



Standard Guide for Measurement of Particle Size Distribution of Nanomaterials in Suspension by Nanoparticle Tracking Analysis (NTA)¹

This standard is issued under the fixed designation E2834; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This guide deals with the measurement of particle size distribution of suspended particles, from ~10 nm to the onset of sedimentation, sample dependent, using the nanoparticle tracking analysis (NTA) technique. It does not provide a complete measurement methodology for any specific nanomaterial, but provides a general overview and guide as to the methodology that should be followed for good practice, along with potential pitfalls.

1.2 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.3 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 ASTM Standards:²

[C322 Practice for Sampling Ceramic Whiteware Clays](#)

[E456 Terminology Relating to Quality and Statistics](#)

[E1617 Practice for Reporting Particle Size Characterization Data](#)

[E2490 Guide for Measurement of Particle Size Distribution of Nanomaterials in Suspension by Photon Correlation Spectroscopy \(PCS\)](#)

2.2 ISO Standards:³

[ISO 13320 Particle Size Analysis—Laser Diffraction Methods](#)

[ISO 13321 Particle Size Analysis—Photon Correlation Spectroscopy](#)

[ISO 14488 Particulate Materials—Sampling And Sample Splitting for the Determination of Particulate Properties](#)
[ISO 22412 Particle Size Analysis—Dynamic Light Scattering \(DLS\)](#)

3. Terminology

3.1 Definitions:

3.1.1 *diffusion coefficient, n*—a measure to characterize the rate a particular molecule or particle moves in a particular medium when driven by random thermal agitation (Brownian motion).

3.1.1.1 *Discussion*—After measurement, the value is to be inputted into the Stokes-Einstein equation (Eq 1, see 7.2.1.2(3)). Diffusion coefficient units in nanoparticle tracking analysis (NTA) measurements are typically cm^2/s , rather than the correct SI units of m^2/s .

3.1.2 *repeatability, n*—in NTA and other particle sizing techniques, this usually refers to a measure of the precision of repeated consecutive measurements on the same group of particles under identical conditions and is normally expressed as a relative standard deviation (RSD) or coefficient of variation (CV).

3.1.2.1 *Discussion*—The repeatability value reflects the stability (instrumental, but mainly the sample) of the system over time. Changes in the sample could include dispersion, aggregation and settling.

3.1.3 *reproducibility, n*—in NTA and particle sizing this usually refers to a measure of the deviation of the results obtained from the first aliquot to that obtained for the second and further aliquots of the same bulk sample (and therefore is subject to the homogeneity or heterogeneity of the starting material and the sampling method employed). Normally expressed as a relative standard deviation (RSD) or coefficient of variation (CV).

3.1.3.1 *Discussion*—In a heterogenous and polydisperse (for example, slurry) system, it is often the largest error when repeated samples are taken. Other definitions of reproducibility also address the variability among single test results gathered from different laboratories when inter-laboratory testing is undertaken, or operator-to-operator, instrument-to-instrument, location-to-location, or even day-to-day. It is to be noted that the same group of particles can never be measured in such a

¹ This guide is under the jurisdiction of ASTM Committee E56 on Nanotechnology and is the direct responsibility of Subcommittee E56.02 on Characterization: Physical, Chemical, and Toxicological Properties.

Current edition approved April 1, 2012. Published May 2012. DOI: 10.1520/E2834-12.

² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

³ Available from American National Standards Institute (ANSI), 25 W. 43rd St., 4th Floor, New York, NY 10036, <http://www.ansi.org>.

system of tests and therefore reproducibility values may typically be considerably in excess of repeatability values.

3.1.4 *robustness, n*—a measure of the change of the required parameter with deliberate and systematic variations in any or all of the key parameters that influence it.

3.1.4.1 *Discussion*—For example, dispersion energy input (that is, ultrasound power and duration) almost certainly will affect the reported results. Variation in pH is likely to affect the degree of agglomeration and so forth. A useful discussion of robustness experiment considerations is found in the ICH Validation of Analytical Procedures Q2(R1) Guideline (1).⁴

3.1.5 *rotational diffusion, n*—a process by which the equilibrium statistical distribution of the overall orientation of molecules or particles is maintained or restored.

3.1.6 *translational diffusion, n*—a process by which the equilibrium statistical distribution of molecules or particles in space is maintained or restored.

3.1.7 *visualization, n*—as it relates to the NTA technique, the particles themselves are not imaged, being below the diffraction limit. Each particle acts as a point scatterer, meaning that the imaging system only sees the scattered light from the particle. This allows the position of each particle to be identified and followed with respect to time. See 7.2.

3.1.7.1 *Discussion*—The intensity and shape of the scattered light pattern for each particle may vary, and some additional information may be obtained from these differences, at least qualitatively, but is outside the scope of this guide.

3.1.8 *percentile, n*—a statistical measure of the distribution of sizes. The size below which a certain percent of the distribution falls. For example, the 10th percentile is the size below which 10 percent of the particles may be found. Expressed in ISO form as x_{10} , x_{50} , x_{90} , and also commonly expressed as D10, D50, D90. The 50th percentile is the median.

3.1.9 *coefficient of variation, n*—in statistics, a normalized measure of dispersion of a distribution. Defined as the standard deviation divided by the mean value. (Note: $CV = SD/Mean$)

3.1.10 *relative standard deviation, n*—in statistics, the absolute value of the coefficient of variation, expressed as a percentage. (Note: $RSD = 100 \cdot SD/Mean$)

NOTE 1—Other common statistical measures are defined in Terminology E456.

3.2 Acronyms:

3.2.1 *CV*—coefficient of variation

3.2.2 *CCD*—charge-coupled device

3.2.3 *CMOS*—complementary metal–oxide–semiconductor

3.2.4 *DLS*—dynamic light scattering

3.2.5 *EMCCD*—electron-multiplying charge-coupled device

3.2.6 *NTA*—nanoparticle tracking analysis

3.2.7 *PCS*—photon correlation spectroscopy

3.2.8 *RSD*—relative standard deviation

4. Summary of Guide

4.1 Nanoparticle tracking analysis (NTA) is a method for the direct and real-time visualization and analysis of nanoparticles in liquids. Particles in suspension are illuminated with a focused laser beam. Light scattered from each particle is visible through magnifying optics fitted to a digital camera such as a CCD. The software analyzes the video stream from the camera, identifying and tracking the motion of each particle with time. Because each particle in the field of view is being simultaneously but separately tracked and analyzed, the particle size distribution profile obtained by NTA is a direct number-based distribution.

