera</i> Gaarder, 1977	cell	sphere	
<i>Corymbellus</i> J.C.Green, 1976	cell	spheroid	
<i>Corythodinium</i> Loeblich Jr. & Loeblich III, 1966	cell	spindle	
<i>Coscinodiscus</i> Ehrenberg, 1839	cell	cylinder	$h=0,52 \times d$
<i>Coscinodiscus asteromphalus</i> Ehrenberg, 1844	cell	cylinder	$h=0,47 \times d$
<i>Coscinodiscus centralis</i> Ehrenberg, 1844	cell	cylinder	$h=0,30 \times d$
<i>Coscinodiscus commutatus</i> Grunow, 1884	cell	cylinder	$h=0,48 \times d$
<i>Coscinodiscus concinnus</i> W.Smith, 1856	cell	cylinder	$h=0,39 \times d$
<i>Coscinodiscus jonesianus</i> (Greville) Ostenfeld, 1915	cell	cylinder \times 0,9	
<i>Coscinodiscus radiatus</i> Ehrenberg, 1840	cell	cylinder	$h=0,40 \times d$
<i>Coscinodiscus wailesii</i> Gran & Angst, 1931	cell	cylinder	$h=0,71 \times d$
<i>Coscosira</i> Gran, 1900	cell	cylinder	$h=0,38 \times d$
<i>Cosmarium</i> Corda ex Ralfs, 1848	half-cell	ellipsoid	$d_2=0,60 \times d_1$
<i>Cosmioneis</i> D.G.Mann & Stickle, 1990	cell	elliptic cylinder \times 0,9	$h=0,75 \times d_2$
<i>Cosmocladium</i> Brébisson, 1856	half-cell	spheroid	
<i>Cosmoeca</i> Thomsen, 1984	cell	spheroid	
<i>Craspedostauros</i> E.J.Cox, 1999	cell	elliptic cylinder	
<i>Craticula</i> Grunow, 1867	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Craticula accomoda</i> (Hustedt) D.G.Mann, 1990	cell	elliptic cylinder \times 0,9	
<i>Craticula acidoclinata</i> Lange-Bertalot & Metzeltin, 1996	cell	rhombic prism	$h=0,75 \times d_2$
<i>Craticula ambigua</i> (Ehrenberg) D.G.Mann, 1990	cell	elliptic cylinder \times 0,9	$h=0,75 \times d_2$
<i>Craticula buderi</i> (Hustedt) Lange-Bertalot, 2000	cell	lanceolate cylinder \times 0,9	
<i>Craticula cuspidata</i> (Kützing) D.G.Mann, 1990	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Craticula dissociata</i> (E.Reichardt) E.Reichardt, 1997	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Craticula halophila</i> (Grunow) D.G.Mann, 1990	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Craticula minusculoides</i> (Hustedt) Lange-Bertalot, 2001	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Craticula molestiformis</i> (Hustedt) Mayama, 1999	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Craticula riparia</i> (Hustedt) Lange-Bertalot, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Craticula submolesta</i> (Hustedt) Lange-Bertalot, 1996	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Crenalithus</i> P.H.Roth, 1973	cell	sphere	
<i>Cricosphaera</i> Braarud, 1960	cell	sphere	
<i>Crinalium</i> Crow, 1927	filament	cylinder	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Crinolina</i> H.A.Thomsen, 1976	cell	spheroid	
<i>Crucigenia</i> Morren, 1830	cell	spheroid	
<i>Crucigenia fenestrata</i> (Schmidle) Schmidle, 1900	cell	trapezoid prism	$l_2=0,6 \times l_1; h=1,00 \times l_1$
<i>Crucigenia lauterbornii</i> (Schmidle) Schmidle, 1900	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Crucigenia mucronata</i> (G.M.Smith) J.Komárek	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Crucigenia quadrata</i> Morren, 1830	cell	sphere	
<i>Crucigenia quadrata</i> var. <i>secta</i> Playfair	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Crucigenia tetrapedia</i> (Kirchner) Kuntze, 1898	cell	tetrahedron	
<i>Crucioplacolithus</i> W.Hay & Mohler, 1967	cell	sphere	
<i>Crucispina</i> Espeland & Thronsdén, 1986	cell	spheroid	
<i>Cryothecomonas</i> Thomsen Buck, Bolt & Garrison, 1991	cell	spheroid	
<i>Cryptaulax</i> Skuja, 1948	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Crypthecodinium</i> Biecheler, 1938	cell	spheroid	
<i>Cryptoglana</i> Ehrenberg, 1831	cell	spheroid	
<i>Cryptoglana skujae</i> Marin & Melkonian, 2003	cell	ellipsoid	$d_2=0,85 \times d_1$
<i>Cryptomonas</i> Ehrenberg, 1831	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Crystallolithus</i> Gaarder & Markali, 1957	cell	sphere	
<i>Ctenophora</i> (Grunow) D.M.Williams & Round, 1986	cell	elliptic cylinder	$d_2=0,15 \times d_1; h=1,00 \times d_2$
<i>Cuspidothrix</i> Rajaniemi, Komárek, Willame, Hrouzek, Ka, 2005	filament	cylinder	
<i>Cyanarcus</i> Pascher, 1914	filament	elliptic cylinder	
<i>Cyanobacterium</i> Rippka & Cohen-Bazire, 1983	cell	spheroid	
<i>Cyanobium</i> Rippka & Cohen-Bazire, 1983	cell	spheroid	
<i>Cyanocatena</i> Hindák, 1975	cell	cylinder	
<i>Cyanocystis</i> Borzi, 1882	cell	spheroid	
<i>Cyanodermatium</i> Geitler, 1933	cell	spheroid	
<i>Cyanodictyon</i> Pascher, 1914	cell	sphere	
<i>Cyanodictyon balticum</i> Cronberg	cell	cylinder	
<i>Cyanodictyon iac</i> G.Cronberg & J.Komárek, 1994	cell	spheroid	
<i>Cyanodictyon planctonicum</i> B.A.Mayer, 1994	cell	spheroid	
<i>Cyanodictyon tubiforme</i> Cronberg, 1988	cell	spheroid	
<i>Cyanogranis</i> Hindák, 1982	cell	sphere	
<i>Cyanokybus</i> Schiller, 1956	cell	spheroid	
<i>Cyanonephron</i> Hickel, 1985	cell	spheroid	
<i>Cyanophanon</i> Geitler, 1955	filament	cylinder	
<i>Cyanosaccus</i> Lukas & Golubic, 1981	cell	spheroid	
<i>Cyanosarcina</i> Kováčik, 1988	cell	spheroid	