4.2 The laser beam is focused such that only particles in the focal plane of the magnifying optics are illuminated. Particles out of the focal plane are not illuminated and at the size range under discussion are not visible to the camera. This yields a high signal to noise image, allowing particles as small as 10 nm to be visualized, depending on sample material. While outside the scope of this document, the technique is generally able to measure particles as large as approximately 1 μm .

4.3 The average distance each particle moves in the image is automatically calculated by the software. From this value, the particle diffusion coefficient can be obtained and through the use of the Stokes-Einstein equation, particle size can be determined.

4.4 This Guide discusses the scientific basis for the technique, size limits, concentration ranges, sampling and sample preparation considerations, condition and analysis selection, data interpretation and comparison to other complementary techniques.

5. Significance and Use

5.1 NTA is one of the very few techniques that are able to deal with the measurement of particle size distribution in the nano-size region. This Guide describes the NTA technique for direct visualization and measurement of Brownian motion, generally applicable in the particle size range from several nanometers until the onset of sedimentation in the sample. The NTA technique is usually applied to dilute suspensions of solid material in a liquid carrier. It is a first principles method (that is, calibration in the standard understanding of this word, is not involved). The measurement is hydrodynamically based and therefore provides size information in the suspending medium (typically water). Thus the hydrodynamic diameter will almost certainly differ from size diameters determined by other techniques and users of the NTA technique need to be aware of the distinction of the various descriptors of particle diameter before making comparisons between techniques (see 8.7). Notwithstanding the preceding sentence, the technique is routinely applied in industry and academia as both a research and development tool and as a QC method for the characterization of submicron systems.

6. Reagents

6.1 In general, no reagents specific to the technique are necessary. However, dispersing and stabilizing agents often are

⁴ The boldface numbers in parentheses refer to the list of references at the end of this standard.

required for a specific test sample in order to preserve colloidal stability during the measurement. A suitable diluent is used to achieve a particle concentration appropriate for the measurement. The apparent hydrodynamic size or diffusion coefficient may undergo change on dilution, as the ionic environment, within which the particles are dispersed, changes in nature or concentration. This is particularly noticeable when diluting a monodisperse latex. A latex that is measured as 60 nm in 1×10^{-3} M NaCl can have a hydrodynamic diameter of over 70 nm in 1×10^{-6} M NaCl (close to deionized water).

6.2 In order to minimize any changes in the system on dilution, it is common to use the “mother liquor”. This is the liquid in which the particles exist in stable form and is usually obtained by centrifuging of the suspension or making up the same ionic composition of the dispersant liquid if knowledge of these components is available. Many biological materials are measured in a buffer (often phosphate buffered saline), which confers the correct (range of) conditions of pH and ionic strength to assure stability of the system. Instability (usually through inadequate zeta potential—see (2)) can promote agglomeration leading to settling or sedimentation in a solid-liquid system or creaming in a liquid-liquid system (emulsion). Such fundamental changes interfere with the stability of the suspension and need to be minimized as they affect the quality (accuracy and repeatability) of the reported measurements. These should be investigated in a robustness experiment.

7. Procedure

7.1 Verification:

7.1.1 The instrument to be used in the measurement should be verified for correct performance, within pre-defined quality control limits, by following protocols issued by the instrument manufacturer. These confirmation tests normally involve the use of one or more NIST-traceable spherical particle size standards. In the sub-micron ($< 1 \times 10^{-6}$ m) region, these standards (for example, NIST, Duke Scientific - now part of ThermoFisher Scientific) tend to be nearly monodisperse (that is, narrow, single mode distribution, $CV < 17\%$) and, while confirming the x (size) axis, do not verify the y (or quantity) axis of the size distribution. Note that NTA is a first principles measurement and thus calibration in the formal sense (adjustment of the instrument to read a true and known value) need not be undertaken. In the event of a “failure” at the verification stage, then the issues to check involve quality of the dilution water, state of dispersion and stability of the standard under dilution plus instrumental issues such as thermal stability, cleanliness and alignment of optical components. The raw video data can be examined during and after acquisition. Such examination can provide useful qualitative information about the sample condition, particle concentration, and system operation. During data acquisition one looks for a consistent number of particles in the field of view, well-separated particles, a low level of random or background noise, and particle tracking lengths sufficient for accurate measurements of each particle. Manufacturers also provide other means of assuring the reliability of the data and it is recommended that these protocols are consulted, as appropriate.

7.1.2 Given the likelihood that the size standard has been certified by electron microscopy, care needs to be exercised in direct comparison of the results. While electron microscopy is carried out under high-vacuum conditions, NTA performs the analysis with particles in suspension. NTA measures the diffusion coefficient of the particle and the diameter given by the Stokes-Einstein relation (Eq 1) is that of the sphere-equivalent, hydrodynamic diameter (the particle itself plus the molecular-scale layer of solvent associated with the particle surface). This solvent layer may be significant relative to the size of the standard particles being analyzed, increasing the apparent size of the particles. For larger particles, however, the effect of this hydrodynamic layer is minimal.

7.1.3 Note that verification of a system only demonstrates that the instrument is performing adequately with the prescribed standard materials. Practical considerations for real-world materials (especially “dispersion” if utilized in sample preparation or if the distribution is relatively polydisperse) mean that the method used to measure that real-world material needs to be carefully evaluated for precision (repeatability).

7.2 Measurement:

7.2.1 Introduction:

7.2.1.1 The measurement of particle size distribution in the nano- (sub 100 nm) region by nanoparticle tracking analysis depends on the interaction of light with matter and the random or Brownian motion that a particle exhibits in liquid medium in free suspension (3). There must be an inhomogeneity in the refractive indices of a particle and the medium within which it exists in order for light scattering to occur. Without such an inhomogeneity (for example, in so-called index-matched systems) there is no scattering and the particle is invisible to light and no measurements can be made by the NTA or any other technique making use of light scattering. A coating or functionalization of the primary particle may affect this refractive index sufficiently to impact the light scattering properties. While some physical phenomena used by the NTA measurement technique are in common with the dynamic light scattering technique (photon correlation spectroscopy), as defined in Guide E2490 and ISO 22412, NTA is a distinctly different technique.