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<i>Cyanostylon</i> Geitler, 1928	cell	spheroid	
<i>Cyanothece</i> Komárek, 1976	cell	spheroid	
<i>Cyathomonas</i> Fromentel, 1874	cell	ellipsoid	
<i>Cyclidiopsis</i> Korshikov, 1917	cell	spindle	
<i>Cyclonexis</i> A.Stokes, 1886	cell	spheroid	
<i>Cyclophora</i> Castracane, 1878	cell	elliptic cylinder	
<i>Cyclostephanos</i> Round ex Theriot, Hakansson, Kociolek, Round & Stoermer, 1987	cell	cylinder	$h=0,60 \times d$
<i>Cyclotella</i> (Kützing) Brébisson, 1838	cell	cylinder	$h=0,61 \times d$
<i>Cyclotella atomus</i> Hustedt, 1937	cell	cylinder	$h=0,50 \times d$
<i>Cyclotella austriaca</i> (M.Peragallo, Handmann & Schiedler) Hustedt, 1948	cell	elliptic cylinder	
<i>Cyclotella choctawhatcheeana</i> Prasad, 1990	cell	cylinder	$h=0,62 \times d$
<i>Cyclotella meneghiniana</i> Kützing, 1844	cell	cylinder	$h=0,86 \times d$
<i>Cyclotella radiosa</i> (Grunow) Lemmermann, 1900	cell	cylinder	$h=0,47 \times d$
<i>Cylindrocapsa</i> Reinsch, 1867	cell	cylinder	
<i>Cylindrocystis</i> Meneghini ex De Bary, 1858	cell	cylinder $\times 0,88$	
<i>Cylindrospermopsis</i> G.Seenayya & N.Subba Raju, 1972	filament	cylinder	
<i>Cylindrospermum</i> Kützing ex Bornet & Flahault, 1886	cell	cylinder	
<i>Cylindrotheca</i> Rabenhorst, 1859	cell	spindle	
<i>Cymatoneis</i> Cleve, 1894	cell	rhombic prism	
<i>Cymatopleura</i> W.Smith, 1851	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Cymatopleura elliptica</i> (Brébisson) W.Smith, 1851	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Cymatopleura elliptica</i> var. <i>hibernica</i> (W.Smith) Hustedt	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Cymatosira</i> Grunow, 1862	cell	elliptic cylinder	$d_2=0,33 \times d_1; h=1,00 \times d_2$
<i>Cymbella</i> C.Agardh, 1830	cell	elliptic cylinder $\times 0,8$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella cymbiformis</i> C.Agardh, 1830	cell	lanceolate cylinder $\times 0,8$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella excisa</i> Kützing, 1844	cell	lanceolate cylinder $\times 0,8$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella excisa</i> var. <i>procera</i> Krammer, 2002	cell	lanceolate cylinder $\times 0,8$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella excisiformis</i> Krammer, 2002	cell	lanceolate cylinder $\times 0,8$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella laevis</i> Nägeli, 1849	cell	lanceolate cylinder $\times 0,8$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella lanceolata</i> (C.Agardh) C.Agardh, 1830	cell	lanceolate cylinder $\times 0,65$	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Cymbella lancettula</i> (Krammer) Krammer, 2002	cell	lanceolate cylinder \times	$d_2=0,83 \times h; h=1,20 \times d_2$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
		0,8	
<i>Cymbella lange-bertalotii</i> Krammer, 2002	cell	lanceolate cylinder \times 0,8	$d_2=0,83 \times h$; $h=1,20 \times d_2$
<i>Cymbella parva</i> (W.Smith) Kirchner, 1878	cell	lanceolate cylinder \times 0,8	$d_2=0,83 \times h$; $h=1,20 \times d_2$
<i>Cymbella subhelvetica</i> Krammer, 2002	cell	lanceolate cylinder \times 0,8	$d_2=0,83 \times h$; $h=1,20 \times d_2$
<i>Cymbellafalsa</i> H.Lange-Bertalot & D.Metzeltin, 2009	cell	elliptic cylinder	
<i>Cymbellonitzschia</i> Hustedt, 1924	cell	elliptic cylinder	
<i>Cymbodinium</i> J.Cachon & M.Cachon, 1967	cell	sphere	
<i>Cymbomonas</i> J.Schiller, 1913	cell	spheroid	
<i>Cymbopleura</i> (K.Krammer) K.Krammer, 1999	cell	elliptic cylinder	
<i>Cymbopleura amphicephala</i> (Nägeli) Krammer, 2003	cell	elliptic cylinder \times 0,9	
<i>Cymbopleura anglica</i> (Lagerstedt) Krammer, 2003	cell	elliptic cylinder \times 0,9	
<i>Cymbopleura angustata</i> (W.Smith) Krammer, 2003	cell	elliptic cylinder \times 0,9	
<i>Cymbopleura apiculata</i> Krammer, 2003	cell	elliptic cylinder \times 0,9	
<i>Cymbopleura cuspidata</i> (Kützing) Krammer, 2003	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h$; $h=1,20 \times d_2$
<i>Cyrtosphaera</i> A.Kleijne, 1992	cell	spheroid	
<i>Cystodinedria</i> Pascher, 1944	cell	ellipsoid	
<i>Cystodinium</i> Klebs, 1912	cell	ellipsoid	
<i>Cystomonas</i> Ettl & Gaertner, 1987	cell	ellipsoid	
<i>Cystopleura</i> Brébisson ex O.Kuntze, 1891	cell	elliptic cylinder \times 0,65	$d_2=0,83 \times h$; $h=1,20 \times d_2$
<i>Dactyliosolen</i> Castracane, 1886	cell	cylinder	
<i>Dactylococcopsis</i> Hansgirg	cell	spindle	
<i>Dactylosphaerium</i> Steinecke, 1916	cell	sphere	
<i>Daktylethra</i> Gartner, 1969	cell	spheroid	
<i>Dasygloea</i> Thwaites ex Gomont, 1892	filament	cylinder	
<i>Decussata</i> (Patrick) Lange-Bertalot & Metzeltin, 2000	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Delicata</i> K.Krammer, 2003	cell	elliptic cylinder	
<i>Delphineis</i> G.W.Andrews, 1977	cell	elliptic cylinder	$d_2=0,40 \times d_1$; $h=0,60 \times d_2$
<i>Dendromonas</i> F.Stein, 1878	cell	spheroid	
<i>Denticella</i> C.G.Ehrenberg, 1838	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Denticula</i> Kützing, 1844	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Derepyxis</i> A.Stokes, 1885	cell	spheroid	
<i>Dermocarpella</i> Lemmermann, 1907	cell	sphere	
<i>Desmarella</i> W.S.Kent, 1878	cell	spheroid	
<i>Desmattractum</i> West & G.S West, 1902	cell	spheroid	
<i>Desmidium</i> C.Agardh ex Ralfs, 1848	cell	cylinder	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Desmidium swartzii</i> C.Agardh ex Ralfs, 1848	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$;
<i>Desmidium swartzii</i> var. <i>amblyodon</i> (Itzigsohn) Rabenhorst, 1863	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$;
<i>Desmidium swartzii</i> var. <i>gotlandicum</i> R.L.Grönblad	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$;
<i>Desmococcus</i> F.Brand, 1925	cell	sphere	
<i>Desmodesmus</i> (R.Chodat) S.S.An, T.Friedl & E.Hegewald, 1999	cell	spheroid	
<i>Desmomastix</i> Pascher, 1914	cell	ellipsoid	
<i>Desmosiphon</i> Borzì, 1907	filament	cylinder	
<i>Detonula</i> F.Schütt ex De Toni, 1894	cell	cylinder	$h=0,80 \times d$
<i>Detonula confervacea</i> (Cleve) Gran, 1900	cell	cylinder	
<i>Deutschlandia</i> Lohmann, 1912	cell	sphere	
<i>Diachros</i> Pascher, 1937	cell	sphere	
<i>Diacronema</i> Prauser, 1958	cell	spheroid	
<i>Diadesmis</i> Kützing, 1844	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Diadesmis biceps</i> G.A.Arnott	cell	cuboid × 0,85	
<i>Diadesmis contenta</i> (Grunow ex Van Heurck) D.G.Mann, 1990	cell	cuboid × 0,95	$c=1,00 \times b$
<i>Diadesmis contenta</i> var. <i>biceps</i> (Grunow) P.B.Hamilton, 1992	cell	cuboid	
<i>Diaphanoeca</i> W.Ellis, 1930	cell	spheroid	
<i>Diatoma</i> Bory de Saint-Vincent, 1824	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Diatoma constricta</i> (Grunow) Williams	cell	cuboid	$b=1,00 \times c$; $c=1,00 \times b$
<i>Diatoma gracillima</i> Hantzsch, 1861	cell	cuboid	$b=1,00 \times c$; $c=1,00 \times b$
<i>Diatoma hiemalis</i> (Roth) Heiberg, 1863	cell	cuboid	
<i>Diatoma moniliformis</i> Kützing, 1833	cell	cuboid × 0,9	
<i>Diatoma tenuis</i> C.Agardh, 1812	cell	cuboid	$b=1,00 \times c$; $c=1,00 \times b$
<i>Diatoma vulgaris</i> Bory de Saint-Vincent, 1824	cell	elliptic cylinder	$d_2=0,25 \times d_1$; $h=0,50 \times d_1$
<i>Diatomella</i> Greville, 1855	cell	elliptic cylinder	
<i>Dicellula</i> Svirenko, 1926	cell	spheroid	
<i>Dichothrix</i> Zanardini ex Bornet & Flahault, 1886	filament	cylinder	
<i>Dichotomococcus</i> Korshikov, 1928	cell	cone with half sphere	
<i>Dickieia</i> Berkeley ex Kützing, 1844	cell	elliptic cylinder	
<i>Dicloster</i> C.-C.Jao, Y.S.Weï & H.C.Hu, 1976	cell	spindle	
<i>Dicranochaete</i> Hieronymus, 1890	filament	cylinder	
<i>Dicrateria</i> Parke, 1949	cell	sphere	
<i>Dicroerisma</i> F.J.R.Taylor & S.A.Cattell, 1970	cell	cone × 0,5	$z=\sqrt{(s^2-0,25 \times d^2)}$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Dictyocha</i> Ehrenberg, 1837	cell	sphere $\times 0,5$	
<i>Dictyochlorella</i> P.C.Silva, 1959	cell	spheroid	
<i>Dictyococcus</i> Gerneck, 1907	cell	sphere	
<i>Dictyoneis</i> Cleve, 1890	cell	elliptic cylinder	
<i>Dictyosphaerium</i> Nägeli, 1849	cell	spheroid	
<i>Didymocystis</i> Korshikov, 1953	cell	spheroid	
<i>Didymoeca</i> Doweld, 2003	cell	spheroid	
<i>Didymogenes</i> Schmidle, 1905	cell	spheroid	
<i>Didymosphenia</i> M.Schmidt, 1899	cell	elliptic cylinder $\times 0,9$	
<i>Dimeregramma</i> Ralfs, 1861	cell	elliptic cylinder	$d_2=0,45 \times d_1$; $h=1,00 \times d_2$
<i>Dimorphococcus</i> Braun, 1855	cell	spheroid	
<i>Dinema</i> Perty, 1852	cell	spheroid	
<i>Dinematomonas</i> P.C.Silva, 1960	cell	spheroid	
<i>Dinobryon</i> Ehrenberg, 1834	cell	spheroid	
<i>Dinococcus</i> Fott, 1960	cell	ellipsoid	
<i>Dinophysis</i> Ehrenberg, 1839	cell	ellipsoid	$d_2=0,45 \times d_1$
<i>Dinophysis acuminata</i> Claparède & Lachmann, 1859	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Dinophysis acuta</i> Ehrenberg, 1839	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Dinophysis caudata</i> Saville-Kent, 1881	cell	ellipsoid $\times 1,05$	$d_2=0,60 \times d_1$
<i>Dinophysis contracta</i> (Kofoid & Skogsberg) Balech, 1973	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis dens</i> Pavillard, 1915	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis groenlandica</i> (Schiller) Balech	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis hastata</i> Stein, 1883	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis mucronata</i> (Kofoid & Skogsberg) Sournia, 1973	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis nasuta</i> (Stein) Parke & Dixon, 1968	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis norvegica</i> Claparède & Lachmann, 1859	cell	ellipsoid	$d_2=0,49 \times d_1$
<i>Dinophysis odiosa</i> (Pavillard) Tai & Skogsberg, 1934	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Dinophysis ovata</i> Claparède & Lachmann, 1859	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis pulchella</i> (Lebour) Balech, 1967	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis punctata</i> Jørgensen, 1923	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis recurva</i> Kofoid & Skogsberg, 1928	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis ruudii</i> (Braarud) Balech, 1967	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Dinophysis tripos</i> Gourret, 1883	cell	ellipsoid	$d_2=0,60 \times d_1$
<i>Dinoporella</i> Halim, 1960	cell	spheroid	$h=0,60 \times d$
<i>Dinosphaera</i> Kofoid & Michener, 1912	cell	ellipsoid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Diplochlois</i> Korshikov, 1939	cell	spindle	
<i>Diplochlois hortobagyi</i> B.Fott	cell	spheroid	
<i>Diplochlois lunata</i> (Fott) Fott, 1979	cell	spheroid	
<i>Diploeca</i> W.N.Ellis	cell	spheroid	
<i>Diplomenora</i> K.L.Blazé, 1984	cell	elliptic cylinder	
<i>Diplomitella</i> P.C.Silva, 1960	cell	spheroid	
<i>Diploneis</i> Ehrenberg ex Cleve, 1894	cell	elliptic cylinder	$d_2=0,45 \times d_1$; $h=0,60 \times d_2$
<i>Diploneis crabro</i> (Ehrenberg) Ehrenberg, 1854	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Diploneis didyma</i> (Ehrenberg) Ehrenberg, 1845	cell	elliptic cylinder	$h=0,63 \times d_2$
<i>Diploneis elliptica</i> (Kützing) Cleve, 1894	cell	elliptic cylinder	$d_2=0,57 \times d_1$; $h=0,70 \times d_2$
<i>Diploneis incurvata</i> (Gregory) Cleve, 1894	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Diploneis interrupta</i> (Kützing) Cleve, 1894	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Diplopelta</i> Stein ex Jörgensen, 1912	cell	spheroid	$h=0,80 \times d$
<i>Diplopsalis</i> Bergh, 1881	cell	spheroid	
<i>Diplopsalis acuta</i> (Apstein) Entz, 1904	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Diplopsalopsis</i> Meunier, 1910	cell	sphere	
<i>Diplopsalopsis bomba</i> (Stein ex Jörgensen) J.D.Dodge & S.Toriumi, 1993	cell	spheroid	$h=0,80 \times d$
<i>Diplosiga</i> Frenzel, 1891	cell	spheroid	
<i>Diplostauron</i> A.A.Korschikov, 1925	cell	spheroid	
<i>Discocelis</i> Ehrenberg, 1836	cell	sphere	
<i>Discolithina</i> A.R.Loeblich Jr. & Tappan, 1963	cell	sphere	
<i>Discoplastis</i> Triemer, 2006	cell	spindle	
<i>Discosphaera</i> Haeckel, 1894	cell	sphere	
<i>Discostella</i> Houk & Klee, 2004	cell	cylinder	$h=0,65 \times d$
<i>Dispora</i> H.Printz, 1913	cell	spheroid	
<i>Dissodinium pseudocalani</i> (Gonnert) Drebes ex Elbrachter & Drebes, 1978	spore	sphere	
<i>Dissodium</i> T.H.Abé, 1941	cell	spheroid	$h=0,80 \times d$
<i>Distephanus</i> E.Stöhr, 1880	cell	sphere × 0,5	
<i>Distigma</i> Ehrenberg, 1831	cell	spindle	
<i>Distigma gracile</i> E.G.Pringsheim, 1942	cell	cone with half sphere	
<i>Ditylum</i> J.W.Bailey, 1861	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3)} \times l$
<i>Docidium</i> Brébisson ex Ralfs, 1848	cell	cylinder	
<i>Dolichomastix</i> Manton, 1977	cell	spheroid	
<i>Dolichospermum</i> (Ralfs ex Bornet & Flahault) P.Wacklin, L.Hoffmann & J.Komárek, 2009	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Dolichospermum fuscum</i> (H.Hill) P.Wacklin, L.Hoffmann & J.Komárek, 1976	cell	spheroid	
<i>Dolichospermum planctonicum</i> (Brunnth.) Wacklin, L.Hoffm. & Komárek, 2009	cell	spheroid	
<i>Dolichospermum smithii</i> (Komárek) Wacklin L.Hoffm. & Komárek, 2009	cell	spheroid	
<i>Donkinia</i> Ralfs, 1861	cell	elliptic cylinder	
<i>Draparnaldia</i> Bory de Saint-Vincent, 1808	filament	cylinder	
<i>Druridgea</i> De Toni, 1894	cell	cylinder \times 0,95	
<i>Ducellieria</i> Teilling, 1957	cell	spheroid	
<i>Dunaliella</i> Teodoresco, 1905	cell	spheroid	
<i>Durinskia</i> S.Carty & E.R.Cox, 1986	cell	sphere	
<i>Durinskia baltica</i> (K.M.Levander) S.Carty & E.R.Cox, 1986	cell	sphere \times 0,8	
<i>Dysmorphococcus</i> Takeda, 1916	cell	spheroid	
<i>Dzensia</i> Voronichin, 1929	cell	spheroid	
<i>Ebria</i> Borgert, 1891	cell	sphere \times 0,35	
<i>Ecballocystis</i> Bohlin, 1897	cell	spheroid	
<i>Echinocoleum</i> C.-C.Jao & K.T.Lee, 1947	cell	spheroid	
<i>Echinosphaerella</i> G.M.Smith, 1920	cell	sphere	
<i>Ehrenbergiulva</i> A.Witkowski, Lange-Bertalot & D.Metzeltin, 2004	cell	cylinder	
<i>Elakatothrix</i> Wille, 1898	cell	spindle	
<i>Ellerbeckia</i> R.M.Crawford, 1988	cell	cylinder	
<i>Ellipsoidion</i> Pascher, 1937	cell	sphere	
<i>Emergosphaera</i> W.W.Miller, 1921	cell	spheroid	
<i>Emiliana</i> W.W.Hay & H.P.Mohler, 1967	cell	sphere	
<i>Enallax</i> Pascher, 1943	cell	spindle	
<i>Enallax costatus</i> (Schmidle) Pascher, 1943	cell	spheroid	
<i>Encyonema</i> Kützing, 1834	cell	elliptic cylinder \times 0,65	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema alpinum</i> (Grunow) D.G.Mann, 1990	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema brehmii</i> (Hustedt) D.G.Mann, 1990	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema caespitosum</i> Kützing, 1849	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema elginense</i> (Krammer) D.G.Mann, 1990	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema gaeumannii</i> (Meister) E.Y.Haworth & M.G.Kelly, 1998	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema hebridicum</i> Grunow ex Cleve, 1891	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema minutum</i> (Hilse) D.G.Mann, 1990	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema norvegica</i> (Grunow) Bukhtiyarova, 1995	cell	elliptic cylinder \times 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Encyonema obscurum</i> (Krasske) D.G.Mann, 1990	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema paucistriatum</i> (Cleve-Euler) D.G.Mann, 1990	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema perpusillum</i> (A.Cleve) D.G.Mann, 1990	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema prostratum</i> (Berkeley) Kützing, 1844	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema reichardtii</i> (Krammer) D.G.Mann, 1990	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema silesiacum</i> (Bleisch) D.G.Mann, 1990	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonema ventricosum</i> (C.Agardh) Grunow in A.Schmidt et al., 1885	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Encyonopsis</i> K.Krammer, 1997	cell	elliptic cylinder	
<i>Encyonopsis cesatii</i> (Rabenhorst) K.Krammer, 1997	cell	lanceolate cylinder × 1,15	
<i>Encyonopsis cesatii</i> var. <i>geitleri</i> Krammer	cell	rhombic prism	
<i>Encyonopsis descripta</i> (Hustedt) Krammer	cell	elliptic cylinder × 0,9	
<i>Encyonopsis descripta</i> var. <i>asymmetrica</i> Krammer	cell	elliptic cylinder × 0,9	
<i>Encyonopsis microcephala</i> (Grunow) Krammer, 1997	cell	elliptic cylinder × 0,9	
<i>Encyonopsis subminuta</i> Krammer & E.Reichardt	cell	elliptic cylinder × 0,9	
<i>Endictya</i> Ehrenberg, 1845	cell	cylinder	
<i>Endoderma</i> Lagerheim, 1883	cell	spheroid	
<i>Endodinium</i> Hovasse, 1922	cell	ellipsoid	
<i>Ensiculifera</i> Balech, 1967	cell	cone with half sphere	
<i>Entocladia</i> Reinke, 1879	cell	spheroid	
<i>Entomoneis</i> Ehrenberg, 1845	cell	elliptic cylinder	$d_2=0,20 \times d_1$
<i>Entomoneis ornata</i> (J.W.Bailey) R.M.Patrick &, 1975	cell	elliptic cylinder	$d_2=0,15 \times d_1; h=0,67 \times d_1$
<i>Entophysalis</i> Kützing, 1843	cell	spheroid	
<i>Entosiphon</i> Stein, 1878	cell	ellipsoid	
<i>Eolimna</i> Lange-Bertalot & W.Schiller, 1997	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Epicystis</i> Pascher, 1930	cell	sphere	
<i>Epigloeosphaera</i> Komárková-Legnerová, 1991	cell	spheroid	
<i>Epilithia</i> Ercegovic, 1932	cell	spheroid	
<i>Epipyxis</i> Ehrenberg, 1838	cell	spheroid	
<i>Epithemia</i> Kützing, 1844	cell	elliptic cylinder × 0,65	$d_2=0,83 \times h; h=1,20 \times d_2$
<i>Ercegovicia</i> De Toni, 1936	cell	spheroid	
<i>Eremosphaera</i> De Bary, 1858	cell	spheroid	
<i>Ericiolus</i> H.A.Thomsen, 1995	cell	spheroid	
<i>Erkenia</i> Skuja, 1948	cell	sphere	
<i>Erythrospidinium</i> P.C.Silva, 1960	cell	ellipsoid	
<i>Ethmodiscus</i> Castracane, 1886	cell	cylinder × 0,9	$h=0,50 \times d$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Euastropsis</i> Lagerheim, 1894	cell	double cone	
<i>Euastrum</i> Ehrenberg ex Ralfs, 1848	cell	ellipsoid	$d_2=0,65 \times d_1$
<i>Eucampia</i> Ehrenberg, 1839	cell	elliptic cylinder	$d_2=0,40 \times d_1$
<i>Eucampia zodiacus</i> Ehrenberg, 1839	cell	elliptic cylinder	$d_2=0,44 \times d_1$
<i>Eucapsis</i> Clements & Shantz, 1909	cell	sphere	
<i>Eucoconeis</i> Cleve ex F.Meister, 1912	cell	elliptic cylinder	
<i>Eudorina</i> Ehrenberg, 1832	cell	sphere	
<i>Euglena</i> Ehrenberg, 1830	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Euglena acusformis</i> J.Schiller	cell	spindle	
<i>Euglena chlamydophora</i> Mainx, 1927	cell	spindle	
<i>Euglena fenestrata</i> A.Elenkin	cell	cylinder	
<i>Euglena geniculata</i> Dujardin, 1841	cell	spindle	
<i>Euglena granulata</i> (Klebs) F.Schmitz, 1884	cell	spindle	
<i>Euglena guentheri</i> M.Gojdics, 1953	cell	spindle	
<i>Euglena hemichromata</i> Skuja, 1948	cell	spindle	
<i>Euglena ignobilis</i> L.P.Johnson, 1944	cell	cylinder	
<i>Euglena limnophila</i> Lemmermann, 1898	cell	spindle	
<i>Euglena megalithus</i> Skuja, 1939	cell	spindle	
<i>Euglena mutabilis</i> F.Schmitz, 1884	cell	cylinder	
<i>Euglena nana</i> L.P.Johnson, 1944	cell	spindle	
<i>Euglena obtusa</i> F.Schmitz, 1884	cell	spindle	
<i>Euglena pascheri</i> Swirenko, 1915	cell	spindle	
<i>Euglena pisciformis</i> Klebs, 1883	cell	spindle	
<i>Euglena proxima</i> P.A.Dangeard, 1901	cell	spindle	
<i>Euglena retronata</i> L.P.Johnson, 1944	cell	spindle	
<i>Euglena rostrata</i> (Schiller) Triemer, 2006	cell	spindle	
<i>Euglena rostrifera</i> L.P.Johnson, 1944	cell	spindle	
<i>Euglena sociabilis</i> P.A.Dangeard, 1901	cell	spindle	
<i>Euglena texta</i> (Dujardin) Hübner, 1886	cell	spheroid	
<i>Euglena vagans</i> Deflandre, 1932	cell	spindle	
<i>Euglena viridis</i> (O.F.Müller) Ehrenberg, 1830	cell	spindle	
<i>Euglenaria</i>	cell	spindle	
<i>Euglenopsis</i> Klebs, 1892	cell	spindle	
<i>Eunotia</i> Ehrenberg, 1837	cell	elliptic cylinder $\times 1,03$	$r_2=0,50 \times h; h=2,00 \times r_2$
<i>Eunotia zasuminensis</i> (Cabejszekówna) Körner	cell	cuboid $\times 0,85$	$b=1,00 \times c; c=1,00 \times b$
<i>Eunotogramma</i> J.F.Weisse, 1855	cell	elliptic cylinder	$r_2=0,25 \times d_1$
<i>Eupodiscus</i> J.W.Bailey, 1851	cell	cylinder	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Eutetramorus</i> Walton, 1918	cell	sphere	
<i>Eutreptia</i> Perty, 1852	cell	spheroid	
<i>Eutreptiella</i> A.da Cunha, 1914	cell	spheroid	
<i>Eutreptiella pomquetensis</i> (McLachlan, Seguel & Fritz) Marin & Melkonian, 2003	cell	spindle	
<i>Excentrosphaera</i> G.T.Moore, 1901	cell	ellipsoid	
<i>Extubocellulus</i> Hasle, Stosch & Syvertsen, 1983	cell	elliptic cylinder	
<i>Falcula</i> M.Voight, 1960	cell	elliptic cylinder × 1,03	$r_2=0,50 \times h$; $h=2,00 \times r_2$
<i>Fallacia</i> Stickle & D.G.Mann, 1990	cell	elliptic cylinder	$d_2=0,50 \times d_1$; $h=0,65 \times d_2$
<i>Fibrocapsa</i> S.Toriumi & H.Takano, 1973	cell	spheroid	
<i>Fischerella</i> (Bornet & Flahault) Gomont, 1895	filament	cylinder	
<i>Fistulifera</i> Lange-Bertalot, 1997	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Florenciella</i> Mead & De Falla, 1965	cell	sphere	
<i>Florisphaera</i> H.Okada & S.Honjo, 1973	cell	spheroid	
<i>Follicularia</i> W.W.Miller, 1924	cell	sphere	
<i>Fortiea</i> De Toni, 1936	filament	cylinder	
<i>Fortiella</i> Pascher, 1927	cell	ellipsoid	
<i>Fossula</i> G.R.Hasle, E.E.Syvertsen & C.H.von Quillfeldt, 1996	cell	elliptic cylinder	
<i>Fotterella</i> R.Buck, 1978	cell	spheroid	
<i>Fragilaria</i> Lyngbye, 1819	cell	elliptic cylinder	$d_2=1,12 \times h$; $h=0,89 \times d_2$
<i>Fragilaria crotonensis</i> Kitton, 1869	cell	rhombic prism × 0,9	$d_2=0,07 \times d_1$
<i>Fragilaria crotonensis</i> var. <i>oregona</i> H.E.Sovereign, 1958	cell	rhombic prism	
<i>Fragilaria gaillonii</i> (Bory) Lange-Bertalot	cell	elliptic cylinder	$d_2=0,05 \times d_1$; $h=2,00 \times d_2$
<i>Fragilaria heidenii</i> Østrup, 1910	cell	elliptic cylinder	$d_2=0,22 \times d_1$
<i>Fragilaria islandica</i> Grunow ex Van Heurck, 1881	cell	elliptic cylinder	$d_2=0,07 \times d_1$
<i>Fragilaria istvanffyvi</i> Pantocsek, 1902	cell	elliptic cylinder	$d_2=0,24 \times d_1$
<i>Fragilaria reicheltii</i> (Voigt) L-B, 1986	cell part	cuboid	$c=1,00 \times b$
<i>Fragilariforma</i> D.M.Williams & Round, 1988	cell	elliptic cylinder	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Fragilariopsis</i> Hustedt, 1913	cell	elliptic cylinder	
<i>Fragilariopsis cylindriformis</i> (Hasle) Hasle, 1993	cell	cuboid	
<i>Fragilariopsis cylindrus</i> (Grunow) Krieger, 1954	cell	cuboid × 0,9	$b=1,00 \times c$; $c=1,00 \times b$
<i>Fragilidium</i> Balech ex Loeblich III, 1965	cell	spheroid	
<i>Franceia</i> Lemmermann, 1898	cell	spheroid	
<i>Frustulia</i> Rabenhorst, 1853	cell	elliptic cylinder	
<i>Frustulia crassinervia</i> (Brébisson) Lange-Bertalot & Krammer, 1996	cell	elliptic cylinder × 0,9	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Frustulia erifuga</i> Lange-Bertalot & Krammer, 1996	cell	lanceolate cylinder	
<i>Frustulia krammeri</i> Lange-Bertalot & Metzeltin, 1998	cell	lanceolate cylinder	
<i>Frustulia pangaeopsis</i> Lange-Bertalot, 2001	cell	lanceolate cylinder	
<i>Frustulia quadrisinuata</i> Lange-Bertalot, 1996	cell	lanceolate cylinder	
<i>Frustulia rhomboides</i> (Ehrenberg) De Toni, 1891	cell	rhombic prism	
<i>Frustulia rhomboides</i> var. <i>amphipleuroides</i> (Grunow) Cleve, 1894	cell	lanceolate cylinder	
<i>Frustulia saxonica</i> Rabenhorst, 1853	cell	lanceolate cylinder	
<i>Frustulia septentrionalis</i> Lange-Bertalot, 1996	cell	lanceolate cylinder	
<i>Fusola</i> Snow, 1903	cell	spindle	
<i>Gambierdiscus</i> Adachi & Fukuyo, 1979	cell	ellipsoid	
<i>Gardnerula</i> G.De Toni, 1936	filament	cylinder	
<i>Geissleria</i> Lange-Bertalot & Metzeltin, 1996	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Geissleria decussis</i> (Østrup) Lange-Bertalot & Metzeltin, 1996	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Geissleria ignota</i> (Krasske) Lange-Bertalot & Metzeltin, 1996	cell	cuboid $\times 0,85$	$c=1,00 \times b$
<i>Geitleribactron</i> Komárek, 1975	cell	cylinder	
<i>Geitlerinema</i> (Anagnostidis & Komárek) Anagnostidis, 1989	filament	cylinder	
<i>Gemellicystis</i> Teiling, 1946	cell	spheroid	
<i>Geminella</i> Turpin, 1828	cell	cylinder	
<i>Geminigera</i> D.R.A.Hill, 1991	cell	ellipsoid	
<i>Genicularia</i> De Bary, 1858	cell	cylinder	
<i>Gephyria</i> Arnott, 1858	cell	elliptic cylinder	
<i>Gephyrocapsa</i> Kamptner, 1943	cell	sphere	
<i>Germainiella</i> Lange-Bertalot & Metzeltin, 2005	cell	ellipsoid	
<i>Glaucocystis</i> Itzigsohn, 1866	cell	spheroid	
<i>Glaucospira</i> Lagerheim, 1892	filament	cylinder	
<i>Glenodiniopsis</i> Woloszynska, 1916	cell	ellipsoid	
<i>Glenodinium</i> Ehrenberg, 1836	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Gliscolithus</i> R.E.Norris, 1985	cell	spheroid	
<i>Glochiococcus</i> G.B.De Toni, 1888	cell	sphere	
<i>Gloeoactinium</i> G.M.Smith, 1926	cell	sphere	
<i>Gloeobacter</i> Rippka, Waterbury & Cohen-Bazire, 1974	cell	spheroid	
<i>Gloeobotrys</i> Pascher, 1930	cell	sphere	
<i>Gloeocapsa</i> Kützing, 1843	cell	sphere	
<i>Gloeocapsa siderochlamys</i> (Skuja) Starmach, 1966	cell	spheroid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Gloeocapsopsis</i> Geitler ex Komárek, 1993	cell	spheroid	
<i>Gloeochaete</i> Lagerheim, 1883	cell	sphere	
<i>Gloeochloris</i> Pascher, 1932	cell	spheroid	
<i>Gloeochrysis</i> Pascher, 1925	cell	spheroid	
<i>Gloeococcus</i> A.Braun, 1850	cell	spheroid	
<i>Gloeocystis</i> Nägeli, 1849	cell	spheroid	
<i>Gloeocystis minuta</i> J.Komárek, 1979	cell	sphere	
<i>Gloeomonas</i> Klebs, 1886	cell	spheroid	
<i>Gloeoplax</i> Schmidle, 1899	cell	spheroid	
<i>Gloeopodium</i> Pascher, 1939	cell	ellipsoid	
<i>Gloeothece</i> Nägeli, 1849	cell	cylinder	
<i>Gloeotila</i> Kützing, 1843	cell	cylinder	
<i>Glootrichia</i> J.Agardh ex Bornet & Flahault, 1886	filament	cylinder	
<i>Glyphodesmis</i> Greville, 1862	cell	elliptic cylinder	
<i>Golenkinia</i> Chodat, 1894	cell	sphere	
<i>Golenkiniopsis</i> Korshikov, 1953	cell	sphere	
<i>Gomontiella</i> Teodoresco, 1901	filament	elliptic cylinder × 0,5	
<i>Gomphocymbella</i> Otto Müller, 1905	cell	elliptic cylinder	
<i>Gomphocymbellopsis</i> K.Krammer, 2003	cell	elliptic cylinder	
<i>Gomphoneis</i> Cleve, 1894	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Gomphonema</i> Ehrenberg, 1832	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Gomphonemopsis</i> L.K.Medlin, 1986	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Gomphonitzschia</i> Grunow, 1868	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Gomphoseptatum</i> L.K.Medlin, 1986	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Gomphosphaeria</i> Kützing, 1836	cell	spheroid	
<i>Gomphosphenia</i> Lange-Bertalot, 1995	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Gonatozygon</i> de Bary, 1858	cell	cylinder	
<i>Gongrosira</i> Kützing, 1843	filament	cylinder	
<i>Gonioceros</i> H.Peragallo & M.Peragallo, 1907	cell	elliptic cylinder	$d_2=0,80 \times d_1$
<i>Goniochloris</i> Geitler, 1928	cell	triangular prism (equilateral) × 0,52	$m=0,50 \times \sqrt{(3)} \times l; h=0,30 \times l$
<i>Goniodoma</i> Stein, 1883	cell	sphere	
<i>Goniodoma polyedricum</i> (Pouchet) Jörgensen, 1899	cell	spheroid	
<i>Goniomonas</i> Stein, 1878	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Gonium</i> O.F.Müller, 1773	cell	spheroid	
<i>Gonyaulax</i> Diesing, 1866	cell	cone with half sphere	$h=1,30 \times d$
<i>Gonyaulax alaskensis</i> Kofoid, 1911	cell	spheroid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Gonyaulax apiculata</i> (Pénard) Entz, 1904	cell	spheroid	
<i>Gonyaulax diegensis</i> Kofoid, 1911	cell	spheroid	
<i>Gonyaulax grindleyi</i> Reinecke, 1967	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Gonyaulax verior</i> Sournia, 1973	cell	cone with half sphere \times 0,75	
<i>Gonyostomum</i> K.Diesing, 1866	cell	ellipsoid	$d_2=0,65 \times d_1$
<i>Gonyostomum semen</i> (Ehrenberg) Diesing, 1866	cell	ellipsoid	$d_2=0,33 \times d_1$
<i>Gossleriella</i> Schütt, 1892	cell	cylinder	
<i>Gotoius</i> T.H.Abé ex Matsuoka, 1988	cell	spheroid	
<i>Grammatophora</i> Ehrenberg, 1840	cell	elliptic cylinder	$d_2=0,20 \times d_1$; $h=0,40 \times d_1$
<i>Grammatophora marina</i> (Lyngbye) Kützing, 1844	cell	elliptic cylinder	$d_2=0,25 \times d_1$; $h=0,50 \times d_1$
<i>Grammatophora oceanica</i> Ehrenberg, 1840	cell	elliptic cylinder	$d_2=0,13 \times d_1$; $h=0,22 \times d_1$
<i>Grammatophora serpentina</i> Ehrenberg, 1844	cell	elliptic cylinder	$d_2=0,18 \times d_1$; $h=0,60 \times d_1$
<i>Grammonema</i> C.Agardh, 1832	cell	elliptic cylinder	
<i>Granulocystis</i> Hindák, 1977	cell	spheroid	
<i>Granulocystopsis</i> Hindák, 1977	cell	spheroid	
<i>Groenbladia</i> Teiling, 1952	cell	cylinder	
<i>Grunowia</i> Rabenhorst, 1864	cell	elliptic cylinder	
<i>Grunowia solgensis</i> (Cleve-Euler) M.Aboal, 2003	cell	elliptic cylinder \times 1,15	
<i>Grunowia tabellaria</i> (Grunow) Rabenhorst, 1864	cell	rhombic prism \times 1,15	
<i>Guillardia</i> D.R.A.Hill & R.Wetherbee, 1990	cell	spheroid	
<i>Guinardia</i> H.Peragallo, 1892	cell	cylinder	
<i>Gymnodinium</i> Stein, 1878	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Gymnodinium abbreviatum</i> Kofoid & Swezy, 1921	cell	spheroid	
<i>Gymnodinium corollarium</i> A.M.Sundström, Kremp & Daugbjerg, 2009	cell	sphere	
<i>Gymnodinium gracile</i> Bergh, 1881	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Gymnodinium helveticum</i> Penard, 1891	cell	double cone	
<i>Gymnodinium lantzschii</i> Utermöhl, 1925	cell	cone with half sphere	$h=0,67 \times d$
<i>Gymnodinium simplex</i> (Lohmann) Kofoid & Swezy, 1921	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Gymnodinium uberrimum</i> (G.J.Allman) Kofoid & Swezy, 1921	cell	spheroid	
<i>Gymnodinium vestificii</i> Schütt, 1885	cell	double cone	
<i>Gyrodinium</i> Kofoid & Swezy, 1921	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Gyrodinium britannicum</i> Kofoid & Swezy, 1921	cell	spindle	
<i>Gyrodinium fissum</i> (Levander) Kofoid & Swezy, 1921	cell	spheroid	
<i>Gyrodinium linguliferum</i> Lebour, 1925	cell	cone with half sphere	

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<i>Gyrodinium pingue</i> (Schütt) Kofoid & Swezy, 1921	cell	spheroid	
<i>Gyrodinium spirale</i> (Bergh) Kofoid & Swezy, 1921	cell	double cone	
<i>Gyromitus</i> Skuja, 1939	cell	ellipsoid	
<i>Gyrosigma</i> Hassall, 1845	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Gyrosigma acuminatum</i> (Kützing) Rabenhorst, 1853	cell	lanceolate cylinder	$h=0,71 \times d_2$
<i>Gyrosigma acuminatum</i> var. <i>brebissonii</i> (Grunow) Cleve, 1894	cell	elliptic cylinder	
<i>Gyrosigma attenuatum</i> (Kützing) Rabenhorst, 1853	cell	lanceolate cylinder	$h=0,60 \times d_2$
<i>Gyrosigma balticum</i> (Ehrenberg) Rabenhorst, 1853	cell	cuboid × 0,9	$c=1,28 \times b$
<i>Gyrosigma macrum</i> (W.Smith) J.W.Griffith & Henfrey, 1856	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Gyrosigma scalproides</i> (Rabenhorst) Cleve, 1894	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Gyrosigma scalproides</i> var. <i>eximia</i> (Thwaites) Cleve, 1894	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Gyrosigma sciotoense</i> (W.S.Sullivant) Cleve	cell	elliptic cylinder	
<i>Gyrosigma strigilis</i> (W.Smith) J.W.Griffin & Henfrey, 1856	cell	elliptic cylinder	$h=0,86 \times d_2$
<i>Gyrosigma tenuissimum</i> (W.Smith) Griffith & Henfrey, 1856	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Gyrosigma wansbeckii</i> (Donkin) Cleve, 1894	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Haematococcus</i> Flotow, 1844	cell	spheroid	
<i>Hafniomonas</i> Ettl & Moestrup, 1980	cell	spheroid	
<i>Halamphora</i> (Cleve) Levkov, 2009	cell	elliptic cylinder × 0,65	$d_2=0,50 \times h; h=2,00 \times d_2$
<i>Halopappus</i> Lohmann, 1912	cell	spheroid	
<i>Halosphaera</i> K.J.F.Schmitz, 1878	cell	sphere	
<i>Hannaea</i> R.M.Patrick, 1966	cell	elliptic cylinder × 1,03	$r_2=0,50 \times h; h=2,00 \times r_2$
<i>Hantzschia</i> Grunow, 1877	cell	cuboid × 0,85	$b=1,00 \times c; c=1,00 \times b$
<i>Hanusia</i> J.A.Deane, D.R.A.Hill, S.J.Brett & G.I.McFadden, 1998	cell	spheroid × 0,9	
<i>Hapalosiphon</i> Nägeli ex Bornet & Flahault, 1886	filament	cylinder	
<i>Haplotaenium</i> T.Bando, 1988	cell	cylinder	
<i>Haptolina</i> Edvardsen & Eikrem, 2011	cell	spheroid	
<i>Haptolina hirta</i> (Manton) Edvardsen & Eikrem, 2011	cell	ellipsoid	$d_2=0,85 \times d_1$
<i>Hariotina</i> P.-A.Dangeard, 1889	cell	sphere	
<i>Haslea</i> Simonsen, 1974	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Haslea ostrearia</i> (Gaillon) Simonsen, 1974	cell	elliptic cylinder	
<i>Haslea spicula</i> (Hickie) Lange-Bertalot, 1997	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Hassallia</i> Berkeley ex Bornet & Flahault, 1887	filament	cylinder	
<i>Hegewaldia</i> T.Proschold, C.Bock, W.Luo & L.Krienitz,	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
2010			
<i>Heimansia</i> Coesel, 1993	half-cell	spheroid	
<i>Helicopedinella</i> Sekiguchi, Kawachi, Nakayama & Inouye, 2003	cell	sphere	
<i>Helicosphaera</i> Kamptner, 1954	cell	sphere	
<i>Helicotheca</i> Ricard, 1987	cell	cuboid	$b=0,09 \times a$
<i>Helladosphaera</i> Kamptner, 1937	cell	spheroid	
<i>Hemiaulus</i> Heiberg, 1863	cell	elliptic cylinder	
<i>Hemidinium</i> Stein, 1878	cell	ellipsoid	
<i>Hemidiscus</i> Wallich, 1860	cell	elliptic cylinder $\times 0,65$	$d_2=1,25 \times h; h=0,80 \times d_2$
<i>Hemiselmis</i> Parke, 1949	cell	cone with half sphere	
<i>Hemitoma</i> Skuja, 1939	cell	spheroid	
<i>Herdmania</i> Dodge, 1981	cell	spheroid	
<i>Heterocapsa</i> Stein, 1883	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Heterocapsa rotundata</i> (Lohmann) G.Hansen, 1995	cell	cone with half sphere	
<i>Heterocapsa triquetra</i> (Ehrenberg) F.Stein, 1883	cell	double cone	
<i>Heterochromonas</i> Pascher, 1912	cell	spheroid	
<i>Heterodesmus</i> Ettl, 1956	cell	cone with half sphere	
<i>Heterodinium</i> Kofoid, 1906	cell	cone with half sphere	
<i>Heterodinium murrayi</i> Kofoid, 1906	cell	spheroid	
<i>Heterogloea</i> Pascher, 1930	cell	sphere	
<i>Heteroleibleinia</i> (Geitler) L.Hoffmann, 1905	filament	cylinder	
<i>Heteronema hexagonum</i> (Playfair) Skuja, 1948	cell	spheroid	
<i>Heterosigma</i> Y.Hada ex Y.Hara & M.Chihara, 1987	cell	spheroid	
<i>Heterosigma akashiwo</i> (Y.Hada) Y.Hada ex Y.Hara & M.Chihara, 1987	cell	sphere	
<i>Heterothrix</i> Pascher, 1932	cell	cylinder	
<i>Heterotrichella</i> H.Reisigl, 1964	cell	cylinder	
<i>Hexamitus</i> Dujardin	cell	ellipsoid	
<i>Hillea</i> Schiller, 1925	cell	cone with half sphere	
<i>Hindakia</i> C. Bock, Proschold & Krienitz, 2010	cell	spheroid	
<i>Hindakochloris</i> A.Comas, 1992	cell	sphere	
<i>Hippodonta</i> Lange-Bertalot, Witkowski & Metzeltin, 1996	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Hippodonta capitata</i> (Ehrenberg) Lange-Bertalot, Metzeltin & Witkowski, 1996	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Hippodonta costulata</i> (Grunow) Lange-Bertalot, Metzeltin & Witkowski, 1996	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Hippodonta costulatifomis</i> Lange-Bertalot, Metzeltin &	cell	lanceolate cylinder	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
Witkowski, 1996			
<i>Hippodonta coxiae</i> Lange-Bertalot, 2001	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Hippodonta lesmonensis</i> (Hustedt) Lange-Bertalot, Metzeltin & Witkowski, 1996	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Hippodonta lueneburgensis</i> (Grunow) Lange-Bertalot, Metzeltin & Witkowski, 1996	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Hippodonta subcostulata</i> (Hustedt) Lange-Bertalot, Metzeltin & Witkowski, 1996	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Histioneis</i> Stein, 1883	cell	ellipsoid	
<i>Holococcolithophora</i> L.Cros & J.R.Young, 2005	cell	sphere	
<i>Holopedia</i> Lagerheim, 1893	cell	spheroid	
<i>Homoeothrix</i> (Thuret ex Bornet & Flahault) Kirchner, 1898	filament	cylinder	
<i>Homozygosphaera</i> Deflandre, 1952	cell	sphere	
<i>Hormidium</i> Kützing, 1843	cell	cylinder	
<i>Hormoscilla</i> K.Anagnostidis & J.Komárek, 1988	filament	cylinder	
<i>Hormotila</i> Borzì, 1883	cell	sphere	
<i>Hortobagyiella</i> L.Hajdu, 1975	cell	ellipsoid	
<i>Hustedtiella</i> Simonsen, 1960	cell	elliptic cylinder	
<i>Hyalodiscus</i> Ehrenberg, 1845	cell	cylinder × 0,83	
<i>Hyalogonium</i> Pascher, 1927	cell	spindle	
<i>Hyaloraphidium</i> Pascher & Korshikov, 1931	cell	spindle	
<i>Hyalotheca</i> Ehrenberg ex Ralfs, 1848	cell	cylinder	
<i>Hydrianum</i> Rabenhorst, 1868	cell	ellipsoid	
<i>Hydrococcus</i> Kützing, 1833	cell	spheroid	
<i>Hydrocoleum</i> Kützing Ex Gomont, 1892	filament	cylinder	
<i>Hydrocoryne</i> H.Schwabe ex Bornet & Flahault, 1886	filament	cylinder	
<i>Hydrodictyon</i> Roth, 1797	cell	cylinder	
<i>Hydrosera</i> G.C.Wallich, 1858	cell	triangular prism (equilateral) × 1,25	$m=0,50 \times \sqrt{(3)} \times l$; $h=1,00 \times l$
<i>Hydrurus</i> C.Agardh, 1824	cell	spheroid	
<i>Hyella</i> Bornet & Flahault, 1888	filament	cylinder	
<i>Hygropetra</i> Krammer & Lange-Bertalot, 2000	cell	elliptic cylinder	
<i>Hymenomonas</i> Stein, 1878	cell	spheroid	
<i>Imantonia</i> N.Reynolds, 1974	cell	sphere	
<i>Isactis</i> Thuret ex Bornet & Flahault, 1886	filament	cylinder	
<i>Isochrysis</i> Parke, 1949	cell	spheroid	
<i>Isocystis</i> Borzì ex Bornet & Flahault, 1888	filament	cylinder	
<i>Isoselmis</i> Butcher, 1967	cell	cone with half sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Isthmia</i> C.Agardh, 1832	cell	elliptic cylinder	
<i>Isthmochloron</i> H.Skuja, 1948	half-cell	spheroid	
<i>Jaaginema</i> Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Johannesbaptistia</i> G.De Toni, 1934	cell	spheroid	
<i>Karayevia</i> Round & L.Bukhtiyarova ex F.E.Round	cell	elliptic cylinder	
<i>Karayevia amoena</i> (Hustedt) Bukhtiyarova, 1999	cell	elliptic cylinder $\times 0,9$	
<i>Karayevia laterostrata</i> (Hustedt) Bukhtiyarova, 1999	cell	elliptic cylinder $\times 0,9$	
<i>Karayevia ploenensis</i> (Hustedt) J.C.Kingston, 2000	cell	elliptic cylinder $\times 0,9$	
<i>Karenia</i> G.Hansen & Moestrup, 2000	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Karenia mikimotoi</i> (Miyake & Kominami ex Oda) G.Hansen & Ø.Moestrup, 2000	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Karlodinium</i> J.Larsen, 2000	cell	spheroid	
<i>Karlodinium vitiligo</i> (D.Ballantine) J.Larsen, 2000	cell	ellipsoid	
<i>Katablepharis</i> Skuja, 1939	cell	spheroid	
<i>Katagnymene</i> Lemmermann, 1899	filament	cylinder	
<i>Katodinium</i> Fott, 1857	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Katodinium asymmetricum</i> (Massart) Loeblich, 1965	cell	cone with half sphere $\times 0,5$	
<i>Katodinium fungiforme</i> (Anissimova) A.R.Loeblich III, 1965	cell	spheroid	
<i>Katodinium glaucum</i> (Lebour) Loeblich III, 1965	cell	spindle $\times 0,5$	
<i>Kephyrion</i> Pascher, 1911	cell	spheroid	
<i>Kephyriopsis</i> Pascher & Ruttner, 1913	cell	spheroid	
<i>Keratococcus</i> Pascher, 1915	cell	spindle	
<i>Khawkinea</i> Jahn & McKibben, 1937	cell	spindle	
<i>Kirchneriella</i> Schmidle, 1893	cell	spindle	
<i>Kirchneriella aperta</i> Teiling, 1912	cell	spheroid	
<i>Kirchneriella arcuata</i> G.M.Smith	cell	spheroid	
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Playfair) Komárek, 1979	cell	cylinder	
<i>Kirchneriella microscopica</i> G.N.Nygaard, 1945	cell	cylinder	
<i>Kirchneriella obesa</i> (West) West & G.S.West, 1894	cell	spheroid $\times 0,9$	$h=0,50 \times d$
<i>Kirchneriella obesa</i> var. <i>major</i> (Bernard) G.M.Smith	cell	spheroid $\times 0,9$	$h=0,50 \times d$
<i>Kirchneriella pinguis</i> Hindák, 1977	cell	spheroid	
<i>Klebsormidium</i> P.C.Silva, K.Mattox & W.Blackwell, 1972	cell	cylinder	
<i>Kobayasiella</i> Lange-Bertalot, 1999	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Kobayasiella jaagii</i> (Meister) Lange-Bertalot, 1999	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Kofoidinium</i> Pavillard, 1928	cell	ellipsoid	$d_2=0,25 \times d_1$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Kolbesia</i> Round & L.Bukhtiyarova ex F.E.Round, 1998	cell	elliptic cylinder × 0,9	
<i>Kolbesia gessneri</i> (Hustedt) M.Aboal, 2003	cell	elliptic cylinder	
<i>Kolbesia kolbei</i> (Hustedt) F.E. Round & L. Bukhtiyarova ex Fourtanier & Kociolek, 1999	cell	elliptic cylinder	
<i>Koliella</i> Hindák, 1963	cell	spindle	
<i>Kolkwitzella</i> Lindemann, 1919	cell	cone with half sphere	
<i>Komarekia</i> Fott, 1981	cell	spheroid	
<i>Komma</i> D.R.A.Hill, 1991	cell	cone with half sphere	
<i>Komvophoron</i> Anagnostidis & Komárek, 1988	cell	cylinder	
<i>Korshikoviella</i> P.C.Silva, 1959	cell	spindle	
<i>Krasskella</i> R.Ross & P.A.Sims, 1978	cell	elliptic cylinder	
<i>Kryptoperidinium</i> Lindemann, 1924	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Kryptoperidinium foliaceum</i> (F.Stein) Lindemann, 1924	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Kyrtuthrix</i> Ercegovic, 1929	filament	cylinder	
<i>Laboea</i> Lohmann, 1908	cell	cone	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Lacunastrum</i> H.A.McManus, 2011	colony	cylinder	
<i>Lagerheimia</i> R.Chodat, 1895	cell	spheroid	
<i>Lagynion</i> Pascher, 1912	cell	spheroid	
<i>Lampriscus</i> A.Schmidt, 1882	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$
<i>Lanceola</i> F.Hindák, 1988	cell	spindle	
<i>Lauderia</i> Cleve, 1873	cell	cylinder	$h=0,90 \times d$
<i>Lauterborniella</i> Schmidle, 1900	cell	spindle	
<i>Leibleinia</i> (Gomont) L.Hoffman, 1985	filament	cylinder	
<i>Lemmermanniella</i> Geitler, 1942	cell	cylinder	
<i>Lemmermanniella flexa</i> Hindák, 1985	cell	spheroid	
<i>Lemnicola</i> Round & P.W.Basson, 1977	cell	elliptic cylinder	$d_2=0,35 \times d_1; h=0,80 \times d_2$
<i>Lennoxia</i> Thomsen & Buck, 1993	cell	spindle	
<i>Lepidodinium</i> Watanabe, Suda, Inouye, Sawaguchi & Chihara, 1990	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Lepochromulina</i> Scherffel, 1911	cell	spheroid	
<i>Lepocinclis</i> Perty, 1849	cell	spheroid	
<i>Lepocinclis acus</i> (O.F.Müller) Marin & Melkonian, 2003	cell	spindle	
<i>Lepocinclis fusca</i> (Klebs) Kosmala & Zakrys, 2005	cell	spindle	
<i>Lepocinclis helicoidea</i>	cell	spindle	
<i>Lepocinclis oxyuris</i> (Schmarda) Marin & Melkonian, 2003	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Lepocinclis spirogyroides</i> Marin & Melkonian, 2003	cell	ellipsoid	$d_2=0,75 \times d_1$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Lepocinclis spiroides</i> (Lemmermann) Marin & Melkonian, 2003	cell	spindle	
<i>Lepocinclis tripteris</i> (Dujardin) Marin & Melkonian, 2003	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Leptocylindrus</i> Cleve, 1889	cell	cylinder	
<i>Leptodiscus</i> Hertwig, 1877	cell	cylinder	
<i>Leptolyngbya</i> Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Leptothrix</i> Kützing, 1843	filament	cylinder	
<i>Lessardia</i> J.F.Saldarriaga & F.J.R.Taylor, 2003	cell	spindle	
<i>Leucocryptos</i> Butcher, 1967	cell	cone with half sphere	
<i>Leyanella</i> G.R.Hasle, H.A.von Stosch & E.E.Syvetsen, 1983	cell	elliptic cylinder	
<i>Licmophora</i> C.Agardh, 1827	cell	pyramid $\times 0,9$	$l_1=0,75 \times l_2$; $l_2=1,33 \times l_1$
<i>Limnococcus</i> (Komárek & Anagnostidis) Komárková, Jezberová, O.Komárek & Zapomelová, 2010	cell	spheroid	
<i>Limnothrix</i> Meffert, 1987	filament	cylinder	
<i>Lingulodinium</i> D.Wall, 1967	cell	cone with half sphere	$h=1,20 \times d$
<i>Lissodinium</i> Matzenauer, 1933	cell	ellipsoid	
<i>Lithocapsa</i> Ercegovic, 1925	cell	spheroid	
<i>Lithodesmium</i> Ehrenberg, 1839	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3)} \times l$; $h=0,80 \times l$
<i>Lobochlamys</i> T.Pröschold, B.Marin, U.W.Schlösser & M.Melkonian, 2001	cell	spheroid	
<i>Lobocystis</i> R.H.Thompson, 1952	cell	spheroid	
<i>Lobomonas</i> P.-A.Dangeard, 1899	cell	spheroid	
<i>Lophodinium</i> Lemmermann, 1910	cell	double cone	
<i>Lunella</i> P.Snoeijs, 1996	cell	elliptic cylinder	
<i>Lutherella</i> Pascher, 1930	cell	sphere	
<i>Luticola</i> D.G.Mann, 1990	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Luticola nivalis</i> (Ehrenberg) D.G.Mann, 1990	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Luticola paramutica</i> (Bock) D.G.Mann, 1990	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Luticola ventricosa</i> (Kützing) D.G.Mann, 1990	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Lyngbya</i> C.Agardh ex Gomont, 1892	filament	cylinder	
<i>Lyngbyopsis</i> N.L.Gardner, 1927	filament	cylinder	
<i>Lyrella</i> Karajeva, 1978	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Macrospermum</i> Komárek, 2008	filament	cylinder	
<i>Mallomonas</i> Perty, 1852	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Mallomonas akrokomos</i> Ruttner, 1913	cell	spindle	
<i>Mallomonas caudata</i> Ivanov, 1899	cell	cone with half sphere	

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<i>Mallomonas crassisquama</i> (Asmund) Fott, 1962	cell	spheroid	
<i>Mallomonas globosa</i> Schiller	cell	sphere	
<i>Mallomonas hirsuta</i> Conrad	cell	spheroid	
<i>Mallomonas punctifera</i> Korshikov, 1941	cell	spheroid	
<i>Mallomonas zellensis</i> Fott, 1962	cell	spheroid	
<i>Mamiella</i> Ø.Moestrup, 1984	cell	spheroid	
<i>Mantoniella</i> T.V.Desikachary, 1972	cell	spheroid	
<i>Martyana</i> Round, 1990	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Marvania</i> F.Hindák, 1976	cell	sphere	
<i>Mastigocoleus</i> Lagerheim ex Bornet & Flahault, 1886	filament	cylinder	
<i>Mastogloia</i> Thwaites ex W.Smith, 1856	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Mastogloia baltica</i> Grunow, 1880	cell	elliptic cylinder × 0,9	
<i>Mastogloia smithii</i> Thwaites ex W.Smith, 1856	cell	elliptic cylinder × 0,9	
<i>Mayamaea</i> Lange-Bertalot, 1997	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Mediopyxis</i> L.K.Medlin & Kühn, 2006	cell	elliptic cylinder	$d_2=0,22 \times d_1$
<i>Medusa</i> Loureiro, 1790	cell	sphere	
<i>Melosira</i> C.Agardh, 1824	cell	cylinder	
<i>Melosira dubia</i> C.G.Kützing	cell	cylinder × 0,9	
<i>Membraneis</i> Paddock, 1988	cell	elliptic cylinder	
<i>Menoidium</i> Perty, 1852	cell	ellipsoid	
<i>Menoidium tortuosum</i> (A.Stokes) Senn, 1900	cell	spindle	
<i>Meridion</i> C.Agardh, 1824	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Meringosphaera</i> Lohmann, 1932	cell	sphere	
<i>Merismopedia</i> Meyen, 1839	cell	sphere	
<i>Merismopedia elegans</i> A.Braun ex Kützing, 1849	cell	spheroid	
<i>Merotricha</i> Mereschkowsky, 1877	cell	ellipsoid	
<i>Mesocena</i> Ehrenberg, 1843	cell	sphere	
<i>Mesodinium</i> von Stein, 1862	cell	spheroid	
<i>Mesopedinella</i> Daugbjerg, 1996	cell	sphere	
<i>Mesoporos</i> Lillick, 1937	cell	spheroid	$h=0,60 \times d$
<i>Mesotaenium</i> Nägeli, 1849	cell	cylinder	
<i>Meuniera</i> P.C.Silva, 1996	cell	elliptic cylinder	$d_2=0,30 \times d_1$
<i>Michaelsarsia</i> Gran, 1912	cell	spheroid	
<i>Micracantha</i> Korshikov, 1953	cell	ellipsoid	
<i>Micracanthodinium</i> Deflandre, 1937	cell	sphere	
<i>Micractinium</i> Fresenius, 1858	cell	sphere	
<i>Micractinium belenophorum</i> (Korshikov) T.Prochold,	cell	ellipsoid	