7.2.1.2 For particles < 100 nm, as considered in this guide, several facts hold true:

(1) The amount of scattering is weak in relative terms and depends highly on the size of the particle as well as particle composition. In the Rayleigh approximation region (typically $d < \lambda/10$ in which d is the diameter of particle and λ is the wavelength of light employed), then this intensity of scattering is proportional to d^6 —or (volume)² or (relative molecular mass)². With a commonly utilized diode laser (638 nm), then the upper size limit of this Rayleigh scattering behavior is approximately 64 nm. This means, in practice, that a 60 nm particle scatters 1 million times as much light as a 6 nm particle of the same composition. Thus, for a sample that contains a wide range of particle sizes or includes contaminating particles (for example dust) that are often present in the local environment and are usually considerably larger than the material that requires measurement, caution must be exercised in selecting the appropriate measurement conditions to properly analyze all

particles. NTA is able to measure over a wide range of sizes, but it may be difficult to find a set of instrument settings to measure all particles in a single analysis. This would mean filtering liquids used to contain or dilute the particles to at least the same level as the size of the particles that require characterizing unless the user is conscious of the inclusion of these particles in the final result. Alternatively, two separate analyses may be conducted, with the conditions optimized for either the smaller or larger particles, then the results from each added together. Unless the two populations are totally distinct, caution must be exercised in interpreting this combined result particularly in the region where the two analyses overlap.

(2) The intensity of scattering in the Rayleigh region is inversely proportional to the fourth power of the wavelength of light employed. Thus, if the wavelength of incident light could be halved then the intensity of scattering that would be observed is increased by a factor of 16. It is possible to use lasers of a lower wavelength than 638 nm to increase the amount of scattering and, hence, signal. This is usually preferable to increasing the power of the laser with possible undesired effects (for example, heating, convection currents). However, note that lower wavelengths sometimes overlap an absorption edge for some molecular species leading to a loss of signal intensity. The detector is a digital video camera (such as CCD, higher-sensitivity EMCCD, or CMOS) of an appropriate frame rate and spectral response to the laser wavelength being used. Sensitivity of the camera varies with manufacturer and together with the other variables discussed such as laser wavelength and power, particle and liquid refractive indices, and optical configuration will determine the lower detection limit of the system.

(3) The measurement of size in the sub-100 nm region relies on the measurement of the amount of Brownian motion (in particular the diffusion coefficient) of the particle as formulated in the Stokes-Einstein equation:

$$d_H = \frac{K_B T}{3\pi\eta D_t} \quad (1)$$

Where d_H is the particle hydrodynamic diameter, K_B is the Boltzmann constant, T is absolute temperature in Kelvin, η is viscosity in centipoise and D_t the (measured) translational diffusion coefficient in cm^2/s .

(4) Note that, in Eq 1, the density of the particle plays no role in Brownian motion (although, of course, it does in settling; see Point 9 below), even though this appears to be counterintuitive. Note also that a hydrodynamic diameter is derived. This refers to an equivalent size in spherical terms to that of a particle moving with the same diffusion coefficient as the observed particle. Thus, for an irregularly shaped particle or one with significant external morphology or surface coatings (or both), then the derived diameter is not likely to correspond to any measured axis of the image of the particle (4). The viscosity refers to the medium in which the particle is dispersed. In a dilute system it is assumed that the particles do not interact, so the viscosity can be assumed to be that of the medium or diluent.

(5) Note the term diffusion coefficient. There are two types of diffusion to be considered for particles in free suspension:

(a) Translational, where the so-called Stokes-Einstein relationship given in Eq 1 applies. Rewriting with the diffusion coefficient on the left:

$$D_t = \frac{K_B T}{3\pi\eta d_H} \quad (2)$$

(b) Rotational, where the Stokes-Einstein-Debye relation applies:

$$D_r = \frac{K_B T}{\pi\eta d_H^3} \quad (3)$$

(6) Association of particles (or molecules) leads to changes in the rotational diffusion coefficient, and also affects the translational diffusion coefficient. Hence, interactions between particles can complicate the interpretation of the observed diffusion coefficient, which for nonspherical particles, is a combination of the translational and rotational diffusion coefficients. These particle-particle interactions tend to be concentration rather than size dependent, and both translational and rotational diffusion coefficients are dependent on the viscosity of the surrounding fluid. For the concentrations appropriate to NTA measurements, the particle concentration is generally too dilute for these effects to be significant.

(7) Brownian translational motion occurs in three dimensions but NTA observes motion only in two dimensions. It is possible to determine a diffusion coefficient based on the measurement of one, two or three dimensions (and theoretically up to six if rotation could be measured). By measuring a higher order of $D_{t,i}$ a more accurate approximation can be made of the particle size for a given number of steps that contribute towards a track. D_t is derived from measuring the mean squared displacement of a particle in one, two or three dimensions (Eq 4-6 respectively) (5, 6).

$$\overline{(x^2)} = \frac{2TK_B t}{3\pi\eta d} \quad (4)$$

$$\overline{(x,y)^2} = \frac{4TK_B t}{3\pi\eta d} \quad (5)$$

$$\overline{(x,y,z)^2} = \frac{2TK_B t}{\pi\eta d} \quad (6)$$

Where t is the time between sequential displacement measurements, in this case the frame to frame period.

(a) In the case where measurement of movement in two dimensions is made:

$$\frac{\overline{(x,y)^2}}{4t} = D_t = \frac{TK_B}{3\pi\eta d} \quad (7)$$

and results thus obtained by NTA are shown as a function of particle diameter, d .

(8) The motion of the particles must be random. Nonrandom particle motion is the main reason for apparent failure or nonapplicability of the technique. Such nonrandom motion can occur through convection currents being present in the system, through particles (too large or dense for the technique) settling during the measurement sequence, or through particles interacting due to chemical reaction or electronic charge. While the system can measure and compensate for this motion, it is preferable to reduce this through accurate temperature measurement and stabilization where possible. If settling/sedimentation occurs in the measurement, other than to a very

minor extent, then the result is almost certainly compromised, as it will reflect a changing and unstable system. If visible settled solid is present at the bottom of a container, then it is very likely that the NTA technique is not recommended. In this case conventional laser light scattering (laser diffraction) is likely to be the preferred technique. If settling can be observed either in the sample container or in the measurement chamber, then it is certain that the original material being measured is not “nano” or is unstable during the measurement time frame.

(9) With respect to size and density, consider the calculations in **Table 1** using Stokes’ Law:

(10) It can be deduced from **Table 1** that if a material is truly < 100 nm, it tends to remain in suspension and exhibits little if any settling tendency. In many situations, for example a gel, the particle density is significantly lower due to incorporation of water into the particle matrix and thus the settling time increased further.

(11) Sometimes it is thought that placing the particles in a material of higher viscosity reduces or even eliminates any settling tendency. This is true, but the Brownian motion is also reduced accordingly and no gain is achieved.

(a) Most dry powder materials are difficult to fully disperse back to a primary size and thus size measurements from diffusion reflect the state of agglomeration of the system rather than to a primary “as produced” size. Hence this Guide assumes that the reader has access to a well dispersed liquid suspension or preparation of nano-size particles for the measurement or that the measurement of the agglomerates is of interest.

(12) Note from **Eq 1** the obvious points that:

(a) As the size of particle increases then the amount of Brownian motion decreases.

(b) As the viscosity of the medium increases then the amount of Brownian motion decreases.

(c) As the temperature is increased then the amount of Brownian motion increases correspondingly.

7.3 Measurement and Analysis of Diffusion Coefficient:

7.3.1 It is necessary to measure the diffusion coefficient to input into **Eq 1** in order to derive a particle size. The diffusion coefficient for each particle in the field of view is determined individually and the resulting sizes are summed to produce the final particle size distribution. This section deals with the measurement of the diffusion coefficient and the objective of providing a particle size distribution from the measured data.

7.3.2 In viewing the scattered light from a group of suspended moving particles over a period of time, each particle moves under Brownian motion. Each particle is identified and tracked for the duration of its presence in the sampling volume, which is defined by the field of view of the camera, the dimensions of the illumination beam, and the focal plane of the magnification optics. The mean squared displacement of each particle is directly measured from the video file images. Dimensions of the displacement (in pixels) are translated into actual displacement through knowledge of the magnification of the system and dimensions of each pixel. This should be calibrated or verified to improve accuracy of the displacement measurement. From the mean squared displacement, the diffusion coefficient is calculated and entered into **Eq 1**.

7.3.3 Each particle must be tracked for a sufficient length of time required to accurately determine the average displacement. The software sizes particles based on analysis of their Brownian motion by identifying and tracking the centers of the spots of light scattered by the individual particles and then measuring the distance moved between frames. Track lengths of only 0.5 seconds duration generate statistically useful data for most particles of interest, assuming a camera frame rate of

TABLE 1 Settling Calculations Based on Stokes’ Law as a Function of Size and Density (T = 298K)

Diameter μm	Diameter nm	ρ (Material) kg/m ³	ρ (Water) kg/m ³	η (Water) 298K, Poise	Time to settle 1 cm (1 × 10 ⁻² m) in water		
					Minutes	Hours	Days
0.01	10	2500	997	0.008955	1815494.39	30258	1261
0.1	100	2500	997	0.008955	18154.94	302.58	12.61
1	1000	2500	997	0.008955	181.55	3.03	0.126
10	10000	2500	997	0.008955	1.82	0.03	0.001
100	100000	2500	997	0.008955	0.02	0.00	0.000
0.01	10	3500	997	0.008955	1089296.64	18154.94	756
0.1	100	3500	997	0.008955	10892.97	181.55	7.56
1	1000	3500	997	0.008955	108.93	1.82	0.076
10	10000	3500	997	0.008955	1.09	0.02	0.001
100	100000	3500	997	0.008955	0.01	0.00	0.000
0.01	10	4200	997	0.008955	851013.00	14183.55	591
0.1	100	4200	997	0.008955	8510.13	141.84	5.91
1	1000	4200	997	0.008955	85.10	1.42	0.059
10	10000	4200	997	0.008955	0.85	0.01	0.001
100	100000	4200	997	0.008955	0.01	0.00	0.000
0.01	10	5500	997	0.008955	605164.80	10086.08	420
0.1	100	5500	997	0.008955	6051.65	100.86	4.20
1	1000	5500	997	0.008955	60.52	1.01	0.042
10	10000	5500	997	0.008955	0.61	0.01	0.000
100	100000	5500	997	0.008955	0.01	0.00	0.000

30 frames/s. The software may automatically select the optimum minimum number of steps on which size estimate is based by continuously monitoring estimated particle size (7).

7.3.4 Particles must be detected by the camera in order to be tracked and the lower limit to particle size which can be analyzed by NTA is determined primarily by particle composition and relative refractive indices. The amount of light scattered by a particle in any given direction is a function of many variables including incident illumination power, wavelength, angle and polarization and particle size, complex refractive index and shape, as well as solvent refractive index (8). Similarly, the amount of light falling on a detector and strength of the resultant signal is dependent, of course, on the efficiency of the collection optics (in particular Numerical Aperture) and the spectral response and sensitivity of the camera. Instrument settings, whether manual or automatically adjusted, will have a direct effect on the system response to scattered light.

7.3.5 The derived particle size distribution from an NTA analysis is naturally a number-based distribution. Given the greater likelihood that smaller particles will diffuse in or out of the field of view due to their more-rapid diffusion, relative to larger particles, during the duration of the measurement, the smaller particles would be over-represented if the final result were reported as a count of particles measured. This effect can be negated by counting each step a particle moves as a single event rather than counting a particle's complete track as a single event. This can be normalized to the total number of frames analyzed. With a known sampling volume, the total concentration is also measured and the size distribution reported as a concentration of particles in each size class (3).