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C.Block, W.Luo & L.Kreinitz, 2010			
<i>Micractinium pusillum</i> Fresenius, 1858	coenobium	sphere	
<i>Micrasterias</i> C.Agardh ex Ralfs, 1848	cell	elliptic cylinder $\times 0,8$	$h=0,30 \times d_2$
<i>Microchaete</i> Thuret ex Bornet & Flahault, 1886	filament	cylinder	
<i>Microcoleus</i> Desmazières ex Gomont, 1892	filament	cylinder	
<i>Microcostatus</i> Johansen & Sray, 1998	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Microcostatus naumannii</i> (Hustedt) Lange-Bertalot	cell	elliptic cylinder $\times 0,9$	
<i>Microcrocis</i> Richter, 1892	cell	spheroid	
<i>Microcystis</i> Lemmermann, 1907	cell	sphere	
<i>Microglena</i> Ehrenberg, 1832	cell	spheroid	
<i>Micromonas</i> I.Manton & M.Parke, 1960	cell	spheroid	
<i>Microsiphona</i> C.I.Weber, 1970	cell	cylinder $\times 0,9$	
<i>Microspora</i> Thuret, 1850	cell	cylinder	
<i>Microtabella</i> Round, 1990	cell	elliptic cylinder	
<i>Microthamnion</i> J.Agardh, 1892	filament	cylinder	
<i>Minidiscus</i> Hasle, 1973	cell	cylinder	
<i>Minuscula</i> Lebour, 1925	cell	cone $\times 0,5$	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Minutocellus</i> Hasle, Stosch & Syvertsen, 1983	cell	elliptic cylinder	
<i>Miosira</i> K.Krammer, Lange-Bertalot & W.Schiller, 1997	cell	cylinder $\times 0,9$	
<i>Mischococcus</i> Nägeli, 1849	cell	sphere	
<i>Monactinus</i> Corda, 1839	colony	cylinder	
<i>Monactinus simplex</i> (Meyen) Corda, 1839	cell	triangular prism $\times 0,67$	$h=1,00 \times l$
<i>Monallantus</i> Pascher, 1939	cell	spheroid	
<i>Monas</i> O.F.Müller, 1773	cell	sphere	
<i>Monoceros</i> A.C.J.Van Goor, 1924	cell	cylinder	
<i>Monochrysis</i> Skuja, 1948	cell	spheroid	
<i>Monochrysis aphanaster</i> Skuja, 1943	cell	ellipsoid	
<i>Monocosta</i> Thomsen, 1979	cell	spheroid	
<i>Monodus</i> Chodat, 1913	cell	spheroid	
<i>Monomastix</i> Scherffel, 1912	cell	ellipsoid	
<i>Monomorphina</i> Mereschkowsky, 1877	cell	ellipsoid	$d_2=0,43 \times d_1$
<i>Monoraphidium</i> Komárková-Legnerová, 1969	cell	spindle	
<i>Monoraphidium minutum</i> (Nägeli) Komárková-Legnerová, 1969	cell	spheroid	
<i>Monoraphidium nanum</i> (Ettl) Hindák, 1980	cell	spheroid	
<i>Monoraphidium skujae</i> B.Fott	cell	cylinder	
<i>Monosiga</i> W.S.Kent, 1878	cell	spheroid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Mougeotia</i> C.Agardh, 1824	cell	cylinder	
<i>Mougeotiopsis</i> Palla, 1894	cell	cylinder	
<i>Mucidosphaerium</i> C.Bock, Proschold & Krienitz, 2011	cell	spheroid	
<i>Muelleria</i> (Frenguelli) Frenguelli, 1945	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Muriella</i> J.B.Petersen, 1932	cell	sphere	
<i>Mychonastes</i> P.D.Simpson & S.D.Van Valkenburg, 1978	cell	spheroid	
<i>Mychonastes jurisii</i> (Hindák) Krienitz, C.Bock, Dadheech & Proschold, 2011	cell	sphere	
<i>Myochloris</i> J.H.Belcher & Swale, 1961	cell	spheroid	
<i>Myxobaktron</i> Schmidle, 1904	cell	spheroid	
<i>Myxohyella</i> Geitler, 1925	cell	spheroid	
<i>Myxosarcina</i> Printz, 1921	cell	spheroid	
<i>Nannochloris</i> Naumann, 1921	cell	spheroid	
<i>Nannochloropsis</i> D.J.Hibberd, 1981	cell	sphere	
<i>Nannoeca</i> Thomsen, 1988	cell	spheroid	
<i>Navicula</i> Bory de Saint-Vincent, 1822	cell	elliptic cylinder	$h=0,85 \times d_2$
<i>Navicula amphibola</i> Cleve, 1891	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Navicula bourrellyivera</i> Lange-Bertalot, A.Witkowski & K.Stachura, 1998	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula capitata</i> var. <i>capitata</i>	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Navicula capitatoradiata</i> Germain, 1981	cell	lanceolate cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Navicula cataracta-rheni</i> Lange-Bertalot, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula caterva</i> Hohn & Hellermann, 1963	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula clementis</i> Grunow, 1882	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Navicula cryptocephala</i> Kützing, 1844	cell	lanceolate cylinder	$h=0,56 \times d_2$
<i>Navicula cryptocephala</i> var. <i>veneta</i> (Kützing) Rabenhorst, 1864	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula cryptofallax</i> Lange-Bertalot & Hofmann, 1993	cell	rhombic prism	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Navicula cryptotenella</i> Lange-Bertalot, 1985	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula cryptotenelloides</i> Lange-Bertalot, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula densilineolata</i> (Lange-Bertalot) Lange-Bertalot, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula detenta</i> Hustedt, 1943	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Navicula digitoradiata</i> (Gregory) Ralfs, 1861	cell	elliptic cylinder	$h=0,83 \times d_2$
<i>Navicula digito-radiata</i> (Gregory) Ralfs, 1861	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula directa</i> (W.Smith) Ralfs, 1861	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula flantica</i> Grunow, 1860	cell	lanceolate cylinder	$h=1,00 \times d_2$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Navicula gregaria</i> Donkin, 1861	cell	lanceolate cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula halophiloides</i> Hustedt, 1959	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula hanseatica</i> Lange-Bertalot & Stachura, 1998	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula hasta</i> Pantocsek, 1892	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula irmengardis</i> Lange-Bertalot, 1996	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula iserentantii</i> Lange-Bertalot & Witkowski, 2000	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula kohlmaieri</i> Lange-Bertalot, 1998	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula kotschyi</i> Grunow	cell	elliptic cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula kotschyi</i> f. <i>kotschyi</i> Grunow	cell	elliptic cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula leptostriata</i> Jørgensen, 1948	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula lucinensis</i> (Hustedt) Mann, 1990	cell	elliptic cylinder	$d_2=0,50 \times d_1$; $h=0,65 \times d_2$
<i>Navicula menisculus</i> Schumann, 1867	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula monoculata</i> var. <i>omissa</i> (Hustedt) D.G.Mann, 1990	cell	elliptic cylinder	$d_2=0,50 \times d_1$; $h=0,65 \times d_2$
<i>Navicula mutica</i> var. <i>ventricosa</i> (Kützing) D.G.Mann, 1990	cell	elliptic cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula notha</i> Wallace, 1960	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula oahuensis</i> (Hustedt) Krammer, 1985	cell	pyramid \times 0,9	$l_1=0,75 \times l_2$; $l_2=1,33 \times l_1$
<i>Navicula oligotrphenta</i> Lange-Bertalot & Hofmann, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula oppugnata</i> Hustedt, 1945	cell	rhombic prism	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Navicula palpebralis</i> Brébisson ex W.Smith, 1853	cell	elliptic cylinder	$h=0,89 \times d_2$
<i>Navicula peregrina</i> (Ehrenberg) Kützing, 1844	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula peregrina</i> var. <i>meniscus</i> (J.Schumann) Grunow, 1895	cell	rhombic prism	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Navicula phyllepta</i> Kützing, 1844	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula phylleptosoma</i> Lange-Bertalot, 1999	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula platystoma</i> Ehrenberg, 1838	cell	elliptic cylinder	$h=0,82 \times d_2$
<i>Navicula pseudanglica</i> E.J.Cox, 1987	cell	elliptic cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula pseudolanceolata</i> Lange-Bertalot, 1980	cell	rhombic prism	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Navicula radiosa</i> Kützing, 1844	cell	rhombic prism	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Navicula reinhardtii</i> (Grunow) Grunow, 1877	cell	elliptic cylinder	$h=0,83 \times d_2$
<i>Navicula rhynchocephala</i> Kützing, 1844	cell	lanceolate cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula rhynchotella</i> Lange-Bertalot, 1993	cell	lanceolate cylinder \times 0,9	$h=0,75 \times d_2$
<i>Navicula ricardae</i> Lange-Bertalot, 2001	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula salinarum</i> Grunow, 1880	cell	lanceolate cylinder \times	$h=0,75 \times d_2$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
		0,9	
<i>Navicula salinarum</i> var. <i>minima</i> R.Kolbe, 1927	cell	lanceolate cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula salinarum</i> var. <i>rostrata</i> (Hustedt) Lange-Bertalot, 2001	cell	lanceolate cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula salinarum</i> var. <i>salinarum</i> Grunow, 1880	cell	lanceolate cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula schmassmannii</i> Hustedt, 1943	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula sieminskiae</i> Lange-Bertalot & Witkowski, 2001	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula streckeriae</i> Lange-Bertalot & Witkowski, 2000	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula subconcentrica</i> Lange-Bertalot, 2001	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula sublucidula</i> (Hustedt) Mann, 1990	cell	elliptic cylinder	$d_2=0,50 \times d_1$; $h=0,65 \times d_2$
<i>Navicula subrhynchocephala</i> Hustedt, 1935	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula supergregaria</i> Rumrich & Lange-Bertalot, 2000	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula tenerrima</i> (Hustedt) D.G.Mann, 1990	cell	elliptic cylinder	$d_2=0,50 \times d_1$; $h=0,65 \times d_2$
<i>Navicula transitans</i> Cleve, 1883	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula transitans</i> var. <i>derasa</i> (Grunow) Cleve, 1883	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula transitans</i> var. <i>derasa</i> f. <i>delicatula</i> Heimdal, 1970	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula tripunctata</i> (O.F.Müller) Bory de Saint-Vincent, 1822	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Navicula trivialis</i> Lange-Bertalot, 1980	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula vandamii</i> Schoeman & Archibald, 1987	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula vaneei</i> Lange-Bertalot, 1998	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula vanhoeffenii</i> Gran, 1897	cell	lanceolate cylinder × 0,85	$h=1,00 \times d_2$
<i>Navicula viridula</i> (Kützing) Ehrenberg, 1838	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Navicula viridula</i> var. <i>germainii</i> (Wallace) Lange-Bertalot, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Navicula wellneri</i> (Lange-Bertalot) Lange-Bertalot	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Navicula wildii</i> Lange-Bertalot, 1993	cell	lanceolate cylinder	$h=1,00 \times d_2$
<i>Naviculadicta</i> Lange-Bertalot, 1994	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Navicymbula</i> K.Krammer, 2003	cell	elliptic cylinder × 0,8	$d_2=0,83 \times h$; $h=1,20 \times d_2$
<i>Navigiolum</i> Lange-Bertalot	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Neglectella</i> Vodeničarov & Benderliev, 1971	cell	spheroid	
<i>Neidiopsis</i> Lange-Bertalot & Metzeltin, 1999	cell	cuboid × 0,85	$c=1,00 \times b$
<i>Neidium</i> Pfitzer, 1871	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Neidium affine</i> (Ehrenberg) Pfizer, 1871	cell	cuboid × 0,85	

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<i>Neidium bisulcatum</i> (Lagerstedt) Cleve, 1894	cell	cuboid \times 0,85	
<i>Neidium hitchcockii</i> (Ehrenberg) Cleve, 1894	cell	cuboid \times 0,8	
<i>Nematodinium</i> Kofoid & Swezy, 1921	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Nematopsides</i> Greuet, 1973	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Nematoradaisia</i> Geitler, 1925	cell	spheroid	
<i>Neocalyptrella</i> D.U.Hernández-Becerril & M.E.Meave del Castillo, 1997	cell	double cone	
<i>Neoceratium</i> F.Gómez, D.Moreira & P.López-García, 2010	cell	Ceratium - girdle diameter (see Annex A)	
<i>Neoceratium arcticum</i> (Ehrenberg) F.Gómez, D.Moreira & P.López-García, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium arietinum</i> (Cleve) F.Gómez, D.Moreira & P.López-García, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium azoricum</i> (Cleve) F.Gómez, D.Moreira & P.López-García, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium breve</i> (Ostenfeld & Schmidt) F.Gómez, D.Moreira & P.López-García, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium candelabrum</i> (Ehrenberg) F.Gómez, D.Moreira & P.López-García, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium carnegiei</i> (Graham & Bronikowsky) F.Gómez, D.Moreira & P.López-García, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium compressum</i> (Gran) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium deflexum</i> (Kofoid) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium extensum</i> (Gourret) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	double cone \times 0,65	
<i>Neoceratium fusus</i> (Ehrenberg) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	double cone \times 0,65	
<i>Neoceratium gibberum</i> (Gourret) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium gravidum</i> (Gourret) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium hexacanthum</i> (Gourret) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium inflatum</i> (Kofoid) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	double cone \times 0,65	
<i>Neoceratium kofoidii</i> (Jørgensen) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	
<i>Neoceratium longirostrum</i> (Gourret) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	double cone \times 0,65	
<i>Neoceratium minutum</i> (Jørgensen) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere \times 0,65	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Neoceratium pentagonum</i> (Gourret) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere × 0,65	
<i>Neoceratium petersii</i> (Steemann Nielsen) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere × 0,65	
<i>Neoceratium setaceum</i> (Jørgensen) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere × 0,65	
<i>Neoceratium symmetricum</i> (Pavillard) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere × 0,65	
<i>Neoceratium teres</i> (Kofoid) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere × 0,65	
<i>Neoceratium trichoceros</i> (Ehrenberg) F.Gomez, D.Moreira & P.Lopez-Garcia, 2010	cell	cone with half sphere × 0,65	
<i>Neochloris</i> Starr, 1955	cell	sphere	
<i>Neocystis</i> F.Hindák, 1988	cell	spheroid	
<i>Neodenticula</i> Akiba & Yanagisawa, 1986	cell	elliptic cylinder	
<i>Neodesmus</i> F.Hindák, 1976	cell	spheroid	
<i>Neonema</i> Pascher, 1925	cell	cylinder	
<i>Neosynedra</i> D.M.Williams & Round, 1986	cell	elliptic cylinder	
<i>Nephrochlamys</i> Korshikov, 1953	cell	spheroid	
<i>Nephrochlamys danica</i> Komárek	cell	spindle	
<i>Nephrocytium</i> Nägeli, 1849	cell	spheroid	
<i>Nephrocytium lunatum</i> West, 1892	cell	spindle	
<i>Nephrوديella</i> Pascher, 1937	cell	spheroid	
<i>Nephroselmis</i> F.Stein, 1878	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Netrium</i> (Nägeli) Itzigsohn & Rothe, 1856	cell	spheroid	
<i>Nitzschia</i> Hassall, 1845	cell	rhombic prism × 1,15	$h=0,50 \times d_2$
<i>Nitzschia abbreviata</i> Hustedt	cell	elliptic cylinder × 1,15	$h=0,50 \times d_2$
<i>Nitzschia acicularis</i> (Kützing) W.Smith, 1853	cell	spindle	
<i>Nitzschia acicularis</i> var. <i>closterioides</i> Grunow	cell	spindle	
<i>Nitzschia angustiforaminata</i> Lange-Bertalot, 1980	cell	elliptic cylinder × 1,15	$h=0,50 \times d_2$
<i>Nitzschia brevissima</i> Grunow, 1881	cell	cuboid × 0,85	$c=0,50 \times b$
<i>Nitzschia calida</i> (Grunow) D.G.Mann, 1990	cell	cuboid × 0,8	$c=0,50 \times b$
<i>Nitzschia clausii</i> Hantzsch, 1860	cell	cuboid × 0,85	$c=0,50 \times b$
<i>Nitzschia denticula</i> Grunow, 1862	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia disputata</i> J.R.Carter, 1971	cell	cuboid × 0,85	$c=0,50 \times b$
<i>Nitzschia dissipata</i> (Kützing) Grunow, 1862	cell	lanceolate cylinder	$h=0,50 \times d_2$
<i>Nitzschia dubia</i> W.Smith, 1853	cell	cuboid × 0,85	$c=0,50 \times b$
<i>Nitzschia epithemoides</i> Grunow, 1880	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia epithemoides</i> var. <i>disputata</i> (J.R.Carter)	cell	elliptic cylinder	$h=0,50 \times d_2$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
Lange-Bertalot, 1987			
<i>Nitzschia flexa</i> Schumann, 1862	cell	cuboid \times 0,9	$c=0,50 \times b$
<i>Nitzschia fonticola</i> (Grunow) Grunow, 1881	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia frigida</i> Grunow, 1880	cell	lanceolate cylinder	$h=0,50 \times d_2$
<i>Nitzschia frustulum</i> (Kützing) Grunow, 1880	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia frustulum</i> var. <i>bulnheimiana</i> (Rabenhorst) Grunow, 1881	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia frustulum</i> var. <i>subsalina</i> Hustedt	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia fruticosa</i> Hustedt	cell	lanceolate cylinder	$h=0,50 \times d_2$
<i>Nitzschia heufleriana</i> Grunow, 1862	cell	cuboid \times 0,85	$c=0,50 \times b$
<i>Nitzschia inconspicua</i> Grunow, 1862	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia intermedia</i> Hantzsch ex Cleve & Grunow, 1880	cell	spindle	
<i>Nitzschia lanceola</i> Grunow	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia lanceola</i> var. <i>minutula</i> Grunow	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia lanceolata</i> W.Smith	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia lanceolata</i> var. <i>lanceolata</i> W.Smith, 1853	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia linearis</i> (C.Agardh) W.Smith, 1853	cell	cuboid \times 0,85	$c=0,50 \times b$
<i>Nitzschia linearis</i> var. <i>subtilis</i> (Grunow) Hustedt, 1923	cell	cuboid \times 0,85	$c=0,50 \times b$
<i>Nitzschia linearis</i> var. <i>tenuis</i> (W.Smith) Grunow, 1880	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia longissima</i> (Brébisson) Ralfs, 1861	cell	spindle	
<i>Nitzschia lorenziana</i> Grunow, 1879	cell	lanceolate cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia microcephala</i> Grunow, 1878	cell	elliptic cylinder \times 0,9	
<i>Nitzschia nana</i> Grunow, 1881	cell	cuboid \times 0,85	$c=0,50 \times b$
<i>Nitzschia paleacea</i> (Grunow) Grunow, 1881	cell	spindle	
<i>Nitzschia parvula</i> W.Smith, 1853	cell	cuboid \times 0,85	$c=0,50 \times b$
<i>Nitzschia perspicua</i> Cholnoky, 1960	cell	elliptic cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia pseudocommunis</i> Hustedt, 1939	cell	elliptic cylinder \times 1,15	$h=0,50 \times d_2$
<i>Nitzschia pusilla</i> (Kützing) Grunow emend.Lange-Bertalot, 1976	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia recta</i> Hantzsch ex Rabenhorst, 1862	cell	cuboid \times 0,8	$c=0,50 \times b$
<i>Nitzschia recta</i> var. <i>robusta</i> Hustedt, 1950	cell	cuboid \times 0,8	$c=0,50 \times b$
<i>Nitzschia regula</i> Hustedt	cell	cuboid \times 0,9	$c=0,50 \times b$
<i>Nitzschia scalpelliformis</i> Grunow, 1880	cell	cuboid \times 0,85	$c=0,50 \times b$