8. Test Method

8.1 Several optical arrangements are used by different manufacturers for NTA based measurement systems. Essentially, the laser source is focused and aligned within the sample cell, while magnifying optics (microscope) equipped with a video camera capture the images. Different approaches to beam focusing and alignment may be used. A generic diagram of an emerging-beam optical arrangement is shown in Fig. 2. A through-cell illumination method is shown in Fig. 3.

8.2 Instrument Components:

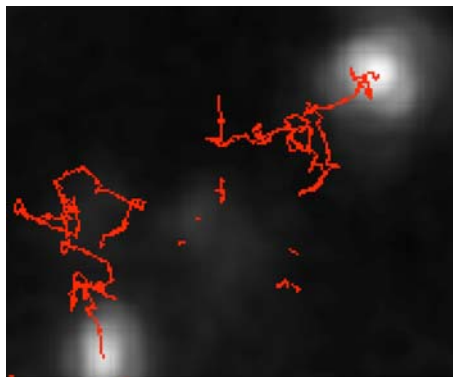


FIG. 1 Tracking of Brownian Motion

8.2.1 Sample analysis module for emerging-beam illumination—which comprises a suitable housing to contain the sample dispersion, the optical element, and the light source, typically a solid-state, single-mode laser diode. The beam is caused to refract at the interface between the liquid sample and the optical element through which it is passed such that it describes a path close to parallel to the glass-sample interface. This angle will be dependent on the dispersion liquid used. This results in a wide, shallow light path that is matched to the depth of field of the optics. It is imperative that only particles within the magnification optics' depth of field be illuminated, otherwise out-of-focus particles will interfere with the measurement by reducing the image signal to noise ratio. The light beam then scatters from the particles in suspension. The optical element may have a metalized surface or be comprised of black glass in order to provide a dark background for high image contrast.

8.2.2 Sample analysis module for through-cell illumination—which comprises a suitable housing to contain the sample dispersion and the light source, typically a solid-state, single-mode laser diode. The beam is focused into the focal point of a microscope lens at a 90° angle. The light beam then scatters from the particles in suspension.

8.2.3 Particles in the beam are visualized by a conventional optical microscope aligned normally to the beam axis and which collects light scattered from each and every particle in the field of view. The total magnification of the system may be quite modest as the system only needs to see the scattered light, not the particles themselves. The particles are seen through the microscope as small points of light moving under Brownian motion. The camera is a digital video camera. Due to the significant variation of the intensity of the image of each particle with size, some adjustment of camera settings (for example gain and shutter speed) is provided for adjustment, either manually or automatically by the software. Care must be taken such that the settings are sufficient to visualize the lower intensity particles while maintaining adequate image quality for the full range of particles in the field of view. This range of settings should be part of any robustness experiments.

8.2.4 A video of typically 30-90 seconds duration is taken of the moving particles by the camera mounted to the optics. The number of particles measured depends on sample concentration, scattering volume, particle size (as this affects diffusion rate), and analysis time. The greater this number, the more representative the analysis will be of the overall samples and the more reproducible the result will be. Wider size distributions will require more total particle measurements, as a statistically significant number of particles need to be measured in each of the size classes of the distribution. Total particle measurements of less than 100 particles may be sufficient for mono-disperse materials while broad distributions may require several hundreds to thousands of particles. The user must define the required reproducibility and the statistical measures for the desired end-purpose. For example, if accurate characterization of the minority parts of the distribution is required, then significantly more particles must be measured than if only the mean/median/mode size is of interest. This total number will depend on polydispersity and shape of the

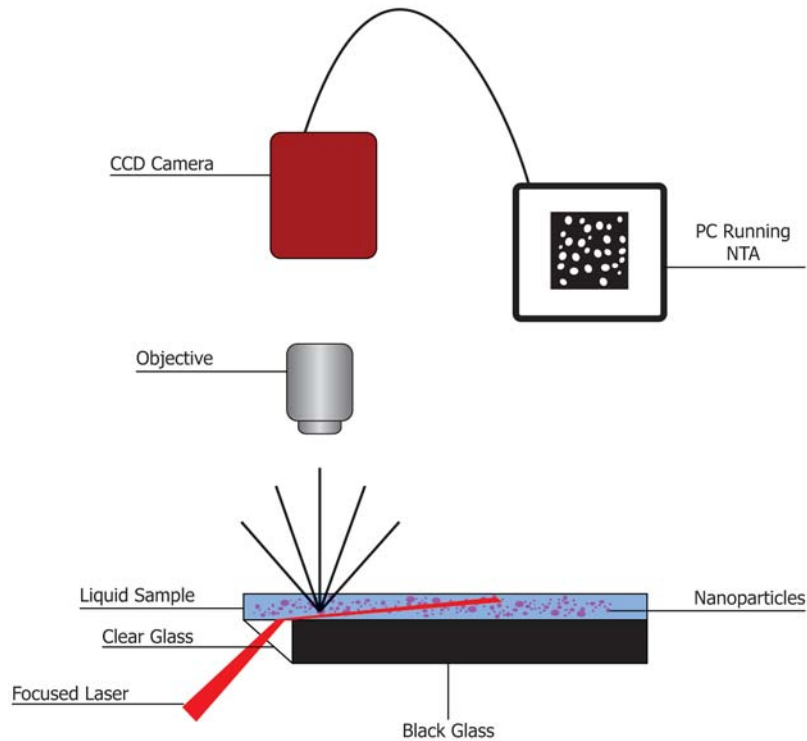
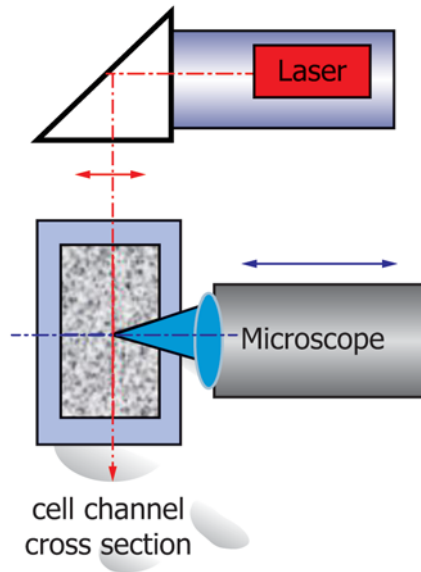


FIG. 2 NTA Measurement System Indicating the Main Components of a Typical Emerging-Beam Illumination Arrangement



NOTE 1—Other optical arrangements are possible including illumination from the side of the sample (orthogonal to the microscope optics) or a generic dark-field microscopy illumination arrangement.