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<i>Nitzschia sigmoidea</i> (Nitzsch) W.Smith, 1853	cell	cuboid × 0,9	$c=0,50 \times b$
<i>Nitzschia sinuata</i> var. <i>sinuata</i> (Thwaites ex W.Smith) Grunow, 1880	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia sublinearis</i> Hustedt, 1930	cell	cuboid × 0,8	$c=0,50 \times b$
<i>Nitzschia tenuis</i> (W.Smith) Grunow, 1880	cell	lanceolate cylinder × 1,15	$h=0,50 \times d_2$
<i>Nitzschia tryblionella</i> Hantzsch, 1860	cell	lanceolate cylinder × 1,15	$h=0,50 \times d_2$
<i>Nitzschia tubicola</i> Grunow, 1880	cell	cuboid × 0,8	$c=0,50 \times b$
<i>Nitzschia valdestriata</i> Aleem & Hustedt, 1951	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Nitzschia vermicularis</i> (Kützing) Hantzsch, 1860	cell	cuboid × 0,9	$c=0,50 \times b$
<i>Nitzschia vitrea</i> G.Norman, 1861	cell	cuboid × 0,9	$c=0,50 \times b$
<i>Noctiluca</i> Suriray, 1836	cell	sphere	
<i>Nodosilinea</i> Perkinson & Casamatta, 2011	filament	cylinder	
<i>Nodularia</i> Mertens ex Bornet & Flahault, 1886	filament	cylinder	
<i>Nostoc</i> Vaucher ex Bornet & Flahault, 1886	cell	cylinder	
<i>Notosolenus</i> Stokes, 1884	cell	ellipsoid	
<i>Nupela</i> Vyverman & Compère, 1991	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Nupela impexiformis</i> (Lange-Bertalot) Lange-Bertalot, 1999	cell	elliptic cylinder × 0,9	
<i>Nupela wellneri</i> (Lange-Bertalot) Lange-Bertalot	cell	elliptic cylinder × 0,9	$h=0,75 \times d_2$
<i>Oblea</i> Balech ex Loeblich Jr. & Loeblich III, 1966	cell	sphere × 0,9	
<i>Ochromonas</i> Vysotskii [Wissotsky], 1887	cell	spheroid	
<i>Ochrosphaera</i> Schussnig, 1930	cell	sphere	
<i>Octacanthium</i> (Hansgirg) P.Compère, 1996	half-cell	spheroid	
<i>Octactis</i> J.Schiller, 1925	cell	sphere	
<i>Odontella</i> C.Agardh, 1832	cell	elliptic cylinder	$d_2=0,50 \times d_1$
<i>Odontella laevis</i> (Ehrenberg) Kützing	cell	cylinder	$h=1,25 \times d$
<i>Odontella longicuris</i> (Greville) Hoban, 1983	cell	cylinder	
<i>Odontella rhombus</i> f. <i>trigona</i> (Cleve ex Van Heurck) R.Ross	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l};$ $h=1,00 \times l$
<i>Odontella sinensis</i> (Greville) Grunow, 1884	cell	elliptic cylinder	$d_2=0,40 \times d_1$
<i>Odontella turgida</i> (Ehrenberg) Kützing, 1844	cell	elliptic cylinder × 0,87	$d_2=0,70 \times d_1$
<i>Oedogonium</i> Link ex Hirn, 1900	filament	cylinder	
<i>Oestrupia</i> Heiden ex Hustedt, 1935	cell	cuboid × 0,85	
<i>Okellya</i> Leliaert & Rueness, 2009	filament	cylinder	
<i>Olisthodiscus</i> N.Carter, 1937	cell	spheroid	
<i>Ollicola</i> Vørs, 1992	cell	spheroid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Oltmannsia</i> Schiller, 1925	cell	spheroid	
<i>Oltmannsiella</i> Zimmermann, 1930	cell	spheroid	
<i>Oltmannsiellopsis</i> M.Cihara & I.Inouye, 1986	cell	spheroid	
<i>Onkonema</i> Geitler, 1933	cell	spheroid	
<i>Oocardium</i> Nägeli, 1849	cell	spheroid	
<i>Oocystis</i> Nägeli ex A.Braun, 1855	cell	spheroid	
<i>Oodinium</i> Chatton, 1912	cell	ellipsoid	
<i>Oolithotus</i> Reinhardt, 1968	cell	sphere	
<i>Oonephris</i> Fott, 1964	cell	spheroid	
<i>Opephora</i> P.Petit, 1889	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Ophiaster</i> Gran, 1912	cell	sphere	
<i>Ophiocytium</i> Nägeli, 1849	cell	cylinder	
<i>Opisthoaulax</i> Calado, 2011	cell	ellipsoid	
<i>Ornithocercus</i> Stein, 1883	cell	sphere	
<i>Orthoseira</i> Thwaites, 1848	cell	cylinder	
<i>Oscillatoria</i> Vaucher ex Gomont, 1892	filament	cylinder	
<i>Ostreococcus</i> C.Courties & M.-J.Chrétiennot-Dinet, 1995	cell	spheroid	
<i>Ostreopsis</i> J.Schmidt, 1901	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Oxyneis</i> Round, 1990	cell	elliptic cylinder $\times 0,85$	$d_2=0,33 \times h; h=3,00 \times d_2$
<i>Oxyrrhis</i> Dujardin, 1841	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Oxytoxum</i> Stein, 1883	cell	spindle	
<i>Oxytoxum criophilum</i> Balech, 1965	cell	double cone	
<i>Oxytoxum gracile</i> Schiller, 1937	cell	double cone	
<i>Oxytoxum laticeps</i> Schiller, 1937	cell	spheroid	
<i>Oxytoxum sphaeroideum</i> Stein, 1883	cell	spheroid	
<i>Pachysoeca</i> Ellis, 1933	cell	spheroid	
<i>Pachysphaera</i> Ostenfeld, 1899	cell	sphere $\times 0,8$	
<i>Palaeophalacroma</i> Schiller, 1928	cell	spheroid	
<i>Palatinus</i> S.C.Craveiro, A.J.Calado, N.Daugbjerg & Moestrup, 2009	cell	ellipsoid	
<i>Palatinus pseudolaevis</i> (M.Lefèvre) S.C.Craveiro, A.J.Calado, N.Daugbjerg & Ø.Moestrup, 2009	cell	spheroid	
<i>Palikiella</i> G.Claus, 1962	filament	cylinder	
<i>Palmella</i> Lyngbye, 1819	cell	spheroid	
<i>Palmellopsis</i> Korshikov, 1953	cell	sphere	
<i>Palmeria</i> Greville, 1865	cell	elliptic cylinder $\times 0,6$	$d_2=1,43 \times h; h=0,70 \times d_2$
<i>Palmodictyon</i> Kützing, 1845	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Palusphaera</i> Lecal, 1966	cell	sphere	
<i>Pandorina</i> Bory de Saint-Vincent, 1824	cell	cone with half sphere	
<i>Pannus</i> B.Hickel, 1991	cell	sphere	
<i>Papenfussiomonas</i> Desikachary, 1972	cell	spheroid	
<i>Papiliocellulus</i> G.R.Hasle, H.A.von Stosch & E.E.Syvetsen, 1983	cell	elliptic cylinder	
<i>Pappomonas</i> I.Manton & K.Oates, 1975	cell	sphere	
<i>Papposphaera</i> K.Tangen, 1972	cell	spheroid	
<i>Paracapsa</i> Naumann, 1924	cell	spheroid	
<i>Parachrysidalis</i> Hulbert, 1965	cell	spheroid	
<i>Paradoxia</i> Svirenko, 1928	cell	spindle	
<i>Paralia</i> Heiberg, 1863	cell	cylinder	
<i>Paramastix</i> Skuja, 1948	cell	spheroid	
<i>Parapediastrum</i> E.Hegewald, 2005	cell	cuboid × 0,67	$c=1,00 \times b$
<i>Parapediastrum biradiatum</i> (Meyen) E.Hegewald, 2005	cell	cuboid × 0,67	$c=1,00 \times b$
<i>Parapedinella</i> S.M.Pedersen, P.L.Beech & H.A.Thomsen, 1986	cell	sphere	
<i>Paraphysomonas</i> De Saedeleer, 1930	cell	sphere	
<i>Parlibellus</i> E.J.Cox, 1988	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Parlibellus rhombicus</i> (Gregory) E.J.Cox	cell	rhombic prism	
<i>Parvicorbicula</i> Deflandre, 1960	cell	spheroid	
<i>Pascherina</i> P.C.Silva, 1959	cell	spheroid	
<i>Pascherinema</i> De Toni, 1936	filament	cylinder	
<i>Pauliella</i> F.E.Round & P.W.Basson, 1997	cell	elliptic cylinder	
<i>Paulinella</i> Lauterborn, 1895	cell	spheroid	
<i>Paulschulzia</i> Skuja, 1948	cell	sphere	
<i>Pavlova</i> Butcher, 1952	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Pectinodesmus</i> Hegewald, Wolf, Keller, Friedl & Krienitz, 2010	cell	spheroid	
<i>Pediastrum</i> Meyen, 1829	cell	cuboid × 0,67	$c=1,00 \times b$
<i>Pediastrum simplex</i> var. <i>biwaense</i> Fukushima	cell	triangular prism × 0,67	$h=1,00 \times l$
<i>Pedinella</i> Vysotskij, 1887	cell	sphere	
<i>Pedinomonas</i> Korshikov, 1923	cell	ellipsoid	
<i>Pelagococcus</i> R.E.Norris, 1977	cell	sphere	
<i>Penium</i> Brébisson ex Ralfs, 1848	cell	cylinder	
<i>Pentapharsodinium</i> Indelicato & Loeblich III, 1986	cell	cone with half sphere	
<i>Peranema</i> Dujardin, 1841	cell	ellipsoid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Peranemopsis</i> Lackey, 1940	cell	spheroid	
<i>Peridiniella</i> Kofoid & Michener, 1911	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Peridiniella catenata</i> (Levander) Balech, 1977	cell	sphere $\times 0,5$	
<i>Peridiniopsis</i> Lemmermann, 1904	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Peridiniopsis cunningtonii</i> Lemmermann, 1907	cell	cone with half sphere	
<i>Peridinium</i> Ehrenberg, 1832	cell	cone with half sphere	$h=1,20 \times d$
<i>Peridinium aciculiferum</i> Lemmermann, 1900	cell	cone with half sphere $\times 0,9$	
<i>Peridinium aciculiferum</i> f. <i>inerme</i> J.Woloszynska	cell	cone with half sphere $\times 0,9$	
<i>Peridinium africanum</i> Lemmermann, 1907	cell	ellipsoid	
<i>Peridinium bipes</i> F.Stein, 1883	cell	cone $\times 0,5$	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Peridinium cinctum</i> (O.F.Müller) Ehrenberg, 1832	cell	sphere	
<i>Peridinium gatunense</i> Nygaard, 1925	cell	spheroid	
<i>Peridinium lomnickii</i> Woloszynska, 1916	cell	ellipsoid	
<i>Peridinium lomnickii</i> var. <i>wierzejskii</i> (Woloszynska) Lindemann, 1928	cell	spheroid	
<i>Peridinium morzinense</i> Lefèvre, 1928	cell	ellipsoid	
<i>Peridinium quinquecorne</i> Abé, 1927	cell	cone with half sphere	$h=1,22 \times d$
<i>Peridinium steinii</i> f. <i>pyriformis</i> O.W.Paulsen	cell	cone with half sphere	$h=1,35 \times d$
<i>Peridinium umbonatum</i> F.Stein, 1883	cell	ellipsoid	$d_2=0,65 \times d_1$
<i>Peridinium umbonatum</i> var. <i>goslaviense</i> (Woloszynska) J.Popovsky & L.Pfeister, 1986	cell	ellipsoid	
<i>Peridinium willei</i> Huitfeldt-Kaas, 1900	cell	ellipsoid	$d_2=0,85 \times d_1$
<i>Periphylophora</i> Kamptner, 1936	cell	sphere	
<i>Perone</i> Pascher, 1932	cell	spheroid	
<i>Peronia</i> Fleming, 1822	cell	pyramid $\times 0,9$	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Peroniella</i> Gobi, 1887	cell	sphere	
<i>Petalomonas</i> F.Stein, 1859	cell	ellipsoid	$d_2=0,80 \times d_1$
<i>Petalonema</i> Berkeley ex Kirchner, 1898	filament	cylinder	
<i>Petrodictyon</i> D.G.Mann, 1990	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Petroneis</i> A.J.Stickle & D.G.Mann, 1990	cell	elliptic cylinder	$h=0,50 \times d_2$
<i>Pfiesteria</i> K.A.Steidinger & J.M.Burkholder, 1996	cell	sphere	
<i>Phacotus</i> Perty, 1852	cell	ellipsoid	$d_2=0,65 \times d_1$
<i>Phacotus glaber</i> Playfair	cell	spheroid	
<i>Phacus</i> Dujardin, 1841	cell	ellipsoid	$d_2=0,30 \times d_1$
<i>Phacus longicauda</i> (Ehrenberg) Dujardin, 1841	cell	ellipsoid	$d_2=0,17 \times d_1$
<i>Phacus pleuronectes</i> (O.F.Müller) Nitzsch ex Dujardin,	cell	ellipsoid	$d_2=0,14 \times d_1$

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1841			
<i>Phaeaster</i> Scherffel, 1927	cell	ellipsoid	
<i>Phaeobotrys</i> H.Ettl, 1966	cell	sphere	
<i>Phaeocystis</i> Lagerheim, 1893	cell	sphere	
<i>Phaeocystis globosa</i> Scherffel, 1899	cell	sphere × 0,9	
<i>Phaeodactylum</i> Bohlin, 1897	cell	spindle	
<i>Phaeodermatium</i> Hansgirg, 1889	cell	spheroid	
<i>Phaeoplaca</i> Chodat, 1926	cell	spheroid	
<i>Phaeoschizochlamys</i> Lemmermann, 1898	cell	spheroid	
<i>Phalacroma</i> Stein, 1883	cell	ellipsoid	$d_2=0,28 \times d_1$
<i>Pheopolykrikos</i> Chatton, 1933	colony	cylinder	
<i>Phormidiochaete</i> Komárek, 2001	filament	cylinder	
<i>Phormidium</i> Kützing ex Gomont, 1892	filament	cylinder	
<i>Phyllomitus</i> Stein	cell	spheroid	
<i>Phytodinium</i> Klebs, 1912	cell	spheroid	
<i>Picochlorum</i> W.J.Henley, J.L.Hironaka, L.Guillou, M.A.Buchheim, J.A.Buchheim, M.W.Fawley & K.P.Fawley, 2004	cell	spheroid	
<i>Pilgeria</i> Schmidle, 1901	cell	spheroid	
<i>Pinnularia</i> Ehrenberg, 1843	cell	cuboid × 0,9	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia balfouriana</i> (Grunow ex Cleve) Krammer & Lange-Bertalot, 2000	cell	elliptic cylinder	
<i>Pinnularia bertrandii</i> K.Krammer, 2000	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia bullacostae</i> K.Krammer & Lange-Bertalot, 1999	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia clevei</i> R.M.Patrick, 1945	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia clevei</i> var. <i>minor</i> (Hustedt) K.Krammer, 2000	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia crucifera</i> Cleve-Euler, 1934	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia divergens</i> var. <i>sublinearis</i> Cleve, 1895	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia elegans</i> (W.Smith) K.Krammer, 1992	cell	elliptic cylinder	$d_2=1,11 \times h; h=0,90 \times d_2$
<i>Pinnularia ivaloensis</i> K.Krammer, 2000	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia krammeri</i> Metzeltin, 1996	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia lange-bertalotii</i> K.Krammer, 1992	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia leptosoma</i> (Grunow) Krammer, 1985	cell	elliptic cylinder × 0,9	
<i>Pinnularia lokana</i> Krammer, 2000	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia polaris</i> H.Heiden	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$
<i>Pinnularia polyonca</i> (Brébisson) W.Smith, 1856	cell	elliptic cylinder × 0,9	
<i>Pinnularia septentrionalis</i> K.Krammer, 2000	cell	cuboid	$b=1,11 \times c; c=0,90 \times b$

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<i>Pinnularia tirolensis</i> var. <i>julma</i> Krammer, 2000	cell	cuboid	$b=1,11 \times c$; $c=0,90 \times b$
<i>Pinnunavis</i> H.Okuno, 1975	cell	elliptic cylinder $\times 1,15$	$d_2=1,11 \times h$; $h=0,90 \times d_2$
<i>Pinnunavis genustriata</i> (Hustedt) Lange-Bertalot & K.Krammer	cell	rhombic prism $\times 1,15$	$d_2=1,11 \times h$; $h=0,90 \times d_2$
<i>Placoma</i> Schousboe ex Bornet & Thuret, 1876	cell	spheroid	
<i>Placoneis</i> Mereschkowsky, 1903	cell	elliptic cylinder $\times 0,9$	$h=0,75 \times d_2$
<i>Placoneis abiskoensis</i> (Hustedt) Lange-Bertalot & D.Metzeltin, 1996	cell	cuboid $\times 0,85$	$c=1,00 \times b$
<i>Placoneis gastrum</i> (Ehrenberg) Mereschkovsky, 1903	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Plagiodiscus</i> Grunow & Eulenstein, 1867	cell	elliptic cylinder	
<i>Plagiogramma</i> Greville, 1859	cell	elliptic cylinder	$d_2=0,40 \times d_1$
<i>Plagiogrammopsis</i> Hasle, von Stosch & Syvertsen, 1983	cell	rhombic prism $\times 0,9$	$d_2=0,20 \times d_1$
<i>Plagioselmis</i> Butcher ex G.Novarino, I.A.N.Lucas & S.Morrall, 1994	cell	cone with half sphere	
<i>Plagiotropis</i> Pfitzer, 1871	cell	rhombic prism $\times 1,15$	
<i>Planctococcus</i> Korshikov, 1953	cell	sphere	
<i>Planctomyces</i>	cell	spheroid	
<i>Planctonema</i> Schmidle, 1903	cell	cylinder $\times 0,9$	
<i>Planktolynghya</i> Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Planktoniella</i> F.Schütt, 1892	cell	cylinder	$h=0,80 \times d$
<i>Planktosphaeria</i> G.M.Smith, 1918	cell	sphere	
<i>Planktothrix</i> Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Planonephros</i> Christensen, 1978	cell	spheroid	
<i>Planophila</i> Gerneck, 1907	cell	cone with half sphere	
<i>Planothidium</i> Round & L.Bukhtiyarova, 1996	cell	elliptic cylinder	
<i>Planothidium dubium</i> (Grunow) Round & Bukhtiyarova, 1996	cell	elliptic cylinder $\times 0,9$	
<i>Planothidium haynaldii</i> (Schaarschmidt) Lange-Bertalot, 1999	cell	elliptic cylinder $\times 0,9$	
<i>Planothidium peragalloi</i> (J.Brun & Héribaud-Joseph) Round & L.Bukhtiyarova, 1996	cell	elliptic cylinder $\times 0,9$	
<i>Planothidium rostratum</i> (Oestrup) Lange-Bertalot, 1999	cell	elliptic cylinder $\times 0,9$	
<i>Platessa</i> Lange-Bertalot, 2004	cell	elliptic cylinder	
<i>Plectonema</i> Thuret ex Gomont, 1892	filament	cylinder	
<i>Pleodorina</i> W.R.Shaw, 1894	cell	sphere	
<i>Pleromonas</i> Pascher, 1914	cell	spheroid	
<i>Pleurasiga</i> J.Schiller, 1925	cell	spheroid	
<i>Pleurocapsa</i> Thuret, 1885	cell	spheroid	

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<i>Pleurochloris</i> Pascher, 1925	cell	sphere	
<i>Pleurochrysis</i> Pringsheim, 1955	cell	sphere	
<i>Pleurosigma</i> W.Smith, 1852	cell	rhombic prism	$h=0,75 \times d_2$
<i>Pleurosira</i> (Meneghini) Trevisan, 1848	cell	elliptic cylinder	$d_2=0,85 \times d_1$; $h=1,00 \times d_2$
<i>Pleurosira laevis</i> f. <i>polymorpha</i> (Kützing) Compère, 1982	cell	cylinder	
<i>Pleurotaenium</i> Nägeli, 1849	cell	cylinder	
<i>Podocapsa</i> Ercegovic, 1931	cell	spheroid	
<i>Podocystis</i> J.W.Bailey, 1854	cell	elliptic cylinder	
<i>Podohedra</i> Düringer, 1958	cell	spindle	
<i>Podolampas</i> Stein, 1883	cell	cone with half sphere	
<i>Podosira</i> Ehrenberg, 1840	cell	cylinder × 0,83	
<i>Polyblepharides</i> P.-A.Dangeard, 1888	cell	spheroid	
<i>Polyedriella</i> Pascher, 1930	cell	sphere	
<i>Polyedriopsis</i> Schmidle, 1899	cell	tetrahedron	
<i>Polyfibula</i> Manton & Bremer, 1981	cell	spheroid	
<i>Polygoniochloris</i> Ettl, 1977	cell	cuboid × 0,52	$c=0,50 \times a$
<i>Polygoniochloris circularis</i> (Bourrelly & Georges) H.Ettl, 1965	cell	cylinder × 0,52	$h=0,50 \times d$
<i>Polykrikos</i> Bütschli, 1873	colony	spheroid	
<i>Polykrikos lebourae</i> Herdman, 1924	colony	ellipsoid	$d_2=0,50 \times d_1$
<i>Polyepidomonas</i> Preisig & D.J.Hibberd, 1983	cell	sphere	
<i>Polyoeca</i> Kent, 1880	cell	spheroid	
<i>Polytoma</i> Ehrenberg, 1831	cell	spheroid	
<i>Polytomella</i> Aragão, 1910	cell	spheroid	
<i>Pomatodinium</i> J.Cachon & Cachon-Enjumet, 1966	cell	sphere	
<i>Pontosphaera</i> Lohmann, 1902	cell	sphere	
<i>Porosira</i> Jörgensen, 1905	cell	cylinder	$h=0,60 \times d$
<i>Porotheca</i> P.C.Silva, 1960	cell	spheroid	$h=0,60 \times d$
<i>Porphyrosiphon</i> Kützing ex Gomont, 1892	filament	cylinder	
<i>Prasinocladus</i> Kuckuck, 1894	cell	spheroid	
<i>Preperidinium</i> Mangin, 1913	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Proboscia</i> Sundström, 1986	cell	cylinder	
<i>Prochlorothrix</i> T.Burger-Wiersma, L.J.Stal & L.R.Mur, 1989	filament	cylinder	
<i>Prodinophysis</i> Balech, 1944	cell	ellipsoid	$d_2=0,28 \times d_1$
<i>Progonoia</i> H.-J.Schrader, 1969	cell	elliptic cylinder	
<i>Pronoctiluca</i> Fabre-Domergue, 1889	cell	ellipsoid	$d_2=0,85 \times d_1$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Pronoctiluca phaeocysticola</i> (Scherffel) Pavillard, 1922	cell	spindle	
<i>Properidinium</i> Meunier, 1919	cell	double cone	
<i>Prorocentrum</i> Ehrenberg, 1834	cell	ellipsoid	$d_2=0,50 \times d_1$
<i>Prorocentrum aporum</i> (Schiller) Dodge, 1975	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Prorocentrum balticum</i> (Lohmann) Loeblich, 1970	cell	sphere $\times 0,9$	
<i>Prorocentrum cassubicum</i> (Woloszynska) Dodge, 1975	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Prorocentrum cordatum</i> (Ostenfeld) Dodge, 1975	cell	ellipsoid	$d_2=0,65 \times d_1$
<i>Prorocentrum dentatum</i> Stein, 1883	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Prorocentrum gracile</i> Schütt, 1895	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Prorocentrum lima</i> (Ehrenberg) F.Stein, 1878	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Prorocentrum nanum</i> J.Schiller, 1918	cell	spheroid	
<i>Prorocentrum scutellum</i> Schröder, 1900	cell	ellipsoid	$d_2=0,70 \times d_1$
<i>Proschkinia</i> N.I.Karajeva, 1978	cell	elliptic cylinder $\times 1,15$	$h=1,00 \times d_2$
<i>Proschkinia complanatula</i> (Hust ex Simonsen) Mann, 1990	cell	rhombic prism $\times 1,15$	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Prosoaulax</i> (F.Stein) Calado & Moestrup, 2005	cell	ellipsoid	
<i>Protaspis</i> Skuja, 1939	cell	ellipsoid	
<i>Proterendothrix</i> West & G.S.West, 1897	filament	cylinder	
<i>Proterospongia</i> Kent, 1882	cell	spheroid	
<i>Proterythropsis</i> Kofoid & Swezy, 1920	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Protoceratium</i> Bergh, 1881	cell	sphere $\times 0,9$	
<i>Protoderma</i> Kützing, 1843	cell	spheroid	
<i>Protodinium</i> Lohmann, 198.	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Protopteridinium</i> Bergh, 1882	cell	cone with half sphere	$h=1,20 \times d$
<i>Protopteridinium achromaticum</i> (Levander) Balech, 1974	cell	sphere $\times 0,9$	
<i>Protopteridinium avellana</i> (Meunier) Balech, 1974	cell	ellipsoid	
<i>Protopteridinium bipes</i> (Paulsen) Balech, 1974	cell	cone $\times 0,5$	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Protopteridinium breve</i> Paulsen	cell	sphere $\times 0,9$	
<i>Protopteridinium brochii</i> (Kofoid & Swezy) Balech	cell	double cone	
<i>Protopteridinium cerasus</i> (Paulsen) Balech, 1973	cell	cone with half sphere	$h=1,00 \times d$
<i>Protopteridinium claudicans</i> (Paulsen) Balech, 1974	cell	ellipsoid	$d_2=0,65 \times d_1$
<i>Protopteridinium crassipes</i> (Kofoid) Balech, 1974	cell	double cone $\times 0,7$	
<i>Protopteridinium curtipes</i> (Jørgensen) Balech, 1974	cell	double cone $\times 0,7$	
<i>Protopteridinium curvipes</i> (Ostenfeld) Balech, 1974	cell	cone with half sphere	$h=1,00 \times d$
<i>Protopteridinium decipiens</i> (Jørgensen) Parke & Dodge, 1976	cell	spheroid	$h=0,60 \times d$
<i>Protopteridinium defectum</i> (Balech) Balech, 1974	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Protopteridinium denticulatum</i> (Gran & Braarud) Balech, 1974	cell	ellipsoid × 0,8	$d_2=0,75 \times d_1$
<i>Protopteridinium depressum</i> (Bailey) Balech, 1974	cell	cone with half sphere × 0,8	$h=1,10 \times d$
<i>Protopteridinium divergens</i> (Ehrenberg) Balech, 1974	cell	cone with half sphere × 0,8	$h=1,20 \times d$
<i>Protopteridinium excentricum</i> (Paulsen) Balech, 1974	cell	double cone	
<i>Protopteridinium globulus</i> (Stein, 1883) Balech, 1974	cell	sphere	
<i>Protopteridinium granii</i> (Ostenfeld) Balech, 1974	cell	cone with half sphere × 0,8	$h=1,00 \times d$
<i>Protopteridinium islandicum</i> (Paulsen) Balech, 1973	cell	spheroid	
<i>Protopteridinium laticeps</i> (Grontved & Seidenfaden) Balech, 1974	cell	cone with half sphere × 0,8	
<i>Protopteridinium leonis</i> (Pavillard) Balech, 1974	cell	double cone × 0,7	
<i>Protopteridinium longispinum</i> (Kofoid) Balech, 1974	cell	cone with half sphere × 0,8	$h=1,20 \times d$
<i>Protopteridinium marie-lebouriae</i> (Paulsen) Balech, 1974	cell	double cone × 0,7	
<i>Protopteridinium mite</i> (Pavillard) Balech, 1974	cell	cone with half sphere	$h=1,00 \times d$
<i>Protopteridinium oblongum</i> (Aurivillius) Parke & Dodge, 1976	cell	cone with half sphere × 0,8	$h=1,40 \times d$
<i>Protopteridinium oceanicum</i> (VanHöffen) Balech, 1974	cell	cone with half sphere × 0,8	$h=1,45 \times d$
<i>Protopteridinium ovatum</i> Pouchet, 1883	cell	ellipsoid	$d_2=0,85 \times d_1$; $h=0,80 \times d_1$
<i>Protopteridinium pallidum</i> (Ostenfeld) Balech, 1973	cell	cone with half sphere × 0,8	$h=1,33 \times d$
<i>Protopteridinium pentagonum</i> (Gran) Balech, 1974	cell	double cone × 0,7	
<i>Protopteridinium punctulatum</i> (Paulsen) Balech, 1974	cell	cone with half sphere × 0,75	$h=1,05 \times d$
<i>Protopteridinium pyriforme</i> (Paulsen) Balech, 1974	cell	cone with half sphere	$h=1,25 \times d$
<i>Protopteridinium quarnerense</i> (Schröder) Balech, 1974	cell	ellipsoid	
<i>Protopteridinium roseum</i> (Paulsen) Balech, 1974	cell	spheroid	
<i>Protopteridinium steinii</i> (Jørgensen) Balech, 1974	cell	cone with half sphere × 0,75	$h=1,35 \times d$
<i>Protopteridinium subcurvipes</i> (Lebour) Balech, 1974	cell	ellipsoid	
<i>Protopteridinium subinerme</i> (Paulsen) Loeblich III, 1969	cell	cone with half sphere × 0,75	$h=0,90 \times d$
<i>Protopteridinium thorianum</i> (Paulsen) Balech, 1974	cell	spheroid × 0,8	
<i>Protopteridinium thulesense</i> (Balech) Balech, 1974	cell	cone with half sphere	$h=0,80 \times d$
<i>Protopsis</i> Kofoid & Swezy, 1921	cell	spheroid	
<i>Provasoliella</i> A.R.Loeblich III, 1967	cell	spheroid	
<i>Prymnesium</i> Massart, 1920	cell	spheroid	