FIG. 3 NTA Measurement System Indicating the Main Components of a Typical Through-Cell Illumination Arrangement

distribution, but in general should be sufficient that the measure of the minority parts of the distribution meet the repeatability and reproducibility requirements defined by the user.

8.2.5 The video is analyzed by the software to locate each particle and track its motion over time. Object-tracking software has many applications and as such there are many approaches to the internal details of the program (9, 10). The specifics of the program and operation will vary by vendor. In general, the video sequence is analyzed, on a frame-by-frame

basis, so that each particle is identified and located automatically and its movement tracked over time. Various parameters may be adjusted either manually or automatically to optimize conditions for particle tracking and rejection of optical noise. These parameters may include thresholds for particle identification, gain, brightness, or blur, thus allowing the user to optimize the image for a particular sample type or can be selected by the program. The video sequence may also be adjusted in terms of image smoothing, background subtraction,

setting of thresholds, or removal of blurring to allow particles of interest to be tracked without interference from stray flare or diffraction patterns which can occasionally occur with non-optimum sample types. As the details of different manufacturer's programs and even different versions of these programs may vary significantly, the user is referred to the instruction manuals for the particular instrument for further details.

8.2.6 A particle size distribution result is then produced either in frequency or histogram format. The user needs to check the derived distribution for reasonableness and repeat consecutive measurements are advised to ascertain the stability of the final answer (dependent both on the stability of the material and the heterogeneity). Replicate samples allow the sample-to-sample variation to be ascertained. A robustness experiment should be performed to establish acceptable limits to these variations.

8.3 *Sampling:*

8.3.1 Preparation of a representative sample in a stable and dispersed state is vital to an accurate and meaningful analysis. To obtain the material in this state is not a trivial matter. ISO 14488 (Particulate materials - sampling and sample splitting for the determination of particulate properties) deals extensively with this subject. Useful guides are to be found in the NIST Practice Guide Special Publication 960-1 Particle Size Characterization (11), the first chapter of T. Allen's Particle Size Measurement (12) as well as a large number of ISO standards such as ISO 13320 and ISO 13321 (particle size analysis standards for techniques which have similar sampling and preparation requirements), and ASTM standards, only a limited number being relevant to nano systems (for example, Practice C322).

8.3.2 The examination of the time trend (size with time, consecutive measurements, input energy—sonication) for the particle size distribution in a repeatability study is vital to ensure stability and confidence in the final reported results. Sample-to-sample reproducibility can be assessed by the taking of replica aliquots or subsamples from the same bulk lot.

8.3.3 To take a representative aliquot of material, the material needs to be moving when the sample is extracted. With a slurry or suspension, sampling is normally carried out by pipetting the required amount of sample from a stirred beaker containing the primary material. If the sample has settled or is settling, and material is extracted only from the supernatant, it is clear that a smaller answer than the bulk material is obtained. Slurry sampling is notoriously difficult to carry out correctly and the use of a Burt sampler (the slurry equivalent of a spinning riffler for powders) is recommended.

8.3.4 The wider the particle size distribution, the more potential problems may be encountered throughout the sampling and measurement especially if a "distribution" is sought. Wider particle size distributions are subject to greater possibilities of segregation or settling which complicate the sampling and measurement issues although again these are likely to be minimal for truly sub-100 nm systems.

8.3.5 When only tiny total amounts of sample are available, then subsampling is not likely to be statistically admissible for the desired degree of accuracy and the entire amount may be required for the analysis. Note that similar constraints apply to

any technique where relatively few particles (= little mass, picograms or nanograms) are sampled.

8.4 *Dispersion:*

8.4.1 Before the experiment is started, it is important to ascertain the purpose of the measurement. Unfortunately, the tendency is often to seek the "smallest possible answer" without regard to whether the real problem is (say) a plant rheology or filtration problem—controlled by the bulk particulate size—or whether information is required relating to the primary particle size (dissolution, reactivity, take up of ions from solution, and so forth). The real purpose of the test and the value of a primary particle size versus a measure of agglomerated state and what that means to process control or the materials application in conditions where it is actually used should be carefully considered. These needs and objectives dictate the amount of energy needed for dispersion prior to the analysis.

8.4.2 The use to which the end results are put is also crucial especially if economic values are at stake (for example, batch control, incoming goods check). We need to consider the implications of what an 'out-of-specification' result will mean in monetary terms. Note that this statement implies that we have a specification in place and test against this specification.

8.4.3 Dispersion for small systems often involves the use of a large amount of input (sonication) energy, especially if the material is in a powdered state to start. Some materials (especially biological or those of high aspect ratio) are not likely to withstand huge amounts of energy input. Given that we are to be measuring the Brownian motion of the particulate system, then co-joined (aggregated or agglomerated or both) groups of primary particles behave as a single larger particle. This fact needs to be borne in mind with any technique that is not able to differentiate between a single (larger) particle and an aggregate or agglomerate of smaller particles.

8.4.4 Note also the discussion of scattering intensity on size from 7.2.1.2. The technique is sensitive to any larger or agglomerated particles in the system. While the technique is able to measure a wide range of sizes simultaneously, it is often desirable to remove (by filtration or centrifugation) even small amounts of any larger material present, recognizing the fact that this is altering the particle size distribution, in particular, on a mass basis. It is to be noted that as the analysis measures all particles present in the sample and identification of particles from the background (diluent) is not possible, a background subtraction 'count' of the solvent is not possible or feasible. Thus cleanliness of the background solvent is essential and filtration is usual. A measurement of particles in the solvent must be carried out for quantification of contamination from that source.

8.5 *Particle Concentration:*

8.5.1 A certain concentration of particles is required in the system in order that a statistically significant number of particles can be analyzed. This concentration is dependent on the measurement volume, the length of time over which the sample is analyzed and the size of the particles present.

However, note that very low concentrations of poorly scattering materials (for example, liposomes) are not likely to generate adequate signal for reasonable measurement, in a number of situations.

8.5.2 Commensurate with the requirements of sufficient particle concentration are those ensuring that the concentration is not so high as to cause particle coincidence. Subject to the specifics of measurement volume, the size of the particles present, and the capabilities of the software, when particles diffuse to within a certain distance of an adjacent particle, tracking is ceased due to the inability of the software to individually identify any one particle, eliminating the possibility of analyzing particle trajectories that cross each other. While this is not likely to prevent the taking of a measurement (unless the solution is so concentrated that the camera is saturated), broken tracks will prevent particles from being measured. Longer analysis times may be required in this case or some effect on results may be seen. The total number of completed tracks or the ratio of completed tracks to total tracks may provide an indication of this effect. In areas where it is difficult or impossible to adjust the dilution stage and measurements are still desired, note that the particle concentration needs to be controlled and recorded.

8.6 *The Measurement:*

8.6.1 In order that random Brownian motion is to be measured accurately, the system needs to be in complete thermal stability (optimally $\pm 0.10^\circ\text{C}$). While the technique may be able to compensate for thermal or convection currents in the measurement cell or cuvette, optimum results are obtained with a completely quiescent system. Follow the manufacturer's guidelines in terms of warm-up and sample equilibration times. Select the conditions for the measurement, ensuring that the correct parameters are set within the instrument and that the sample is described correctly in the documentation. Parameters that need setting or recording are likely to include:

- (1) The wavelength of the laser utilized in the experiments.
- (2) The duration of the experiment and any sub-experiments carried out. It is possible to take measurements as a consecutive set of replicate experiments of identical duration or at fixed time intervals. In this way any changes with time (agglomeration, deagglomeration, dissolution, settling) can be followed.
- (3) The set or measured temperature that the measurements were taken at.
- (4) The viscosity appropriate to the liquid used in the study.
- (5) The name of the operator.

8.6.1.1 It may be useful to take repeated consecutive measurements to assess the stability of the system. In this manner, a system undergoing settling or other untoward effects can be seen. The individual measurement duration needs to correspond with manufacturer's recommendations. Extended measurement times are likely to be needed with larger particles (less Brownian motion) or wider distributions of sizes (fewer counts for each size class).

8.6.2 Examine the measurement chamber before and after measurement for evidence of settling.

8.6.3 At the end of the measurement, a particle size distribution is produced in graphical and tabular form. In some cases it is possible (within the constraints of the system and material) to follow kinetic events such as aggregation, dissolution, or polymerization through repeat measurements at appropriate time intervals.

8.6.4 The main calculated/derived parameters that are most useful are the following:

- (1) Mean
- (2) Mode
- (3) Percentile values—commonly D10, D50, D90
- (4) Concentration

8.6.5 Other information can be displayed, but the assumptions in obtaining this derived information need to be carefully understood and the implications of reporting this information also carefully evaluated. Since the reported concentration distribution is directly measured, volume based distributions derived from nanoparticle tracking analysis measurements can reasonably be used for comparison purposes or for estimating the relative amounts of multimodal (multiple size peaks) samples, with the understanding of limited statistics may lead to significant variability in these alternate distribution forms.

8.7 *Interpretation and Comparison with Other Techniques:*

8.7.1 The result that is derived from NTA is a number-based size distribution that corresponds to how the particle moves within the liquid (hydrodynamic diameter). Non-spherical particles move in different ways along different axes, but NTA measures the total movement over the entire observation time, so a sphere-equivalent, hydrodynamic diameter is derived on the basis of this total diffusion. The derived particle size distribution is based on a summation of all measured particles. If conditions are inappropriate for measurement of a given size (or intensity) range of particle, those particles will not be included in the final result.

8.7.2 PCS is a common nanoparticle sizing technique, as defined by Guide E2490 and ISO 22412. The mean result that is derived from PCS is an intensity-weighted mean (commonly known as the z -average) that corresponds to how the particle moves within the liquid and interacts with light. The z -average is weighted towards the higher end of the particle size distribution, as the intensity is proportional to d^6 (or V^2 or MW^2). In simple terms if we have one 100 nm particle it is equivalent in intensity weighting terms to 10^{12} (1 trillion) 1 nm particles. This differs from say a classic laser diffraction experiment where the volume weighting would have 10^6 (or 1 million) 1 nm particles equivalent to the single 100 nm particle, which is correct in mass terms (for constant density of particle). Thus, a polydisperse sample may result in significantly different results from NTA. See Filipe (13) for an extensive comparison of the two techniques. Note that with any form of microscopy the 1 nm particle has equal equivalence (or weighting) to the 100 nm particle, even though they differ greatly in mass (and therefore weighting in the final reported size distributions). Thus, conversions to other distributions to which the reader is likely to be familiar (for example, volume or number), are fraught with dangers—see 8.8.

8.7.3 The accuracy of the presented distribution is related to the limits of the method discussed previously. Thus the user

will benefit from having prior information as to the nature of the sample in order to conclude whether a reported answer is reasonable or not and comparison with other techniques is beneficial to generate confidence in a set of results. In all cases, some form of direct visualization of a statistically valid number of particles, is desirable. For the nano-size range (< 100 nm) only electron microscopy or atomic force microscopy would be applicable. Note that it is often very difficult to get images for a number of biological systems, for example, micelles, proteins or other materials such as emulsions which usually require specialized techniques (for example, freeze-fracture).

8.7.4 For narrow distributions (RSD <10 %) there is little inherent problem in the analysis of the raw video files to determine particle size information. For wider distributions (RSD >10 %), care must be exercised to ensure that the appropriate methods and conditions are selected to ensure reliable results. Analysis time, image capture settings, and sample concentration are variables that need to be considered.

8.8 Conversion of the Concentration Distribution to Other Particle Size Distributions:

8.8.1 With any technique, the most reliable results are those reported in that technique's natural format. Conversion to other distribution bases may be mathematically valid, but the assumptions in obtaining this derived information need to be carefully understood and the implications of reporting this information also carefully evaluated. Small changes in collected data can give rise to enormous changes in derived result and as such treat any derived result with caution and skepticism. To convert from concentration to volume distribution would involve the manipulation of perfect noise-free experimental data. The wider the initial distribution the more serious are the potential errors in the conversion.