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<i>Psammodictyon</i> D.G.Mann, 1990	cell	elliptic cylinder	
<i>Psammodiscus</i> Round & D.G.Mann, 1980	cell	cylinder	
<i>Psammothidium</i> L.Buhtkiyarova & Round, 1996	cell	elliptic cylinder	
<i>Pseudanabaena</i> Lauterborn, 1915	filament	cylinder	
<i>Pseudendozonium</i> Wille, 1901	filament	cylinder	
<i>Pseudoactiniscus</i> Bursa, 1969	cell	sphere	
<i>Pseudobodo</i> Greissmann, 1913	cell	spheroid	
<i>Pseudocapsa</i> Ercegovic, 1925	cell	spheroid	
<i>Pseudocarteria</i> H.Ettl, 1958	cell	spheroid	
<i>Pseudocharaciopsis</i> K.W.Lee & Bold, 1973	cell	spindle	
<i>Pseudocharacium</i> Korshikov, 1953	cell	spindle	
<i>Pseudochattonella</i> (Y.Hara & Chihara) Hosoi-Tanabe, Honda, Fukaya, Inagaki & Sako, 2007	cell	spheroid $\times 0,88$	
<i>Pseudochattonella verruculosa</i> (Y.Hara & M.Chihara) S.Tanabe-Hosoi, D.Honda, S.Fukaya, Y.Inagaki & Y.Sako, 2007	cell	ellipsoid	$d_2=0,85 \times d_1$
<i>Pseudococcomyxa</i> Korshikov, 1953	cell	spheroid	
<i>Pseudodictyosphaerium jurisii</i> (Hindák) Krienitz, C.Bock, Dadheech & Proschold, 2011	cell	sphere	
<i>Pseudodidymocystis</i> Hegewald & Deason, 1989	cell	spheroid	
<i>Pseudogomphonema</i> L.K.Medlin, 1986	cell	pyramid $\times 0,9$	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Pseudogoniochloris</i> L.Krienitz, E.Hegewald, O.L.Reymond & T.Peschke, 1993	cell part	cuboid $\times 0,67$	$c=1,00 \times b$
<i>Pseudoguinaridia</i> von Stosch, 1986	cell	cylinder	
<i>Pseudohimantidium</i> Hustedt & Krasske, 1941	cell	elliptic cylinder $\times 0,65$	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Pseudokephyrion</i> Pascher, 1913	cell	spheroid	
<i>Pseudokirchneriella contorta</i> (Schmidle) F.Hindák, 1990	cell	cylinder	
<i>Pseudoncobyrsa</i> Geitler, 1925	cell	spheroid	
<i>Pseudo-nitzschia</i> H.Peragallo, 1900	cell	rhombic prism $\times 0,7$	$d_2=1,33 \times h; h=0,75 \times d_2$
<i>Pseudopediastrum</i> E.Hegewald, 2005	cell	cuboid $\times 0,67$	$c=1,00 \times b$
<i>Pseudopedinella</i> N.Carter, 1937	cell	sphere	
<i>Pseudopedinella erkensis</i> Skuja, 1948	cell	cylinder $\times 0,8$	
<i>Pseudopfiesteria</i> Litaker, Steidinger, Mason, Shields & Tester, 2005	cell	sphere	
<i>Pseudophormidium</i> (Forti) Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Pseudopodosira</i> A.P.José, 1949	cell	spheroid	$h=0,80 \times d$
<i>Pseudopolyedriopsis</i> Gollerbach, 1962	half-cell	tetrahedron	
<i>Pseudoquadrigula</i> E.N.Lacoste de Díaz, 1973	cell	spindle	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Pseudoschroederia</i> Hegewald & Schnepf, 1986	cell	spindle	
<i>Pseudoscourfieldia</i> I.Manton, 1975	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Pseudoscytonema</i> Elenkin, 1949	filament	cylinder	
<i>Pseudosolenia</i> B.G.Sundström, 1986	cell	cylinder	
<i>Pseudosphaerocystis</i> Woronichin, 1931	cell	spheroid	
<i>Pseudostaurastrum</i> R.Chodat, 1921	half-cell	tetrahedron	
<i>Pseudostaurosira</i> D.M.Williams & Round, 1988	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Pseudotetraedriella</i> Hedgewald, Padisak & Friedl, 2007	cell	tetrahedron	
<i>Pseudotetrastrum</i> Hindák, 1977	cell	cone with half sphere	
<i>Pseudotriceratium</i> Grunow, 1884	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$
<i>Pteridomonas</i> Penard, 1889	cell	sphere	
<i>Pteromonas</i> Seligo, 1887	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Pteroncola</i> R.W.Holmes & D.A.Croll, 1984	cell	elliptic cylinder	
<i>Pterosperma</i> Pochet, 1893	cell	sphere	
<i>Ptychodiscus</i> Stein, 1883	cell	ellipsoid	
<i>Pulchella</i> Krammer, 2000	cell	elliptic cylinder	
<i>Pulvinularia</i> Borzi, 1916	cell	cylinder	
<i>Punctastriata</i> D.M.Williams & Round, 1988	cell	elliptic cylinder	
<i>Puncticulata</i> H.Håkansson	cell	cylinder	
<i>Pycnococcus</i> R.R.L.Guillard, 1991	cell	sphere	
<i>Pyramichlamys</i> H.Ettl & O.Ettl, 1959	cell	ellipsoid	
<i>Pyramidochrysis</i> Pascher, 1909	cell	cone with half sphere	
<i>Pyramimonas</i> Schmarda, 1849	cell	spheroid	
<i>Pyramimonas longicauda</i> L.Van Meel, 1969	cell	cone	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Pyramimonas tertrarhynchus</i> Schmarda	cell	cone	$z=\sqrt{(s^2-0,25 \times d^2)}$
<i>Pyrobotrys</i> Arnoldi, 1916	cell	spheroid × 0,88	
<i>Pyrocystis</i> J.Murray ex Haeckel, 1890	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Pyrocystis fusiformis</i> (Wyville Thomson ex Haeckel) Blackman, 1902	cell	spindle	
<i>Pyrocystis lunula</i> (J.Schütt) J.Schütt, 1896	sporangium	spindle	
<i>Pyrocystis noctiluca</i> Murray ex Haeckel, 1890	sporangium	spindle	
<i>Pyrodinium</i> Plate, 1906	cell	ellipsoid	
<i>Pyrophacus</i> Stein, 1883	cell	ellipsoid × 0,8	$d_2=0,95 \times d_1; h=0,53 \times d_2$
<i>Quadrichloris</i> Fott, 1959	cell	spheroid	
<i>Quadricoccus</i> Fott, 1948	cell	spheroid	
<i>Quadrigula</i> Printz, 1916	cell	spindle	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Raciborskiella</i> S.Wislouch, 1924	cell	spheroid	
<i>Radaisia</i> Sauvageau, 1895	cell	spheroid	
<i>Radaisiella</i> Geitler, 1925	cell	spheroid	
<i>Radiococcus</i> Schmidle, 1902	cell	sphere	
<i>Radiocystis</i> Skuja, 1948	cell	sphere	
<i>Radiocystis geminata</i> Skuja, 1948	cell	spheroid	
<i>Radiofilum</i> Schmidle, 1894	cell	sphere	
<i>Radiosphaera</i> J.Snow, 1918	cell	sphere	
<i>Raphidiopsis</i> F.E.Fritsch & F.Rich, 1929	filament	cylinder	
<i>Raphidocelis</i> Hindák, 1977	cell	spindle	
<i>Raphidonema</i> Lagerheim, 1892	cell	spindle	
<i>Raphidosphaera</i> (Pascher) P.C.Silva, 1979	cell	sphere	
<i>Rayssiella</i> Edelstein & Prescott, 1964	cell	ellipsoid	
<i>Rebecca</i> J.C.Green, 2000	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Reimeria</i> J.P.Kociolek & E.F.Stoermer, 1987	cell	elliptic cylinder	
<i>Resultor</i> Moestrup, 1991	cell	sphere	
<i>Rhabdoderma</i> Schmidle & Lauterborn, 1900	cell	cylinder	
<i>Rhabdogloea</i> Schröder, 1917	cell	cylinder	
<i>Rhabdolithes</i> O.Schmidt, 1870	cell	sphere	
<i>Rhabdomonas</i> Fresenius, 1858	cell	spheroid	
<i>Rhabdomonas costata</i> (Korsikov) Pringsheim, 1942	cell	spindle	
<i>Rhabdonema</i> Kützing, 1844	cell	elliptic cylinder	
<i>Rhaphoneis</i> Ehrenberg, 1844	cell	elliptic cylinder	$h=0,33 \times d_2$
<i>Rhinomonas</i> D.R.A.Hill & Wetherbee, 1988	cell	ellipsoid	
<i>Rhipidodendron</i> F.Stein, 1878	cell	sphere	
<i>Rhizochloris</i> Pascher, 1917	cell	miscellaneous (amoeboid)	
<i>Rhizochrysis</i> Pascher, 1913	cell	spheroid	
<i>Rhizoclonium</i> Kützing, 1843	filament	cylinder	
<i>Rhizosolenia</i> Brightwell, 1858	cell	cylinder	
<i>Rhizosolenia imbricata</i> Brightwell, 1858	cell	cylinder $\times 0,9$	
<i>Rhizosolenia longiseta</i> O.Zacharias, 1893	cell	elliptic cylinder	$d_2=0,90 \times d_1$
<i>Rhodomonas</i> Karsten, 1898	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Rhodomonas baltica</i> Karsten, 1898	cell	cone with half sphere	
<i>Rhodomonas lacustris</i> Pascher & Ruttner, 1913	cell	cone with half sphere	$h=0,90 \times d$
<i>Rhodomonas marina</i> (P.A.Dangeard) Lemmermann, 1899	cell	cone with half sphere	

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<i>Rhodomonas salina</i> (Wislouch) D.R.A.Hill & R.Wetherbee, 1989	cell	cone with half sphere	
<i>Rhodoplax</i> Schmidle & Wellheim, 1901	cell	spheroid	
<i>Rhodostichus</i> Geitler & Pascher, 1931	cell	spheroid	
<i>Rhoicosphenia</i> Grunow, 1860	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Rhoicosphenia abbreviata</i> (C.Agardh) Lange-Bertalot, 1980	cell	elliptic cylinder	$d_2=0,75 \times h; h=1,33 \times d_2$
<i>Rhoikoneis</i> Grunow, 1863	cell	elliptic cylinder	
<i>Rhopalodia</i> Otto Müller, 1895	cell	elliptic cylinder × 0,65	$d_2=0,40 \times h; h=2,50 \times d_2$
<i>Rhynchomonas</i> Klebs, 1892	cell	spheroid	
<i>Richelia</i> J.Schmidt, 1901	filament	cylinder	
<i>Rivularia</i> C.Agardh ex Bornet & Flahault, 1886	filament	cylinder	
<i>Romeria</i> Koczwara, 1932	cell	cylinder	
<i>Roperia</i> Grunow ex Pelletan, 1889	cell	cylinder	$h=0,50 \times d$
<i>Roscoffia</i> Balech, 1956	cell	ellipsoid	
<i>Rosenvingiella</i> P.C.Silva, 1957	filament	cylinder	
<i>Rossithidium</i> Round & L.Bukhtiyarova, 1996	cell	elliptic cylinder	
<i>Roya</i> West & G.S.West, 1896	cell	cylinder	
<i>Rutilaria</i> Greville, 1863	cell	rhombic prism	
<i>Sacchochrysis</i> Korshikov, 1941	cell	spheroid	
<i>Sacconema</i> Borzi ex Bornet & Flahault, 1886	filament	cylinder	
<i>Saepicula</i> Leadbeater, 1980	cell	spheroid	
<i>Salpingoeca</i> H.J.Clark, 1866	cell	spheroid	
<i>Sarcinochrysis</i> Geitler, 1930	cell	sphere	
<i>Saroeca</i> H.A.Thomsen, 1979	cell	spheroid	
<i>Savillea</i> (W.N. Ellis) Loeblich III, 1967	cell	spheroid	
<i>Scenedesmus</i> Meyen, 1829	cell	spheroid	
<i>Scenedesmus acutus</i> f. <i>tetradesmiformis</i> (Wolosz.) Uherkovich	cell	spindle	
<i>Scenedesmus producto-capitatus</i> Schmula, 1909	cell	spindle	
<i>Scenedesmus raciborskii</i> Woloszynska, 1914	cell	spindle	
<i>Scherffelia</i> Pascher, 1911	cell	spheroid	
<i>Schilleriella</i> Pascher, 1932	cell	cylinder	
<i>Schizochlamys</i> Braun ex Kützing, 1849	cell	sphere	
<i>Schizomeris</i> Kützing, 1843	filament	cylinder	
<i>Schizothrix</i> Kützing ex Gomont, 1892	filament	cylinder	
<i>Schroederella</i> Pavillard, 1913	cell	cylinder	$h=0,80 \times d$
<i>Schroederia</i> Lemmermann, 1898	cell	spindle	

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<i>Schroederiella</i> Woloszynska, 1914	cell	ellipsoid	
<i>Schuettiella</i> Balech, 1988	cell	double cone	
<i>Sclerodinium</i> Dodge, 1982	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Scoliopleura</i> Grunow, 1860	cell	elliptic cylinder	
<i>Scoliotropis</i> P.T.Cleve, 1894	cell	cuboid $\times 0,85$	
<i>Scotiella</i> Fritsch, 1912	cell	spheroid	
<i>Scotiellopsis</i> Vinatzer, 1975	cell	spheroid	
<i>Scotinosphaera</i> Klebs, 1881	cell	ellipsoid	
<i>Scourfieldia</i> G.S.West, 1912	cell	ellipsoid	
<i>Scrippsiella</i> Balech ex A.R.Loeblich III, 1965	cell	spheroid	
<i>Scrippsiella mexicana</i> Indelicato & Loeblich III, 1986	cell	cone with half sphere	
<i>Scrippsiella trochoidea</i> (Stein) Balech ex Loeblich III, 1965	cell	cone with half sphere	$h=1,50 \times d$
<i>Scyphosphaera</i> Lohmann, 1902	cell	sphere	
<i>Scytonema</i> C.Agardh ex Bornet & Flahault, 1886	filament	cylinder	
<i>Scytonematopsis</i> E.I.Kiseleva, 1930	filament	cylinder	
<i>Selenastrum</i> Reinsch, 1867	cell	spindle	
<i>Selenochloris</i> Pascher, 1927	cell	spindle	
<i>Selenoderma</i> K.Bohlin, 1897	cell	spindle	
<i>Sellaphora</i> Mereschowsky, 1902	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Sellaphora bacillum</i> (Ehrenberg) D.G.Mann, 1989	cell	cuboid	
<i>Sellaphora mutata</i> (Krasske) Lange-Bertalot, 1996	cell	elliptic cylinder $\times 0,85$	$h=1,00 \times d_2$
<i>Sellaphora nana</i> (Hustedt) Lange-Bertalot, Cavacini, Tagliaventi & Alfinito, 2003	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Sellaphora pupula</i> (Kützing) Mereschkovsky, 1902	cell	cuboid $\times 0,85$	
<i>Seminavis</i> D.G.Mann, 1990	cell	elliptic cylinder $\times 0,65$	$d_2=0,50 \times h; h=2,00 \times d_2$
<i>Semiorbis</i> R.M.Patrick, 1966	cell	elliptic cylinder $\times 1,03$	$r_2=0,50 \times h; h=2,00 \times r_2$
<i>Sheshukovia</i> Z.I.Glezer [S.I.Gleser], 1975	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}$
<i>Shionodiscus</i> A.J.Alverson, S.H.Kang & E.C.Theriot, 2006	cell	cylinder	$h=0,50 \times d$
<i>Siderocelis</i> (Naumann) Fott, 1934	cell	spheroid	
<i>Siderocystopsis</i> Swale, 1964	cell	spheroid	
<i>Sieminskia</i> D.Metzeltin & Lange-Bertalot, 1998	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Simonsenia</i> Lange-Bertalot, 1979	cell	elliptic cylinder	
<i>Sinaiella</i> L.Gruia, 1965	filament	cylinder	
<i>Sinophysys</i> Nie & C.Wang, 1944	cell	ellipsoid	
<i>Siphonema</i> Geitler	filament	cylinder	
<i>Siphononema</i> Geitler, 1925	cell	spheroid	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Sirocoleum</i> Kützing ex Gomont, 1892	filament	cylinder	
<i>Skeletonema</i> Greville, 1865	cell	cylinder × 0,9	
<i>Snowella</i> Elenkin, 1938	cell	sphere	
<i>Snowella fennica</i> J.Komárek & J.Komárková-Legnerová, 1992	cell	spheroid	
<i>Snowella lacustris</i> (Chodat) Komárek & Hindák, 1988	cell	spheroid	
<i>Sokolovia</i> Elenkin, 1926	filament	cylinder	
<i>Solentia</i> Ercegovic, 1927	cell	spheroid	
<i>Sorastrum</i> Kützing, 1845	cell	tetrahedron	
<i>Spatulodinium</i> J.Cachon & M.Cachon, 1976	cell	ellipsoid	$d_2=0,67 \times d_1$
<i>Spermatozopsis</i> Korshikov, 1913	cell	spindle	
<i>Sphaeraspis</i> J.Schiller, 1954	cell	spheroid	
<i>Sphaerellocystis</i> Ettl, 1960	cell	spheroid	
<i>Sphaerellopsis fluviatilis</i> (F.Stein) Pascher, 1927	cell	spheroid × 0,88	
<i>Sphaerellopsis gloeocystiformis</i> (Dill) Gerloff, 1940	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Sphaerobotrys</i> Butcher, 1932	cell	sphere	
<i>Sphaerocalyptra</i> Deflandre, 1952	cell	sphere	
<i>Sphaerocystis</i> R.Chodat, 1897	cell	sphere	
<i>Sphaerodinium</i> Woloszynska, 1916	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Sphaeroeca</i> Lauterborn, 1894	cell	spheroid	
<i>Sphaeroneocystis</i> I.Kostikov, T.Darienko, A.Lukešová & L.Hoffmann	cell	sphere	
<i>Sphaeroplea</i> C.Agardh, 1824	filament	cylinder	
<i>Sphaerospermopsis</i> Zapomělová, Jezberová, Hrouzek, Hisem, Řeháková & Komárková, 2010	cell	sphere	
<i>Sphaerososma</i> Corda ex Ralfs, 1848	half-cell	spheroid	
<i>Sphaleromantis</i> Pascher, 1910	cell	ellipsoid	
<i>Sphenomonas</i> Stein, 1878	cell	spindle	
<i>Spiniferomonas</i> E.Takahashi, 1973	cell	sphere	
<i>Spiraulax</i> Kofoed, 1911	cell	double cone	
<i>Spirodinium</i> Schütt, 1896	cell	spindle × 0,5	
<i>Spirogyra</i> Link, 1820	cell	cylinder	
<i>Spiromonas</i> Dujardin, 1841	cell	ellipsoid	
<i>Spirotaenia</i> Brébisson, 1848	cell	spheroid	
<i>Spirulina</i> Turpin ex Gomont, 1892	filament	cylinder	
<i>Spondylomorom</i> Ehrenberg, 1848	cell	spheroid	
<i>Spondylosium</i> Brébisson ex Kützing, 1849	half-cell	spheroid	
<i>Spongiococcum</i> Deason, 1959	cell	sphere	