8.8.2 Notwithstanding the above caveats and cautions, conversion to a volume-weighted distribution can often provide an indication of the relative importance (prominence) of two or more reported peaks. A common situation is to see an apparently dominant small-size peak virtually disappearing and a low-intensity larger-sized peak becoming the primary mode after conversion to volume weighting.

9. Report

9.1 See Practice E1617.

9.2 As a minimum the following need reporting in addition to graphical and tabular information:

9.2.1 The instrument type and manufacturer and serial number. Version of software employed.

9.2.2 Date and results of the last verification. Details of the traceability of the standards employed.

9.2.3 Date of measurement together with analyst's name and affiliation.

9.2.4 Details of the sample including chemical composition. Shape information if obtained by electron microscopy or atomic force microscopy is helpful.

9.2.5 Details of the dispersion conditions (concentration of material, liquid used, ultrasound time, frequency and power, surfactants and stabilizing agents, if used, and their concentration) and evidence that full dispersion or primary particle size has been reached.

9.2.6 Measurement conditions—time of measurement, wavelength of laser, stabilization period prior to measurement, temperature. Number of measurements.

9.2.7 Minimum of 3 replicate consecutive measurements. This demonstrates the stability of the material (especially) and the instrument during the duration of the measurements. The use of a relative standard deviation (RSD) or coefficient of variation (CV) is highly recommended.

9.2.8 Minimum of 3 separate aliquots/samples from the same bulk lot measured under identical conditions and with each having a minimum of 3 replicate consecutive measurements as above. This demonstrates the homogeneity or heterogeneity of the bulk lot and the aliquot-to-aliquot reproducibility.

9.2.9 Measurement Conditions:

9.2.9.1 Viscosity of medium (for Stokes-Einstein equation). Note that this is the viscosity that the particle(s) experience in their interaction with the medium. It is equivalent to a viscosity at zero stress.

9.2.9.2 Temperature of the sample.

9.2.9.3 Image capture and analysis settings. Note: Include any relevant settings as discussed in Section 8.

10. Keywords

10.1 DLS; dynamic light scattering; nano; nanoparticle tracking analysis; NTA; PCS; photon correlation spectroscopy

REFERENCES

- (1) International Conference on Harmonisation, *Guideline on the Validation of Analytical Procedures: Methodology*, Published in the Federal Register, 19 May 1997, Vol 62, No. 96, p. 27463–7.
- (2) Hunter, R.J., *Zeta Potential in Colloid Science*, Academic Press, New York, 1981.
- (3) Malloy, A., and Carr, R., "NanoParticle Tracking Analysis—The Halo System," *Particle & Particle Systems Characterization* (Special Issue: Particulate Systems Analysis), Vol 23, No. 2, 2006, pp. 197–204.
- (4) Pecora, R., (Ed.) *Dynamic Light Scattering, Applications of Photon Correlation Spectroscopy*, Plenum Press, New York, 1985.
- (5) Chandrasekhar, S., "Brownian Motion, Dynamical Friction, and Stellar Dynamics," *Reviews of Modern Physics*, Vol. 21, Num. 3, July 1949, pp. 383–388.
- (6) Einstein, A., and Fürth, R., translated by Cowper, A., "On the Movement of Small Particles Suspended in a Stationary Liquid Demanded by the Molecular-Kinetic Theory of Heat," *Investigations on the Theory of the Brownian Motion*, Dover Publications, New York, 1926.
- (7) Saveyn, H., De Baets, B., Thas, O., Hole, P., Smith, J., and Van der

Meeren, P., “Accurate particle size distribution determination by Nanoparticle Tracking Analysis based on 2-D Brownian dynamics simulation,” *Journal of Colloid and Interface Science*, 352, December 2010, pp. 593–600.

- (8) Provencher, S.M., “Low-bias Macroscopic Analysis of Polydispersity,” *Laser Light Scattering in Biochemistry*, Royal Society of Chemistry (UK), Special Publication No. 99.
- (9) Crocker, J. and Grier, “Methods of Digital Video Microscopy for Colloidal Studies,” *Journal of Colloid and Interface Science*, 179, 1996, pp. 298–310.
- (10) Kalaizidis, Y., “Multiple objects tracking in fluorescence microscopy,” *Journal of Mathematical Biology*, Vol 58, 2009, pp. 57–80.

- (11) Jillavenkatesa, A., Lin-Sien H. Lum, and Dapkunas S., *Particle Size Characterization*, NIST Recommended Practice Guide, Special Publication 960-1, National Institute of Standards and Technology, January 2001.

- (12) Allen, T., *Particle Size Measurement*, 5th Edition, Volume 1, Chapter 1, Chapman & Hall, London, 1993.

- (13) Filipe, V., Hawe, A., and Jiskoot, W., “Critical Evaluation of Nanoparticle Tracking Analysis (NTA) by NanoSight for the Measurement of Nanoparticles and Protein Aggregates,” *Pharmaceutical Research*, (2010) Vol. 27, No. 5, pp. 796-810.

ASTM International takes no position respecting the validity of any patent rights asserted in connection with any item mentioned in this standard. Users of this standard are expressly advised that determination of the validity of any such patent rights, and the risk of infringement of such rights, are entirely their own responsibility.

This standard is subject to revision at any time by the responsible technical committee and must be reviewed every five years and if not revised, either reapproved or withdrawn. Your comments are invited either for revision of this standard or for additional standards and should be addressed to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee, which you may attend. If you feel that your comments have not received a fair hearing you should make your views known to the ASTM Committee on Standards, at the address shown below.

This standard is copyrighted by ASTM International, 100 Barr Harbor Drive, PO Box C700, West Conshohocken, PA 19428-2959, United States. Individual reprints (single or multiple copies) of this standard may be obtained by contacting ASTM at the above address or at 610-832-9585 (phone), 610-832-9555 (fax), or service@astm.org (e-mail); or through the ASTM website (www.astm.org). Permission rights to photocopy the standard may also be secured from the Copyright Clearance Center, 222 Rosewood Drive, Danvers, MA 01923, Tel: (978) 646-2600; http://www.copyright.com/