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<i>Spongomonas</i> Stein, 1878	cell	spheroid	
<i>Sporotetras</i> Butcher, 1932	cell	spheroid	
<i>Spumella</i> Cienkowski, 1870	cell	cone with half sphere	
<i>Stanieria</i> Komárek & Anagnostidis, 1986	cell	sphere	
<i>Starria</i> N.J.Lang, 1977	filament	elliptic cylinder	
<i>Staurastrum</i> Meyen ex Ralfs, 1848	half-cell	tetrahedron	
<i>Stauridium</i> Corda, 1839	cell	cuboid $\times 0,67$	$c=1,00 \times b$
<i>Staurodesmus</i> Teiling, 1948	half-cell	tetrahedron	
<i>Staurodesmus brevispina</i> (Brébisson) Croasdale, 1957	half-cell	spheroid	
<i>Staurodesmus convergens</i> (Ehrenberg ex Ralfs) S.Lilleroth, 1950	half-cell	spheroid	
<i>Staurodesmus crassus</i> (West & G.S.West) M.-B.Florin, 1957	half-cell	spheroid	
<i>Staurodesmus extensus</i> (Borge) Teiling, 1948	half-cell	spheroid	
<i>Staurodesmus extensus</i> var. <i>isthmus</i> (Heimerl) Coesel, 1993	half-cell	spheroid	
<i>Staurodesmus extensus</i> var. <i>joshuae</i> (Gutwinski) Teiling, 1967	half-cell	spheroid	
<i>Staurodesmus incus</i> (Hassal ex Ralfs) Teiling, 1967	half-cell	spheroid	
<i>Staurodesmus incus</i> var. <i>ralfsii</i> (West) Teiling, 1967	half-cell	spheroid	
<i>Staurodesmus indentatus</i> (West & G.S.West) Teiling, 1948	half-cell	spheroid	
<i>Staurodesmus patens</i> (Nordstedt) Croasdale, 1957	half-cell	spheroid	
<i>Staurodesmus patens</i> var. <i>maximus</i> Teiling, 1967	half-cell	spheroid	
<i>Staurodesmus subtriangularis</i> (O.Borge) Teiling, 1967	half-cell	spheroid	
<i>Staurodesmus triangularis</i> (Lagerheim) Teiling, 1948	half-cell	spheroid	
<i>Stauriforma</i> R.Flower, V.Jones & Round	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Stauroneis</i> Ehrenberg, 1843	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Stauroneis acidoclinata</i> Lange-Bertalot & Werum in Werum & Lange-Bertalot, 2004	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis acuta</i> W.Smith, 1853	cell	rhombic prism	
<i>Stauroneis agrestis</i> J.B.Petersen, 1915	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis anceps</i> Ehrenberg, 1843	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis anceps</i> f. <i>gracilis</i> Rabenhorst, 1864	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis anceps</i> var. <i>anceps</i> Ehrenberg, 1843	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis anceps</i> var. <i>hyalina</i> Peragallo & Brun	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis anceps</i> var. <i>siberica</i> Grunow, 1880	cell	elliptic cylinder $\times 0,9$	
<i>Stauroneis gracilior</i> E.Reichardt	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Stauroneis gracilis</i> Ehrenberg, 1843	cell	lanceolate cylinder	$h=0,75 \times d_2$

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Stauroneis laenburgiana</i> Hustedt, 1950	cell	elliptic cylinder × 0,9	
<i>Stauroneis neohyalina</i> LB nov.stat., 1996	cell	elliptic cylinder × 0,9	
<i>Stauroneis phoenicenteron</i> (Nitzsch) Ehrenberg, 1843	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Stauroneis pseudagrestis</i> Lange-Bertalot & Werum in Werum & Lange-Bertalot, 2004	cell	elliptic cylinder × 0,9	
<i>Stauroneis reichardtii</i> Lange-Bertalot	cell	elliptic cylinder × 0,9	
<i>Stauroneis siberica</i> (Grunow) Lange-Bertalot & Krammer	cell	elliptic cylinder × 0,9	
<i>Stauroneis smithii</i> Grunow, 1860	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Stauroneis smithii</i> var. <i>smithii</i> Grunow, 1860	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Stauronella</i> Mereschkowsky, 1901	cell	elliptic cylinder	
<i>Staurophora</i> Mereschkowsky, 1903	cell	elliptic cylinder	
<i>Staurophora salina</i> (W.Smith) Mereschkowsky, 1903	cell	lanceolate cylinder	$h=0,75 \times d_2$
<i>Stausosira</i> Ehrenberg, 1843	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Stausosira construens</i> Ehrenberg, 1843	cell	elliptic cylinder × 0,85	$h=0,50 \times d_2$
<i>Stausosira construens</i> var. <i>binodis</i> (Ehrenberg) P.B.Hamilton, 1992	cell	elliptic cylinder	$h=1,00 \times d_2$
<i>Stausosira construens</i> var. <i>exigua</i> (W.Smith) H.Kobayasi, 2002	cell	triangular prism (equilateral) × 0,65	$m=0,50 \times \sqrt{(3) \times l}$;
<i>Stausosira venter</i> (Ehrenberg) H.Kobayasi, 2002	cell	elliptic cylinder	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Stausosirella</i> D.M.Williams & Round, 1988	cell	elliptic cylinder	$d_2=1,00 \times h$; $h=1,00 \times d_2$
<i>Stausosirella leptostauron</i> (Ehrenberg) D.M.Williams & Round, 1987	cell	elliptic cylinder × 0,8	$h=0,33 \times d_2$
<i>Stalexomonas</i> Lackey, 1942	cell	spheroid	
<i>Stellarima</i> Hasle & P.A.Sims, 1986	cell	cylinder × 0,85	$h=0,65 \times d$
<i>Stellarima stellaris</i> (Roper) G.R.Hasle & P.A.Sims, 1986	cell	cylinder	$h=0,50 \times d$
<i>Stenokalyx</i> J.Schiller, 1926	cell	spheroid	
<i>Stenopterobia</i> Brébisson ex Van Heurck, 1896	cell	cuboid × 0,85	
<i>Stenopterobia delicatissima</i> (F.W.Lewis) Brébisson ex van Heurck, 1896	cell	rhombic prism × 1,15	
<i>Stephanocodon</i> Pascher, 1942	cell	spheroid	
<i>Stephanocostis</i> Genkal & Kuzmina, 1985	cell	cylinder	$h=0,60 \times d$
<i>Stephanodiscus</i> Ehrenberg, 1845	cell	cylinder	$h=0,60 \times d$
<i>Stephanodiscus astraea</i> (Ehrenberg) Grunow, 1880	cell	cylinder	$h=0,65 \times d$
<i>Stephanodiscus binderanus</i> (Kützing) Krieger, 1927	cell	cylinder	$h=1,50 \times d$
<i>Stephanodiscus hantzschii</i> Grunow, 1880	cell	cylinder	$h=0,77 \times d$
<i>Stephanodiscus hantzschii</i> f. <i>tenuis</i> (Hustedt) H.Håkansson & E.F.Stoermer, 1984	cell	cylinder	$h=0,77 \times d$
<i>Stephanodiscus minutulus</i> (Kützing) Cleve & Möller, 1882	cell	cylinder	$h=0,88 \times d$

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<i>Stephanodiscus parvus</i> Stoermer & Håkansson, 1984	cell	cylinder	$h=0,88 \times d$
<i>Stephanodiscus tenuis</i> Hustedt, 1939	cell	cylinder	$h=0,77 \times d$
<i>Stephanoeca</i> W.Ellis, 1930	cell	spheroid	
<i>Stephanoporos</i> W.Conrad & Pascher, 1940	cell	spheroid	
<i>Stephanopyxis</i> (Ehrenberg) Ehrenberg, 1845	cell	cylinder $\times 0,87$	
<i>Stichococcus</i> Nägeli, 1849	cell	cylinder	
<i>Stichogloea</i> Chodat, 1897	cell	sphere	
<i>Stichosiphon</i> Geitler, 1931	filament	cylinder	
<i>Stictocyclus</i> A.Mann, 1925	cell	cylinder	
<i>Stictodiscus</i> Greville, 1861	cell	cylinder	
<i>Stigeoclonium</i> Kützing, 1843	filament	cylinder	
<i>Stigonema</i> C.Agardh ex Bornet & Flahault, 1886	filament	cylinder	
<i>Stipitochrysis</i> Korshikov, 1941	cell	spheroid	
<i>Stipitococcus</i> West & G.S.West, 1898	cell	spheroid	
<i>Stokesiella</i> Lemmermann, 1908	cell	spheroid	
<i>Storeatula</i> Hill, 1991	cell	spheroid	
<i>Striatella</i> C.Agardh, 1832	cell	elliptic cylinder	$d_2=0,33 \times d_1$
<i>Strombomonas</i> Deflandre, 1930	cell	spheroid	
<i>Stylochrysalis</i> F.Stein, 1878	cell	spheroid	
<i>Stylodinium</i> Klebs, 1912	cell	spheroid	
<i>Stylosphaeridium</i> Geitler & Gimesi, 1925	cell	spheroid	
<i>Subsilicea</i> H.A.von Stosch & B.E.F.Reimann, 1970	cell	elliptic cylinder	$d_2=1,00 \times h$
<i>Surirella</i> Turpin, 1828	cell	elliptic cylinder	$d_2=1,12 \times h; h=0,89 \times d_2$
<i>Surirella angusta</i> Kützing, 1844	cell	cuboid $\times 0,85$	$b=1,33 \times c; c=0,75 \times b$
<i>Surirella biseriata</i> Brébisson, 1835	cell	elliptic cylinder	$d_2=1,23 \times h; h=0,81 \times d_2$
<i>Surirella capronii</i> Brébisson ex F.Kitton	cell	elliptic cylinder $\times 0,7$	$d_2=1,10 \times h; h=0,91 \times d_2$
<i>Surirella crumena</i> Brébisson ex Kützing, 1849	cell	elliptic cylinder	$d_2=1,52 \times h; h=0,66 \times d_2$
<i>Surirella linearis</i> W.Smith, 1853	cell	cuboid $\times 0,85$	$b=1,33 \times c; c=0,75 \times b$
<i>Surirella minuta</i> Brébisson, 1849	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Surirella robusta</i> Ehrenberg	cell	elliptic cylinder	$d_2=1,41 \times h; h=0,71 \times d_2$
<i>Symbiodinium</i> Freudenthal, 1962	cell	sphere	
<i>Symploca</i> Kützing ex Gomont, 1892	filament	cylinder	
<i>Symplocastrum</i> (Gomont) Kirchner, 1898	filament	cylinder	
<i>Syncrypta</i> Ehrenberg	cell	spheroid	
<i>Syndendrium</i> Ehrenberg, 1845	spore	spheroid	
<i>Synechococcus</i> Nägeli, 1849	cell	spheroid	
<i>Synechocystis</i> Sauvageau, 1892	cell	sphere	

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<i>Synedra</i> Ehrenberg, 1830	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Synedra nana</i> F.Meister, 1912	cell	elliptic cylinder	
<i>Synedrella</i> F.E.Round & N.I.Maidana, 2001	cell	elliptic cylinder	
<i>Synedropsis</i> G.R.Hasle, L.K.Medlin & E.E.Syvetsen, 1994	cell	elliptic cylinder	
<i>Synedrosphenia</i> (H.Peragallo) Azpeitia Moros, 1911	cell	pyramid × 0,9	$l_1=0,75 \times l_2; l_2=1,33 \times l_1$
<i>Synura</i> Ehrenberg, 1834	cell	cone with half sphere	
<i>Synura globosa</i> (Schiller) Starmach	cell	spheroid × 0,88	
<i>Synura splendida</i> Korshikov, 1942	cell	spheroid × 0,88	
<i>Synuopsis</i> J.Schiller, 1929	cell	spheroid × 0,88	
<i>Syracolithus</i> (Kamptner) Deflandre, 1952	cell	sphere	
<i>Syracosphaera</i> Lohmann, 1902	cell	sphere	
<i>Tabellaria</i> Ehrenberg ex Kützing, 1844	cell	cuboid	$b=0,50 \times c$
<i>Tabellaria binalis</i> var. <i>elliptica</i> R.J.Flower	cell	elliptic cylinder × 0,85	$d_2=0,33 \times h; h=3,00 \times d_2$
<i>Tabellaria fenestrata</i> (Lyngbye) Kützing, 1844	cell	cuboid	$b=0,10 \times a; c=0,33 \times a$
<i>Tabularia</i> (Kützing) D.M.Williams & Round, 1986	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Tabularia gaillonii</i> (Bory de Saint-Vincent) Bukhtiyarova, 1995	cell	cuboid × 0,8	
<i>Talaroneis</i> W.H.C.F.Kooistra & M.De Stefano, 2004	cell	elliptic cylinder	
<i>Tapinothrix</i> Sauvageau, 1892	filament	cylinder	
<i>Tasmanites</i> E.T.Newton, 1875	cell	sphere	
<i>Teilingia</i> Bourrelly, 1964	half-cell	spheroid	
<i>Teleaulax</i> Hill, 1991	cell	cone with half sphere	
<i>Telonema</i> Griessmann, 1913	cell	cone with half sphere	
<i>Terpsinoe</i> Ehrenberg, 1843	cell	elliptic cylinder	
<i>Tessellaria</i> Playfair, 1918	cell	sphere	
<i>Tetmemorus</i> Ralfs ex Ralfs, 1848	cell	ellipsoid	
<i>Tetrabaena</i> Fromentel	cell	spheroid	
<i>Tetraphlepharis</i> Senn ex Wille, 1909	cell	ellipsoid	
<i>Tetrachlorella</i> Korshikov, 1939	cell	spheroid	
<i>Tetracyclus</i> Ralfs, 1843	cell	elliptic cylinder	
<i>Tetracyclus emarginatus</i> (Ehrenberg) W.Smith, 1856	cell	elliptic cylinder × 0,75	
<i>Tetracyclus glans</i> (Ehrenberg) Mills, 1935	cell	elliptic cylinder × 0,85	$h=0,50 \times d_2$
<i>Tetracystis</i> R.M.Brown, Jr & Bold, 1964	cell	sphere	
<i>Tetradasmus</i> G.M.Smith, 1913	cell	spindle	
<i>Tetradinium</i> Klebs, 1912	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3)} \times l;$
<i>Tetraedriella</i> Pascher, 1930	cell	tetrahedron	

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<i>Tetraedron</i> Kützing, 1845	cell	tetrahedron	
<i>Tetraedron caudatum</i> (Corda) Hansgirg, 1888	cell	cuboid $\times 0,75$	$c=0,67 \times b$
<i>Tetraedron incus</i> (Teiling) G.M.Smith, 1926	cell	cuboid	$c=0,67 \times b$
<i>Tetraedron lunula</i> (Reinsch) Hansgirg, 1889	cell	spindle	
<i>Tetraedron minimum</i> (A.Braun) Hansgirg, 1888	cell	cuboid	$c=0,67 \times b$
<i>Tetraedron minimum</i> var. <i>scrobiculatum</i> Lagerheim, 1888	cell	elliptic cylinder	
<i>Tetraedron minimum</i> var. <i>tetralobulatum</i> Reinsch	cell	elliptic cylinder	
<i>Tetraedron triangulare</i> Korshikov	cell	triangular prism (equilateral) $\times 0,52$	$m=0,50 \times \sqrt{(3) \times l}$; $h=0,50 \times l$
<i>Tetrallantos</i> Teiling, 1916	cell	spindle	
<i>Tetramitus</i> Perty, 1852	cell	miscellaneous	
<i>Tetranephris</i> C.R.Leite & C.E.M.Bicudo, 1977	cell	spheroid	
<i>Tetraplektron</i> Fott, 1957	cell	tetrahedron	
<i>Tetrarcus</i> H.Skuja, 1934	cell	spheroid	
<i>Tetraselmis</i> F.Stein, 1878	cell	spheroid	
<i>Tetraspora</i> Link ex Desvaux, 1818	cell	sphere	
<i>Tetrasporopsis</i> Lemmermann ex Schmidle, 1902	cell	sphere	
<i>Tetrastrum</i> Chodat, 1895	cell	cone with half sphere	
<i>Tetrastrum elegans</i> Playfair, 1917	cell	sphere	
<i>Tetrastrum glabrum</i> (Y.V.Roll) Ahlstrom & Tiffany	cell	sphere	
<i>Tetrastrum staurogeniiforme</i> (Schröder) Lemmermann, 1900	cell	sphere	
<i>Tetrastrum triangulare</i> (Chodat) Komárek, 1974	cell	sphere	
<i>Tetratoma</i> Buetschli, 1884	cell	spheroid	
<i>Thalassiocylus</i> Håkansson & Mahood	cell	cylinder	
<i>Thalassioneis</i> Round, 1990	cell	elliptic cylinder	
<i>Thalassionema</i> Grunow ex Mereschkowsky, 1902	cell	cuboid	$b=1,00 \times c$; $c=1,00 \times b$
<i>Thalassionema bacillare</i> (Heiden) Kolbe, 1955	cell	elliptic cylinder	
<i>Thalassionema frauenfeldii</i> (Grunow) Hallegraef, 1986	cell	cuboid $\times 0,89$	$b=1,00 \times c$; $c=1,00 \times b$
<i>Thalassionema nitzschioides</i> (Grunow) Mereschkowsky, 1902	cell	cuboid $\times 0,89$	$b=1,00 \times c$; $c=1,00 \times b$
<i>Thalassionema pseudonitzschioides</i> (G.Schuette & H.Schrader) G.R.Hasle	cell	elliptic cylinder	
<i>Thalassiophysa</i> P.S.Conger, 1954	cell	elliptic cylinder	
<i>Thalassiosira</i> Cleve, 1873	cell	cylinder	$h=0,49 \times d$
<i>Thalassiosira angulata</i> (W.Gregory) Hasle, 1978	cell	cylinder	$h=0,48 \times d$
<i>Thalassiosira anguste-lineata</i> (A.Schmidt) G.Fryxell & Hasle, 1977	cell	cylinder	$h=0,38 \times d$

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<i>Thalassiosira baltica</i> (Grunow) Ostenfeld, 1901	cell	cylinder	$h=0,42 \times d$
<i>Thalassiosira constricta</i> Gaardner, 1938	cell	cylinder	$h=1,00 \times d$
<i>Thalassiosira delicatula</i> Ostenfeld, 1908	cell	cylinder	$h=1,00 \times d$
<i>Thalassiosira eccentrica</i> (Ehrenberg) Cleve, 1903	cell	cylinder	$h=0,38 \times d$
<i>Thalassiosira gravida</i> Cleve, 1896	cell	cylinder	$h=0,40 \times d$
<i>Thalassiosira hyalina</i> (Grunow) Gran, 1897	cell	cylinder	$h=0,60 \times d$
<i>Thalassiosira hyperborea</i> (Grunow) Hasle	cell	cylinder	$h=0,40 \times d$
<i>Thalassiosira hyperborea</i> var. <i>pelagica</i> (Cleve-Euler) G.R.Hasle	cell	cylinder	$h=0,40 \times d$
<i>Thalassiosira lacustris</i> (Grunow) Hasle, 1977	cell	cylinder	$h=0,54 \times d$
<i>Thalassiosira levanderi</i> van Goor, 1924	cell	cylinder	$h=0,60 \times d$
<i>Thalassiosira minima</i> Gaarder, 1951	cell	cylinder	$h=0,50 \times d$
<i>Thalassiosira nordenskiöldii</i> Cleve, 1873	cell	cylinder	$h=0,61 \times d$
<i>Thalassiosira proschkinae</i> Makarova, 1979	cell	cylinder	$h=0,50 \times d$
<i>Thalassiosira pseudonana</i> Hasle & Heimdal, 1970	cell	cylinder	$h=0,65 \times d$
<i>Thalassiosira punctigera</i> (Castracane) Hasle, 1983	cell	cylinder	$h=0,50 \times d$
<i>Thalassiosira rotula</i> Meunier, 1910	cell	cylinder	$h=0,44 \times d$
<i>Thalassiothrix</i> Cleve & Grunow, 1880	cell	cuboid	$b=1,00 \times c; c=1,00 \times b$
<i>Thaumatomastix</i> Lauterborn, 1899	cell	spheroid	
<i>Thecadinium</i> Kofoid & Skogsberg, 1928	cell	ellipsoid	
<i>Thelesphaera</i> Pascher, 1943	cell	sphere	
<i>Thompsodinium</i> Bourrelly, 1970	cell	spheroid	
<i>Thoracomonas</i> Korshikov, 1925	cell	ellipsoid	
<i>Thorakochloris</i> Pascher, 1932	cell	sphere	
<i>Thorosphaera</i> Ostenfeld, 1910	cell	sphere	
<i>Togula</i> Flo Jorgensen, Murray & Daugbjerg, 2004	cell	ellipsoid	
<i>Tolypothrix</i> Kützing ex Bornet & Flahault, 1886	filament	cylinder	
<i>Torodinium</i> Kofoid & Swezy, 1921	cell	ellipsoid	$d_2=0,90 \times d_1$
<i>Torodinium teredo</i> (Pouchet) Kofoid & Swezy, 1921	cell	ellipsoid	$d_2=0,76 \times d_1$
<i>Tovellia</i> Moestrup, K.Lindberg & N.Daugberg, 2005	cell	spheroid	
<i>Tovellia leopoliensis</i> (Wolosz.) Moestrup, Lindberg & Daugbjerg, 2005	cell	ellipsoid	
<i>Toxonidea</i> Donkin, 1858	cell	elliptic cylinder	
<i>Trachelomonas</i> Ehrenberg, 1835	cell	spheroid	
<i>Trachelomonas rugulosa</i> F.Stein ex Deflandre, 1926	cell	sphere	
<i>Trachelomonas volvocina</i> (Ehrenberg) Ehrenberg, 1834	cell	sphere	
<i>Trachelomonas volvocinopsis</i> Svirenko, 1914	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Trachelophyllum</i> Claparède & Lachmann, 1859	cell	spindle	
<i>Trachychloron</i> Pascher, 1939	cell	spheroid	
<i>Trachydiscus</i> Ettl, 1964	cell	spheroid	
<i>Trachyneis</i> P.T.Cleve, 1894	cell	elliptic cylinder	$d_2=0,80 \times h; h=1,25 \times d_2$
<i>Trachysphenia</i> Petit, 1877	cell	elliptic cylinder	
<i>Trebouxia</i> Puymaly, 1924	cell	sphere	
<i>Trepomonas</i> Dujardin, 1841	cell	spheroid	
<i>Treubaria</i> C.Bernard, 1908	cell	sphere	
<i>Treubaria crassispina</i> G.M.Smith, 1926	cell	tetrahedron	
<i>Treubaria quadrispina</i> (G.M.Smith) Fott & Kovácik, 1975	cell	cuboid	
<i>Treubaria setigera</i> (Archer) G.M.Smith, 1933	cell	triangular prism	
<i>Tribonema</i> Derbès & Solier, 1851	cell	cylinder	
<i>Triceratium</i> Ehrenberg, 1839	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}; h=0,50 \times l$
<i>Trichocoleus</i> Anagnostidis, 2001	filament	cylinder	
<i>Trichodesmium</i> Ehrenberg ex Gomont, 1892	filament	cylinder	
<i>Trichormus</i> (Ralfs ex Bornet & Flahault) Komárek & Anagnostidis, 1989	cell	cylinder	
<i>Trichotoxon</i> F.M.H.Reid & Round, 1988	cell	cuboid	$b=1,00 \times c; c=1,00 \times b$
<i>Trigonaspis</i> H.A.Thomsen, 1980	cell	spheroid	
<i>Trigonium</i> Cleve, 1867	cell	triangular prism (equilateral)	$m=0,50 \times \sqrt{(3) \times l}; h=1,00 \times l$
<i>Trigonomonas</i> Klebs, 1892	cell	spheroid	
<i>Triparma</i> B.C.Booth & H.J.Marchant, 1987	cell	sphere	
<i>Triploceras</i> J.W.Bailey, 1851	cell	cylinder	
<i>Trochiscia</i> Kützing, 1834	cell	sphere	
<i>Tropidoneis</i> Cleve, 1891	cell	rhombic prism	
<i>Tryblionella</i> W.Smith, 1853	cell	elliptic cylinder $\times 1,15$	$h=0,75 \times d_2$
<i>Tryblionella aerophila</i> (Hustedt) D.G.Mann, 1990	cell	cuboid $\times 0,8$	$c=0,75 \times b$
<i>Tryblionella angustata</i> W.Smith, 1853	cell	elliptic cylinder $\times 0,8$	$h=0,75 \times d_2$
<i>Tryblionella apiculata</i> Gregory, 1857	cell	cuboid $\times 0,8$	$c=0,75 \times b$
<i>Tryblionella calida</i> (Grunow) D.G.Mann, 1990	cell	cuboid $\times 0,8$	$c=0,75 \times b$
<i>Tryblionella compressa</i> (J.W.Bailey) M.Poulin, 1990	cell	elliptic cylinder	$h=0,75 \times d_2$
<i>Tryblionella debilis</i> Arnott ex O'Meara, 1873	cell	elliptic cylinder $\times 0,8$	$h=0,75 \times d_2$
<i>Tryblionella divergens</i> (Hustedt) D.G.Mann, 1990	cell	rhombic prism $\times 1,15$	
<i>Tryblionella hungarica</i> (Grunow) Frenguelli, 1942	cell	cuboid $\times 0,8$	$c=0,75 \times b$
<i>Tryblionella marginulata</i> (Grunow) D.G.Mann, 1990	cell	elliptic cylinder $\times 1,15$	$h=0,50 \times d_2$

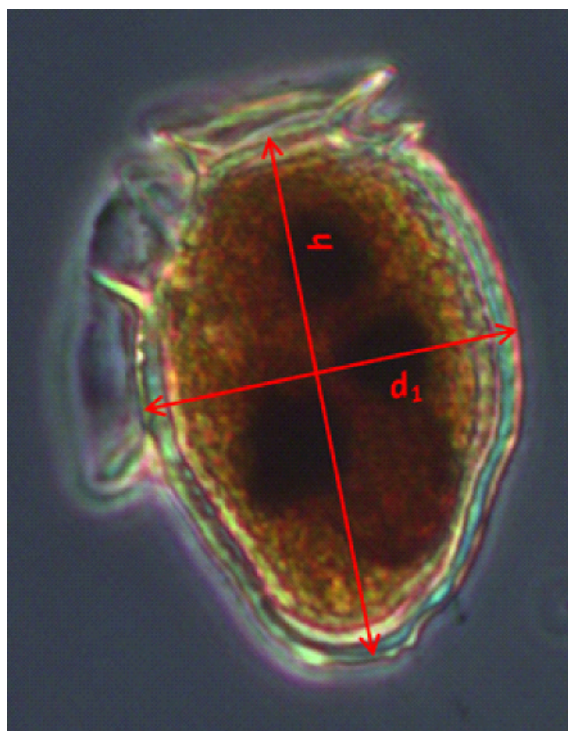
Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (× correction factor)	Hidden dimension estimation*
<i>Tryblionella navicularis</i> (Brébisson) Ralfs, 1861	cell	elliptic cylinder × 0,8	$h=0,75 \times d_2$
<i>Tryblionella plana</i> (W.Smith) Pelletan, 1889	cell	cuboid	$c=0,75 \times b$
<i>Tryblionella salinarum</i> (Grunow) Pantocsek, 1901	cell	cuboid	$c=0,75 \times b$
<i>Tryblionella scalaris</i> (Ehrenberg) P.Siver & P.B.Hamilton, 2005	cell	cuboid × 0,9	$c=0,75 \times b$
<i>Tubiella</i> Hollerbach, 1935	cell	spheroid	
<i>Turrisphaera</i> I.Manton, J.Sutherland & K.Oates, 1976	cell	spheroid	
<i>Tychonema</i> Anagnostidis & Komárek, 1988	filament	cylinder	
<i>Tyrannodinium</i> A.J.Calado, S.C.Craveiro, N.Daugbjerg & Ø.Moestrup, 2009	cell	ellipsoid	
<i>Ulnaria</i> (Kützing) P.Compère, 2001	cell	elliptic cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Ulnaria acus</i> (Kützing) M.Aboal, 2003	cell	lanceolate cylinder	$d_2=1,00 \times h; h=1,00 \times d_2$
<i>Ulnaria capitata</i> (Ehrenberg) P.Compère, 2001	cell	cuboid	
<i>Ulnaria danica</i> (Kützing) Compère & Bukhtiyarova, 2006	cell	cuboid	
<i>Ulnaria delicatissima</i> (W.Smith) M.Aboal & P.C.Silva, 2004	cell	rhombic prism	
<i>Ulnaria delicatissima</i> var. <i>angustissima</i> (Grunow) M.Aboal & P.C.Silva, 2004	cell	rhombic prism	
<i>Ulothrix</i> Kützing, 1833	cell	cylinder	
<i>Umbellosphaera</i> Paasche, 1955	cell	sphere	
<i>Umbilicosphaera</i> Lohmann, 1902	cell	sphere	
<i>Undatella</i> Paddock & P.A.Sims, 1980	cell	elliptic cylinder	
<i>Urceolus</i> C.von Mereschkowsky, 1879	cell	spheroid	
<i>Uroglena</i> Ehrenberg, 1834	cell	spheroid	
<i>Uroglenopsis</i> Lemmermann, 1899	cell	spheroid	
<i>Uronema</i> Lagerheim, 1887	filament	cylinder	
<i>Urosolenia</i> Round & R.M.Crawford, 1990	cell	cylinder	
<i>Vacuolaria</i> Cienkowski, 1870	cell	ellipsoid	
<i>Vaucheria</i> A.P.de Candolle, 1801	filament	cylinder	
<i>Vischeria</i> Pascher, 1938	cell	sphere	
<i>Vitreochlamys</i> Batko, 1970	cell	spheroid	
<i>Vitreochlamys fluviatilis</i> (Stein) Batko, 1970	cell	spheroid × 0,88	
<i>Vitreochlamys gloeocystiformis</i> (O.Dill) A.Nakazawa in Nakazawa Krienitz & Nozaki, 2001	cell	sphere	
<i>Volvox</i> Linnaeus, 1758	cell	sphere	
<i>Volvolina</i> Playfair, 1915	cell	spheroid	
<i>Warnowia</i> Lindemann, 1928	cell	spheroid	
<i>Westella</i> De Wildermann, 1897	cell	sphere	

Taxon (genus, species, subspecies, variety, forma)	Geometrical shape unit	Geometrical shape (\times correction factor)	Hidden dimension estimation*
<i>Westellopsis</i> C.-C.Jao, 1959	cell	sphere	
<i>Wigwamma</i> I.Manton, J.Sutherland & K.Oates, 1977	cell	spheroid	
<i>Willea</i> Schmidle, 1900	cell	spheroid	
<i>Wollea</i> Bornet & Flahault, 1886	filament	cylinder	
<i>Woloszynskia</i> R.H.Thompson, 1951	cell	ellipsoid	$d_2=0,75 \times d_1$
<i>Woloszynskia coronata</i> (Woloszynska) Moestrup, Lindberg & Daugbjerg, 2005	cell	spheroid	
<i>Woloszynskia halophila</i> (Biecheler) M.Elbrächter & A.Kremp, 2005	cell	spheroid	
<i>Wolskyella</i> Claus, 1963	cell	cylinder	
<i>Woronichinia</i> Elenkin, 1933	cell	spheroid	
<i>Xanthidium</i> Ehrenberg ex Ralfs, 1848	half-cell	spheroid	
<i>Xanthonema</i> P.C.Silva, 1979	cell	cylinder	
<i>Xenococcus</i> Thuret, 1880	cell	spheroid	
<i>Xenotholos</i> M.Gold-Morgan, G.Montejano & J.Komárek, 1994	cell	spheroid	
<i>Zygnema</i> C.Agardh, 1817	cell	cylinder	
<i>Zygnemopsis</i> (Skuja) Transeau, 1934	cell	cylinder	
<i>Zygoceros</i> C.G.Ehrenberg, 1839	cell	miscellaneous	
<i>Zygogonium</i> Kützing, 1843	cell	cylinder	
<i>Zygosphaera</i> Kamptner, 1937	cell	sphere	

Annex E (informative)

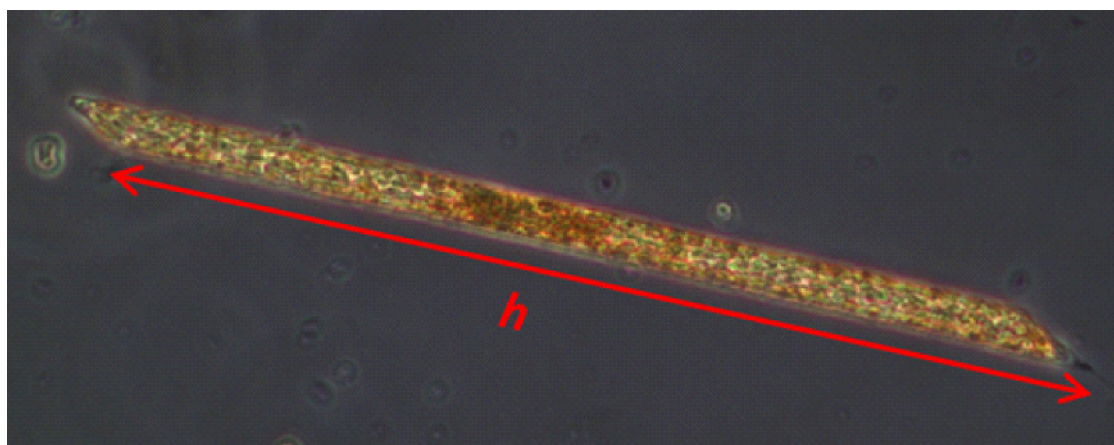
Example: Genus-specific instruction for measurement of dimensions

Figures E.1 to E.6 show examples for some well-chosen taxa to measure the necessary dimensions.



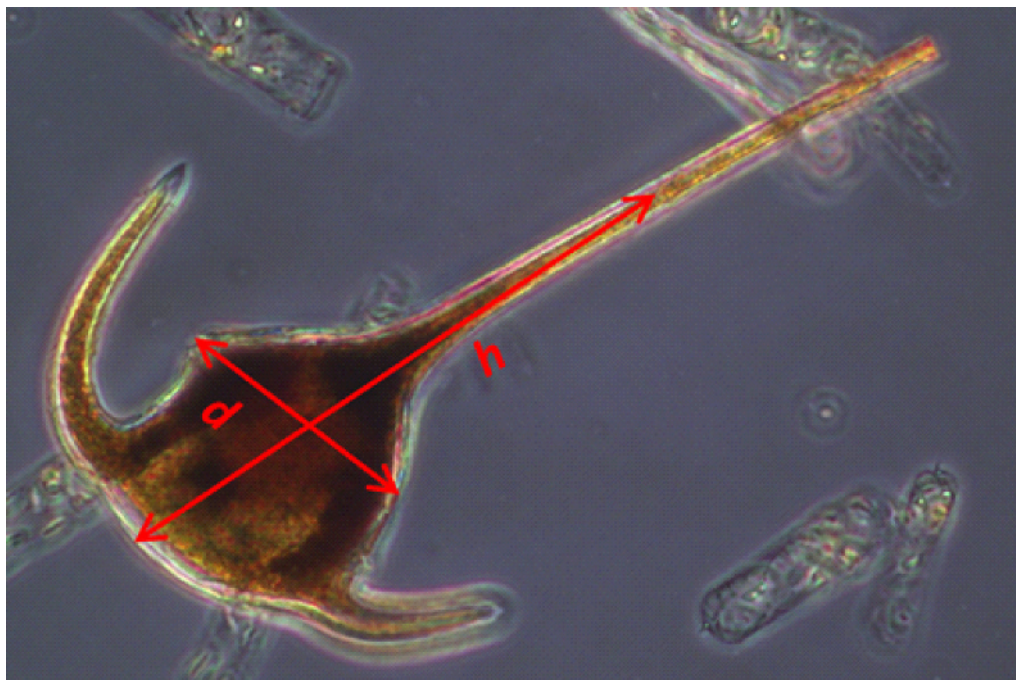
NOTE The necessary dimensions large diameter (d_1) and height (h) of the “ellipsoid” are both measured without the cell “wings”.

Figure E.1 — *Dinophysis*



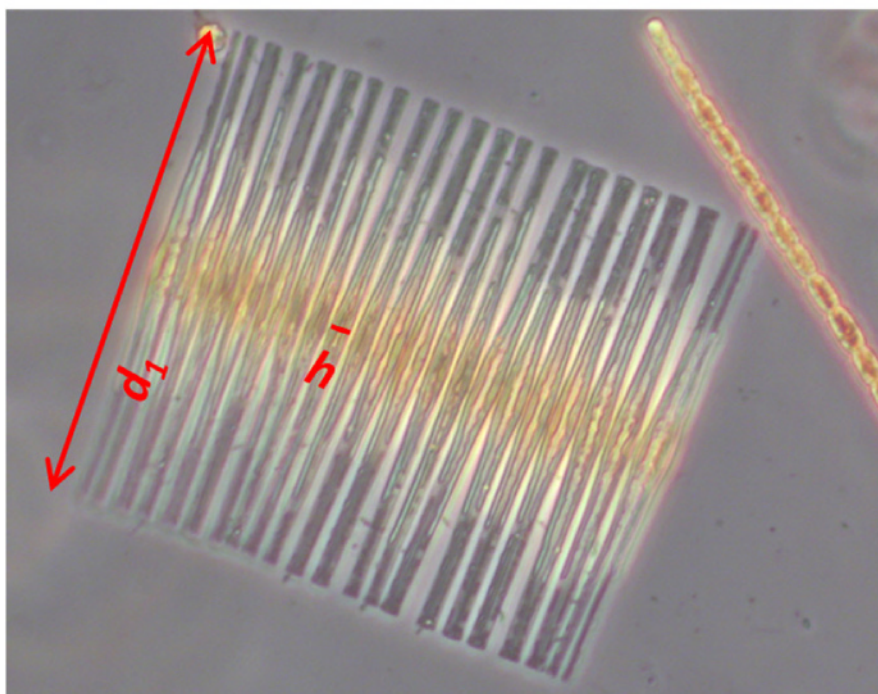
NOTE The height (h) of the “cylinder” is measured on one side from the tip to the beginning of the inclination at the other end. Thus, the unmeasured tip on one side replaces the missing piece on the other side.

Figure E.2 — *Rhizosolenia*



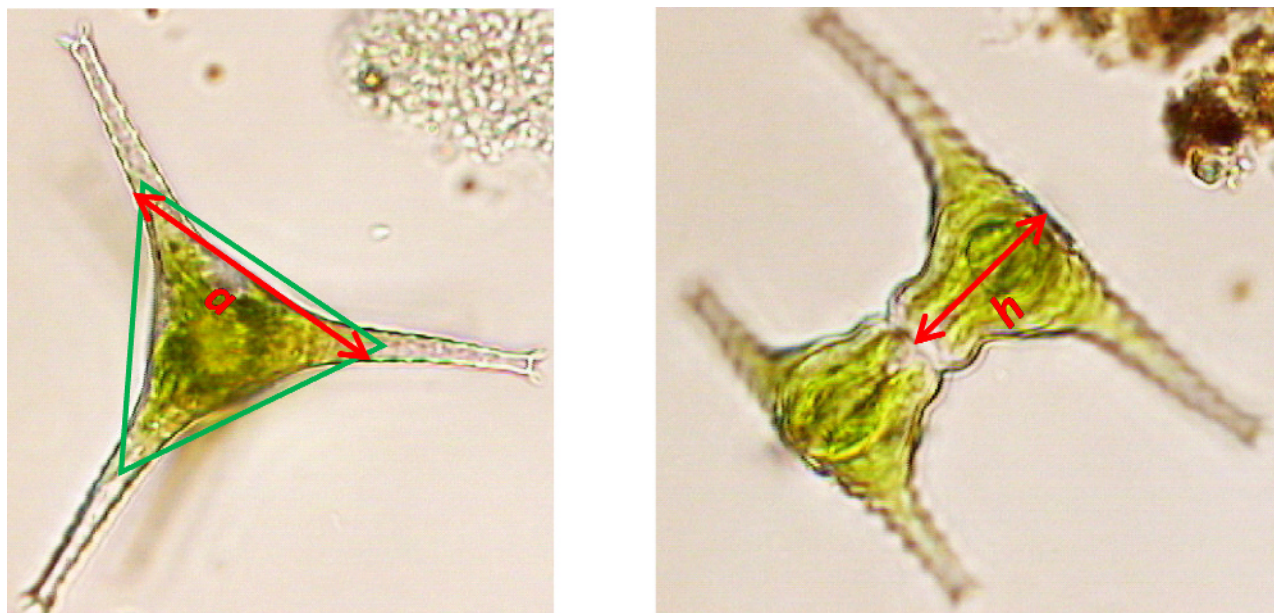
NOTE For those species, for which the volume can be expressed as the special Ceratium-shape (see Annex A) the girdle-diameter (d) is sufficient for calculation. For other species that are assigned as “cone with half sphere” additionally the height (h) is necessary, which can be measured approximately up to half of the horn length. Thus, the unrecognized horns and parts of the “half sphere” compensate the resulting overestimation of the cone. The flattening of the whole cell is addressed by the additional use of a correction factor.

Figure E.3 — *Ceratium*



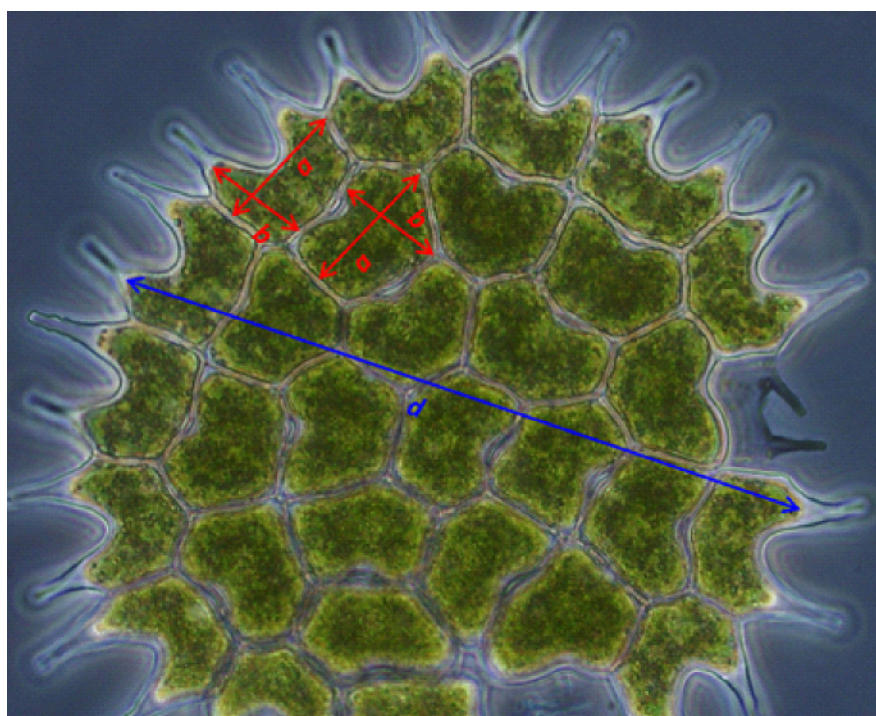
NOTE For that genus, which species can be a “cuboid”, an “elliptic cylinder”, or a “rhombic prism” the height (h) is measured in the middle of the cell. In cases where cells are narrowed towards the apical poles, this is taken into account by using a correction factor.

Figure E.4 — *Fragilaria*



NOTE The shape for this genus is a “tetrahedron” on half-cell basis. The edge length (a) of the basic triangle (left figure) should be measured between the opposite edges of the “arms” after the eyepiece scale has been applied to one cell edge (a). The resulting slight overestimation of this basis is compensated by the “arms” and the underestimation in the area of the upper part of the “tetrahedron”. In the appropriate position, the height (h) can be measured between the basis and the isthmus axis.

Figure E.5 — Staurastrum



NOTE If applying a “cylinder” on coenobium basis it is easy to measure the diameter (d). For the geometrical shape correction factor for the “cuboid” and the hidden dimension factors see Annex D.

Figure E.6 — Pediastrum

If in Figure E.6, the “cuboid” is used on cell basis the length (a) and width (b) should be measured at the widest positions.

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